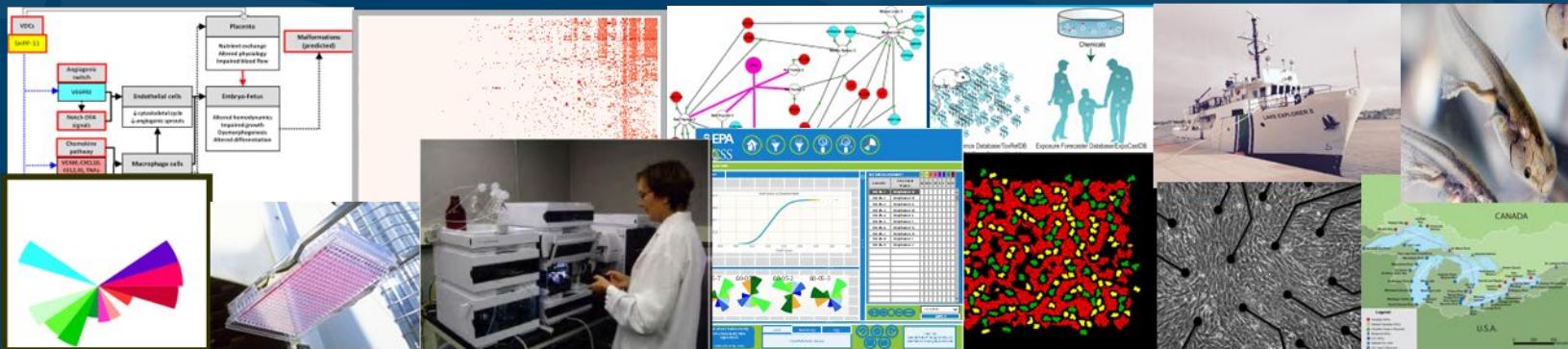


# NAMs Work Plan Progress and Day 1 Overview



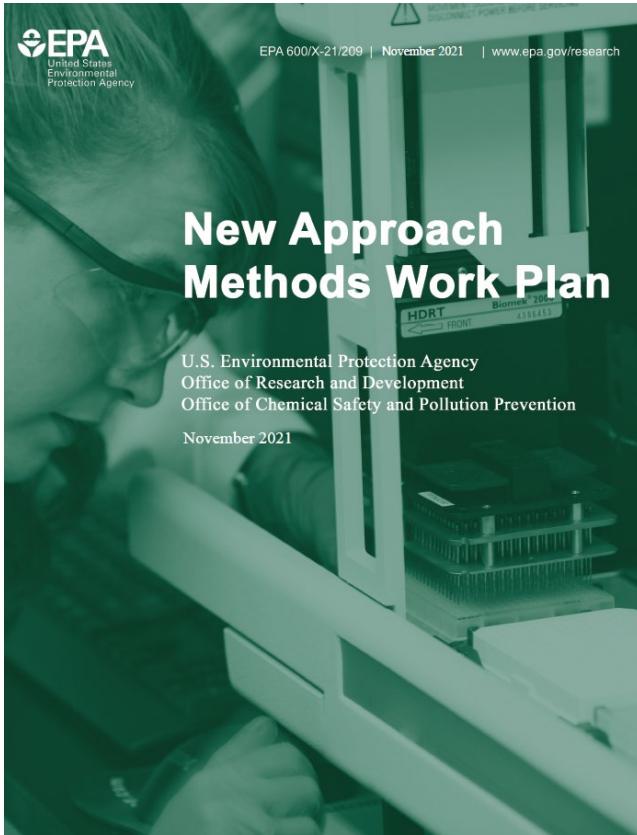
**Fourth EPA NAMs Conference**

**November 5 - 6, 2024**

**Rusty Thomas**  
**Director**  
**Center for Computational Toxicology and Exposure**

The views expressed in this presentation are those of the presenter and do not necessarily reflect the views or policies of the U.S. EPA

# NAMs Work Plan Identified Objectives, Strategies and Deliverables for Applying NAMs



- Five objectives for reducing animal testing and research while ensuring that Agency decisions remain fully protective of human health and the environment
  - Develop Baselines and Metrics
  - Evaluate Regulatory Flexibility
  - Establish Scientific Confidence and Demonstrate Application
  - Develop NAMs to Address Information Gaps
  - Engage and Communicate with Stakeholders
- Updated NAM Work Plan released in December 2021
  - Expansion of the species covered in the work plan to include all vertebrate animals to be consistent with TSCA.
  - Modified deliverable timelines that reflect the expansion of covered species and incorporate feedback received over the preceding years.
  - New case studies for building confidence and demonstrating application of NAMs.
  - A pilot study to develop NAMs training courses and materials.



# Status of NAMs Work Plan Deliverables

Milestones/Deliverables	Projected Dates
<b>Evaluate Regulatory Flexibility for Accommodating the Use of NAMs</b> EPA report on a review of existing statutes, programmatic regulations, policies, and guidance that relate to vertebrate animal testing and the implementation and use of appropriate NAMs for regulatory purposes.	2022 
<b>Develop Baselines and Metrics for Assessing Progress</b> Progress and summary metrics on reducing vertebrate animal testing requests and use.	Annually starting in Q4 2022 



# Status of NAMs Work Plan Deliverables

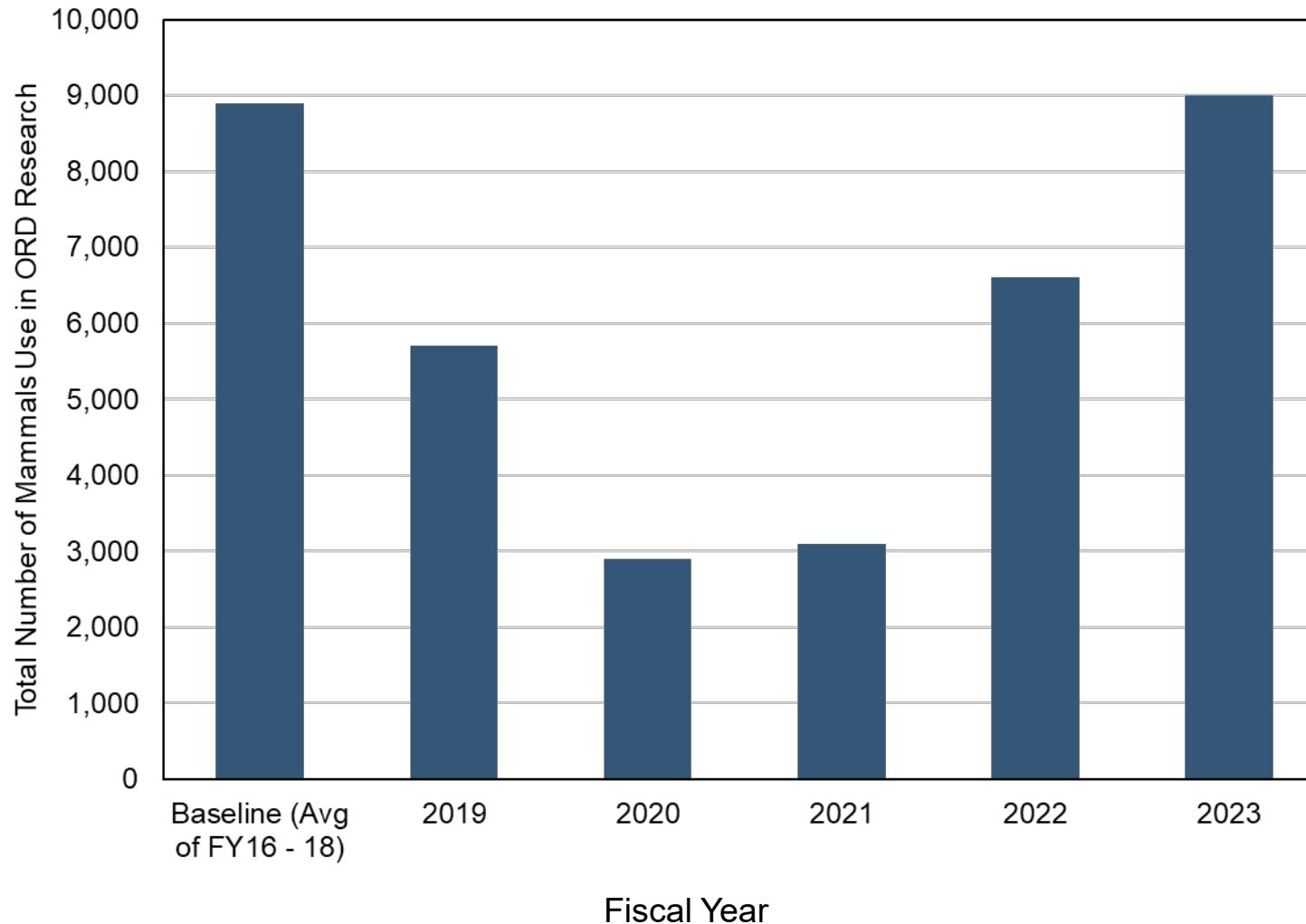
Milestones/Deliverables	Projected Dates
<b>Establish Scientific Confidence in NAMs and Demonstrate Application to Regulatory Decisions</b>	
U.S. National Academies of Sciences, Engineering, and Medicine study that evaluates the variability and relevance of existing mammalian toxicity tests and reviews frameworks for validation and establishing scientific confidence in testing methods.	2023 
A scientific confidence framework to evaluate the quality, reliability, and relevance of NAMs.	Q4 2024
An initial set of reporting templates which may be used by EPA and stakeholders that capture the range of specific NAMs used for Agency decisions.	Q4 2024
Case studies for evaluating application to risk assessment and demonstrating protection of human health and the environment.	Ongoing 



# Status of NAMs Work Plan Deliverables

Milestones/Deliverables	Projected Dates
<b>Develop NAMs to Address Scientific Challenges and Fill Important Information Gaps</b>	
EPA Strategic Research Action Plans outlining research products to develop and apply NAMs.	Q1 2023 
Encourage development of NAMs through mechanisms such as the STAR program and facilitate partnerships with organizations focused on establishing scientific confidence in alternative methods.	Ongoing 
<b>Engage and Communicate with Stakeholders</b>	
EPA website to house information about NAM efforts and progress being upon release of the work plan.	2020 
Public webinars and, where appropriate, peer-review on deliverables from this work plan.	Ongoing 
Complete NAMs pilot training program in the fourth quarter (Q4) of 2023 and provide regular scientific exchanges and progress updates through Agency sponsored and partner organized events.	Q4 2023 and Ongoing 

**Milestone/Deliverable:** Progress and summary metrics on reducing vertebrate animal testing requests and use. (FY22+).

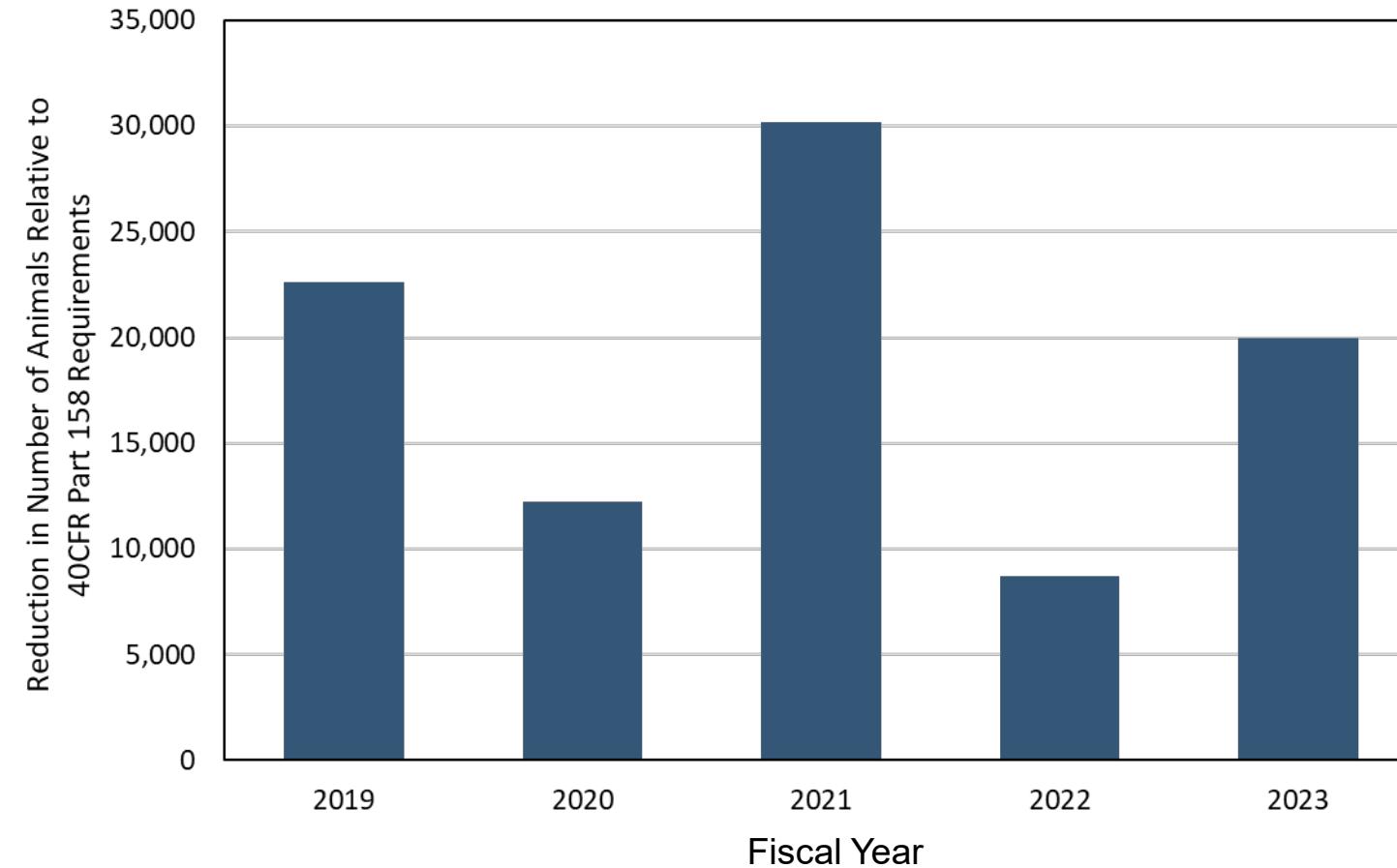


- The numbers in FY19 – 23 include those mammals used in contract research activities.
- Baseline numbers (FY16 – 18) do not include mammals used in contract research activities due to a lack of tracking at that time.
- The numbers in FY19 – 21 are likely reduced due to impacts of the ORD reorganization/lab remodeling and pandemic (FY20 – 21).
- A system for estimating vertebrate animal use in research (includes fish and amphibians in addition to mammals) has been established.
- A baseline for vertebrate animal use will be established after data has been collected over multiple years.

# EPA FY19 – FY23 Animal Reduction Metrics for OPP

Develop  
Baselines and  
Metrics

**Milestone/Deliverable:** Progress and summary metrics on reducing vertebrate animal testing requests and use. (FY22+).



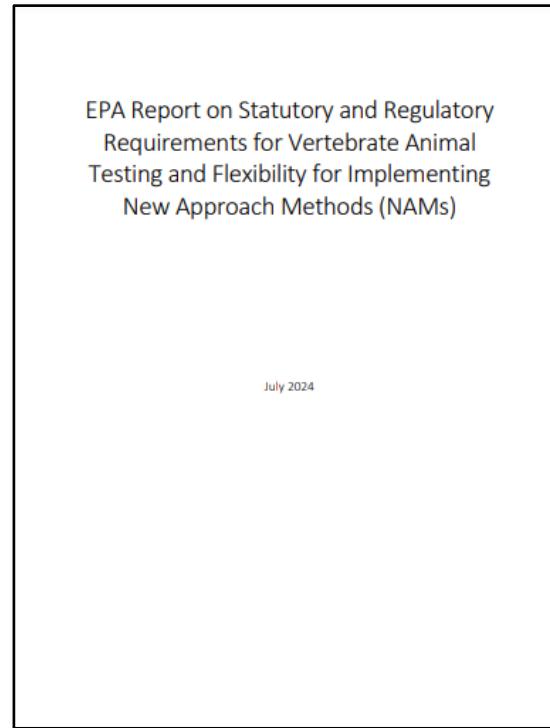
- The reduction in the number of animals were due to Hazard and Science Policy Council (HASPOC), Chemistry and Acute Toxicology Science Advisory Council (CATSAC), and Acute Dermal waivers granted under the updated waiver policies (2016/2020).
- Beginning in 2023, OPP expanded tracking of data waivers granted for the acute “6-pack”.
- The total number waivers granted from FY19 – 23 were:
  - HASPOC - 235
  - CATSAC - 62
  - Acute Dermal - 229
- The number of NAM-related endpoint data submissions from FY19 – 23 were:
  - Eye Irritation - 81
  - Skin Irritation - 57
  - Skin Sensitization - 25



# Review of Statutes and Regulations for Flexibility in Incorporating NAMs

Evaluate  
Regulatory  
Flexibility

**Milestone/Deliverable:** EPA report on a review of existing statutes, programmatic regulations, policies, and guidance that relate to vertebrate animal testing and the implementation and use of appropriate NAMs for regulatory purposes.



[https://www.epa.gov/system/files/documents/2024-09/epa-regulatory-review-report\\_final\\_508\\_0.pdf](https://www.epa.gov/system/files/documents/2024-09/epa-regulatory-review-report_final_508_0.pdf)

- Topics covered in report:
  - Overview of main environmental statutes and language on testing requirements.
  - Regulatory requirements for vertebrate animal testing
  - Research to support regulatory use of NAMs
  - Current use of NAMs in decision making
  - Barriers to implementation and use of NAMs
- Statutes are written broadly in most cases and do not generally preclude the use of scientific information or data from NAMs.
- Some regulations require a minimum set of vertebrate animal testing for decision-making.
- Program offices will need to identify the regulations that require revisions to incorporate data from NAMs when feasible and scientifically justified.



# EPA STAR Grants

**Milestone/Deliverable:** Encourage development of NAMs through mechanisms such as the STAR program and facilitate partnerships with organizations focused on establishing scientific confidence in alternative methods. (Ongoing).

United States Environmental Protection Agency

Environmental Topics ▾ Laws & Regulations ▾ Report a Violation ▾ About EPA ▾

Research Grants

CONTACT US

## Safer Chemicals Research Grants

EPA funds safer chemical research grants supporting the development of innovative science to support safer, more sustainable use of chemicals in consumer products and chemicals used for other purposes such as pesticides. Using safer, more sustainable chemicals will help to better protect human and environmental health, including sensitive populations like children, elderly and endangered species.

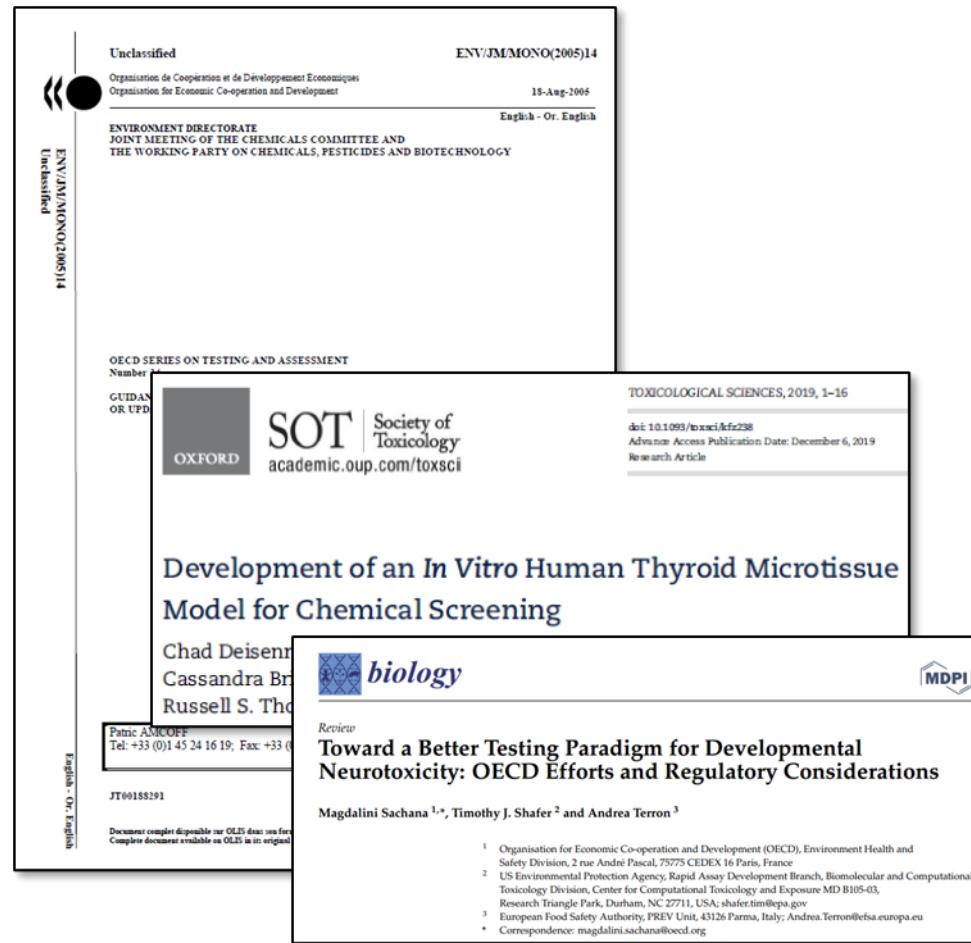
All forms necessary for completing an application are referenced in the RFA and available to download online. [Funding Opportunities: How to Apply and Required Forms](#)

<https://www.epa.gov/research-grants/star>

- EPA STAR grants on *Advancing Actionable Alternatives to Vertebrate Animal Testing for Chemical Safety Testing* (2019-22/25)
  - Awarded ~\$4.2 million to 5 universities
  - Vanderbilt University, University of California Riverside, Louisiana State University, Oregon State University, Johns Hopkins University
- EPA STAR grants on *Advancing Toxicokinetics for Efficient and Robust Chemical Evaluations* (2020 – 2023/2025)
  - Awarded ~\$4 million to 5 institutions
  - Purdue University, Woods Hole Oceanographic Institution, Vanderbilt University, Texas A&M, and University of Nevada Reno
- EPA STAR grants on *Development of Innovative Approaches to Assess the Toxicity of Chemical Mixtures* (2022-25)
  - Awarded ~ \$7.7 to 11 institutions
  - [www.epa.gov/research-grants/development-innovative-approaches-assess-toxicity-chemical-mixtures-research-grants](https://www.epa.gov/research-grants/development-innovative-approaches-assess-toxicity-chemical-mixtures-research-grants)
- EPA STAR grants on *Advancing Sustainable Chemistry*. (Awardees Coming Soon)

# Partnerships with External Organizations Focused on Scientific Confidence

**Milestone/Deliverable:** Encourage development of NAMs through mechanisms such as the STAR program and facilitate partnerships with organizations focused on establishing scientific confidence in alternative methods. (Ongoing).



- Partnering with 4 external organizations on an inter-laboratory prevalidation study of a human thyroid microtissue assay.
- Partnering with 5 external organizations on the development and validation of 17 assays for developmental neurotoxicity.
- Co-leading OECD activity to update Guidance Document 34.
- Collaborating with NICEATM to catalog characteristics of OECD Test Guideline validation studies.
- Participating in the NIH Complement-ARIE program.
- Co-authored ICCVAM report on the validation, qualification, and regulatory acceptance on NAMs in 2024.

**Milestone/Deliverable:** Case studies for evaluating application to risk assessment and demonstrating protection of human health and the environment (Ongoing).



**APCRA**

ACCELERATING THE PACE OF  
CHEMICAL RISK ASSESSMENT



**OECD**



**ECHA**



- Co-leading APCRA case study on using bioactivity for screening level risk assessment.
- Leading APCRA case study on application of *in vitro* toxicokinetics to regulatory decisions.
- Co-leading APCRA case study on incorporating NAMs into species sensitivity distributions for ecological risk assessment.
- Leading international APCRA case study on developing quantitative structure use relationship models for predicting chemical functional use.
- Leading international APCRA case study on evaluating quantitative concordance between human and rodent toxicity data.
- Participating in OECD case study on systemic toxicity.



# EPA NAM Pilot Training Program and Regular Scientific Exchanges and Progress Updates

Engage and Communicate with Stakeholders

**Deliverables:** Completed training pilot and used lessons learned for trainings going forward. Provide regular scientific exchanges and progress updates through Agency sponsored and partner organized events (ongoing – 2026). TSCA New Chemicals Outreach due 2026.

- Public NAMs training website serves as a resource for EPA NAMs training materials and recordings
- Available Resource Hubs & Tool User Guides ([comptox.epa.gov](https://comptox.epa.gov)): ChemExpo Knowledgebase, Cheminformatics Modules, CompTox Chemicals Dashboard, Generalized Read-Across, ECOTOX Knowledgebase, Sequence Alignment to Predict Across-Species Susceptibility
- Available Tool Tips Videos: CompTox Chemicals Dashboard (6), ECOTOX (6)
- NAMs Update email bulletin to share progress and updates, email [NAM@epa.gov](mailto:NAM@epa.gov) to join!



<https://www.epa.gov/chemical-research/new-approach-methods-nams-training>

# NAMs Tools Trainings

Training	Date	Attendees
ECOTOX	May 2022	321
CompTox Chemicals Dashboard	Oct 2022	554
ECOTOX	Feb 2023	575
GenRA	May 2023	730
httR R Package	Nov 2023	521
In-Person multi-tool training	April 2024	88
AOP-Wiki	Sep 2024	298

## Upcoming:

- March 2025: CompTox Tools training at Society of Toxicology conference in a Satellite Meeting
- April 2025: Interactive training on Sequence Alignment to Predict Across-Species Susceptibility (SeqAPASS) tool
- Late 2025: In-person high-throughput toxicokinetics

# NAM Conference Topic Survey



- More than 40 submissions from 30 individuals/organizations
- Many submissions included multiple topics
- **All** of the topics on the agenda are from YOU!!!
  - Validation
  - Exposure
  - Toxicokinetics/IVIVE
  - Omics
- Thank you for your input!



# Overview of Day 1

<b>Validation Update</b>		
1:30 – 2:00 pm	ICCVAM Validation Report and NICETAM Activities	Nicole Kleinstreuer (NICEATM)
2:00 – 2:30 pm	Building Confidence in NAMs via Validation Standard Setting in a Revised OECD GD 34	Alison Harrill (EPA)
<b>Exposure NAMs</b>		
2:30 – 3:00 pm	Development and Application of Exposure NAMs in EPA's ExpoCast Project	Kristin Isaacs (EPA)
3:00 – 3:30 pm	Break	
3:30 – 4:00 pm	Integrating Geospatial Exposure Models with NAMs to Evaluate Health Risks from Environmental Chemicals	Kyle Messier (NIEHS)
4:00 – 4:30 pm	Leveraging Non-Targeted High-Resolution Mass Spectrometry to Reveal the Complete PFAS Fingerprint in Maryland	Sin Urban (Maryland Department of Health)
4:30 – 5:00 pm	Human and Environmental Exposure Framework for Biosolids	Carsten Prasse (Johns Hopkins University)
5:00 – 5:10 pm	Session Wrap Up	Annette Guiseppi-Elie (EPA)



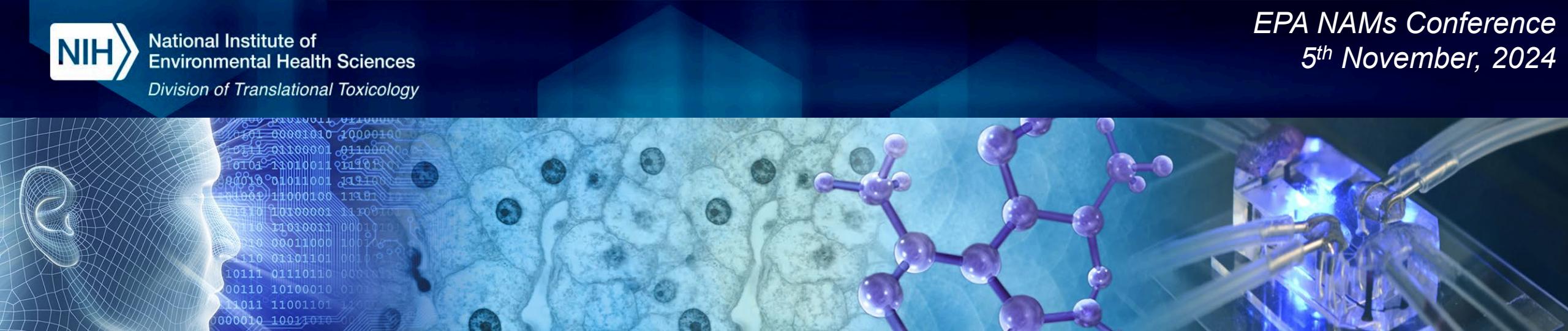
# Acknowledgements

## NAM Work Plan Implementation Team

Christina Baghdikian  
Kathie Dionisio  
Annette Guiseppi-Elie  
Alison Harrill  
Charles Kovatch  
Monica Linnenbrink  
Anna Lowit  
Jeff Morris  
Monique Perron  
Keith Salazar  
Nisha Sipes  
Russell Thomas  
Krystle Yozzo

## Conference Organization

Christina Baghdikian  
Esra Mutlu  
Scarlett VanDyke  
Sammy Hanf  
Carolyn Holmes  
ICF

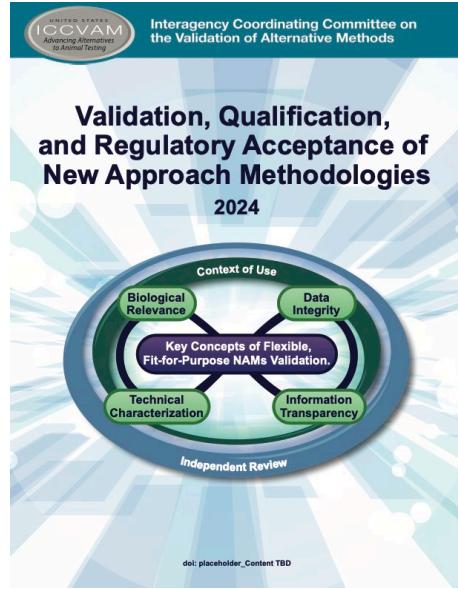
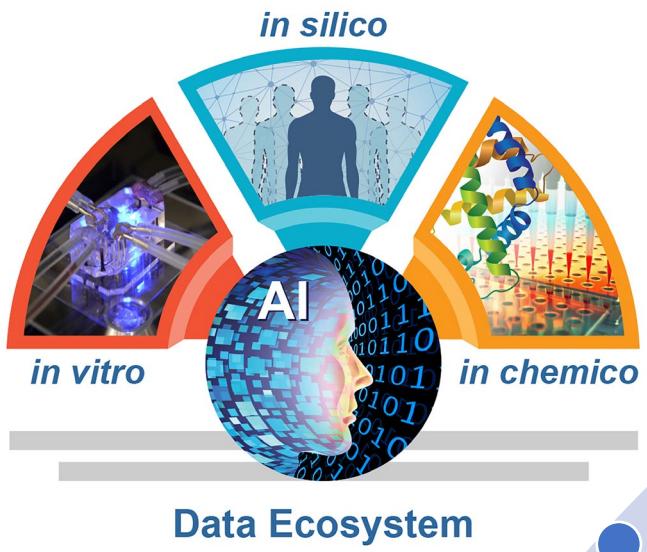


# ICCVAM Validation Report and NICEATM Activities

**Nicole C. Kleinstreuer, PhD**

**Director, NTP Interagency Center for the Evaluation of Alternative  
Toxicological Methods**

**Executive Director, Interagency Coordinating Committee for the  
Validation of Alternative Methods**



etc.

Validation/  
Qualification

Adoption &  
Implementation

Biomedical  
Research

**The NAMs Confidence Continuum:**  
from *Biomedical Research* to  
*Validation/Qualification* to  
*Adoption & Implementation*

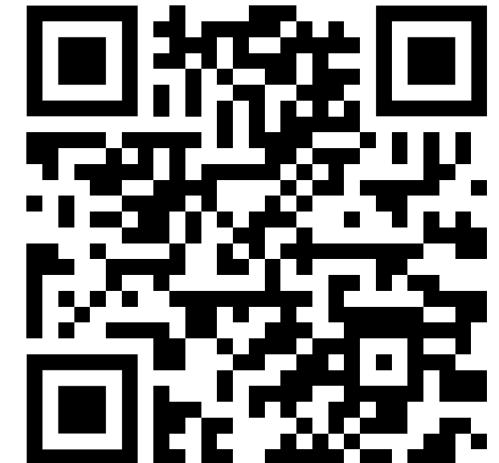
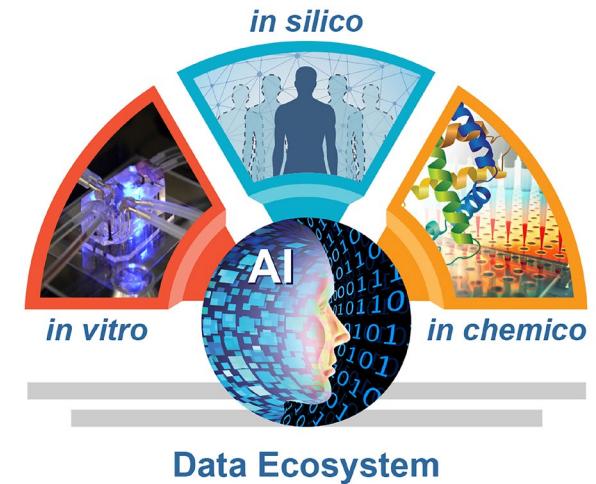
# Complement-ARIE: Complement Animal Research in Experimentation

**Purpose:** To catalyze the development, standardization, validation and use of **human-based new approach**

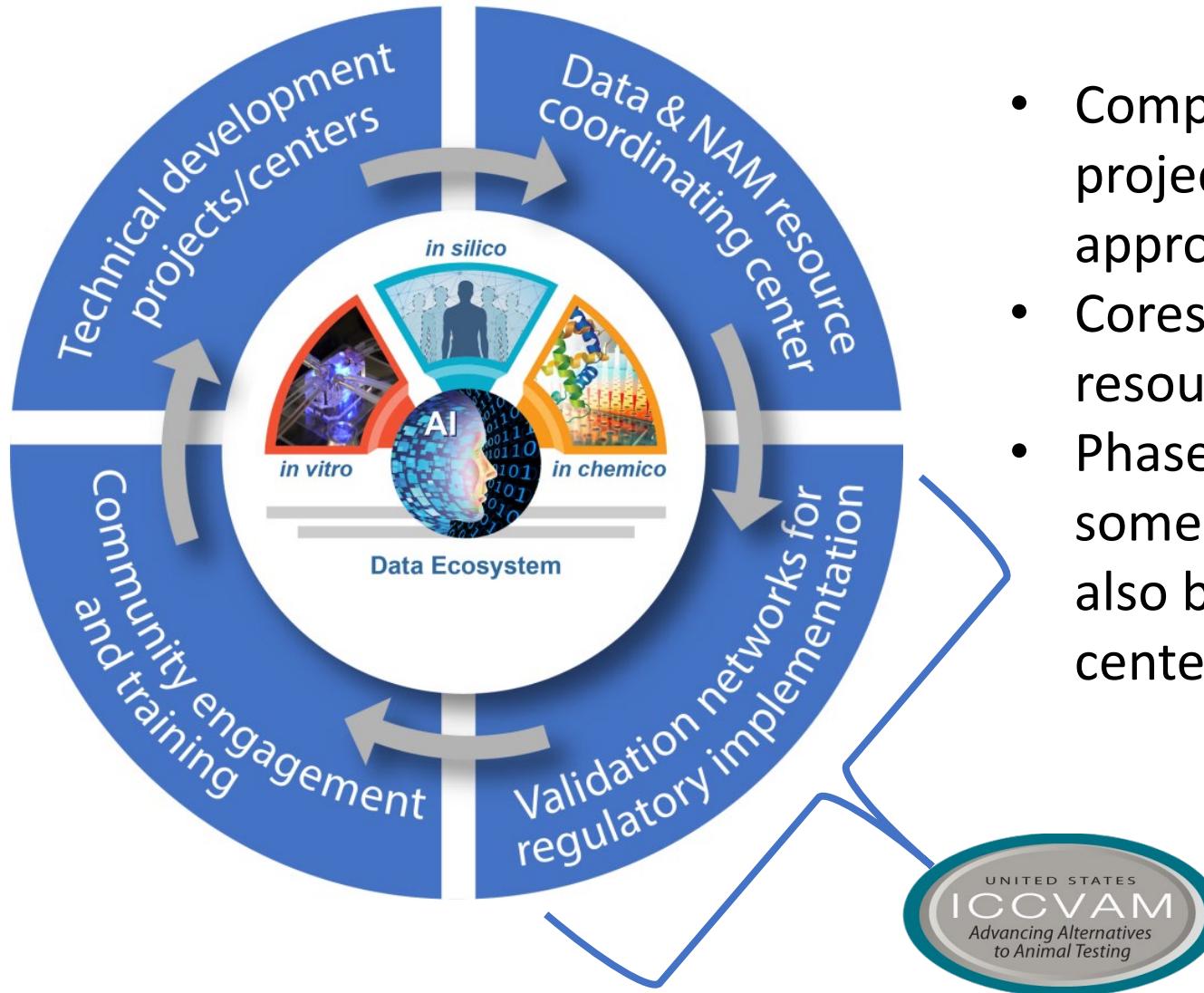
**methodologies (NAMs)** that will transform the way we do basic, translational, and clinical sciences

## Goals:

1. Better model and **understand human health and disease** outcomes **across diverse populations**.
2. Develop NAMs that **provide insight into specific biological processes** or disease states.
3. Validate mature NAMs to **support regulatory use** and standardization.
4. Complement traditional models and **make biomedical research more efficient and effective**.



# Complement-ARIE: Comprehensive center model



- Comprehensive centers will require embedded projects on *in vitro*, *in chemico*, and *in silico* approaches plus combinatorial approaches.
- Cores will include administrative, validation, resources, and training components.
- Phased milestone-driven projects that pilot some of the truly innovative approaches can also be transitioned for integration with the centers.

Key partners for validation networks include: ICCVAM, FDA, EPA, ICATM members, OECD, etc.



# U.S. Validation Body: ICCVAM Authorization Act of 2000

## PUBLIC LAW 106-545 (42 U.S.C. 2851-3)

"To establish, wherever feasible, guidelines, recommendations, and regulations that promote the regulatory acceptance of new or revised scientifically valid toxicological tests that protect human and animal health and the environment while reducing, refining, or replacing animal tests and ensuring human safety and product effectiveness."

- Consumer Product Safety Commission
- Department of Agriculture
- Department of the Interior
- Department of Transportation
- Environmental Protection Agency
- Food and Drug Administration
- Occupational Safety and Health Administration
- National Institute for Occupational Safety and Health
- Agency for Toxic Substances and Disease Registry
- National Cancer Institute



- National Inst of Environmental Health Sciences
- National Library of Medicine
- National Institutes of Health
- Department of Defense
- Department of Energy
- National Institute of Standards and Technology (since 2017)
- Dept of Veterans Affairs Office of Research and Development (since 2020)
- National Center for Advancing Translational Sciences (since 2024)

More information: <https://ntp.niehs.nih.gov/go/iccvam>

ICCVAM Co-chairs



**Suzy Fitzpatrick**  
FDA/CFSAN



**Natalia Vinas**  
DoD



**Nicole Kleinstreuer**  
Executive Director, ICCVAM  
Director, NICEATM

- NTP Interagency Center for the Evaluation of Alternative Toxicological Methods (NICEATM)
- Office within the Division of Translational Toxicology (DTT), National Institute of Environmental Health Sciences (NIEHS)
- Provides scientific leadership and operational support for ICCVAM and ICCVAM workgroup activities
  - NAMs development, evaluation, validation, and implementation
  - Data compilation and review
  - Computational tools development
  - ICCVAM meeting and teleconference support
- Advised by Scientific Advisory Committee on Alternative Toxicological Methods (SACATM)



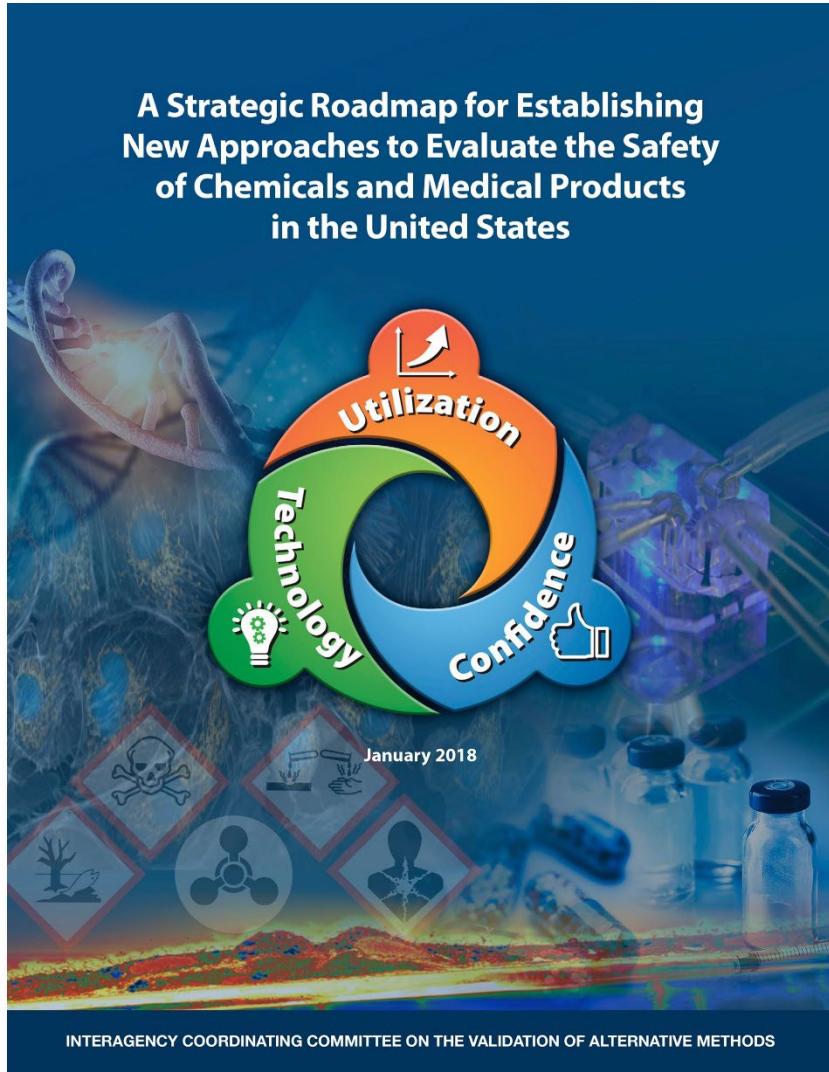
More information: <https://ntp.niehs.nih.gov/go/niceatm>



# Existing ICCVAM Workgroups and Expert Groups

- Acute Toxicity
- Consideration of Alternative Methods
- Ecotoxicology
- In Vitro to In Vivo Extrapolation
- PFAS NAMs
- Read Across
- Validation
- Developmental and Reproductive Toxicology
- Developmental Immunotoxicity
- FAIR Data standards
- Metrics
- Microphysiological Systems
- Nanomaterials
- Ocular and Dermal Irritation
- Skin Sensitization

# Interagency Coordinating Committee on the Validation of Alternative Methods



Connect end users with the developers of alternative methods



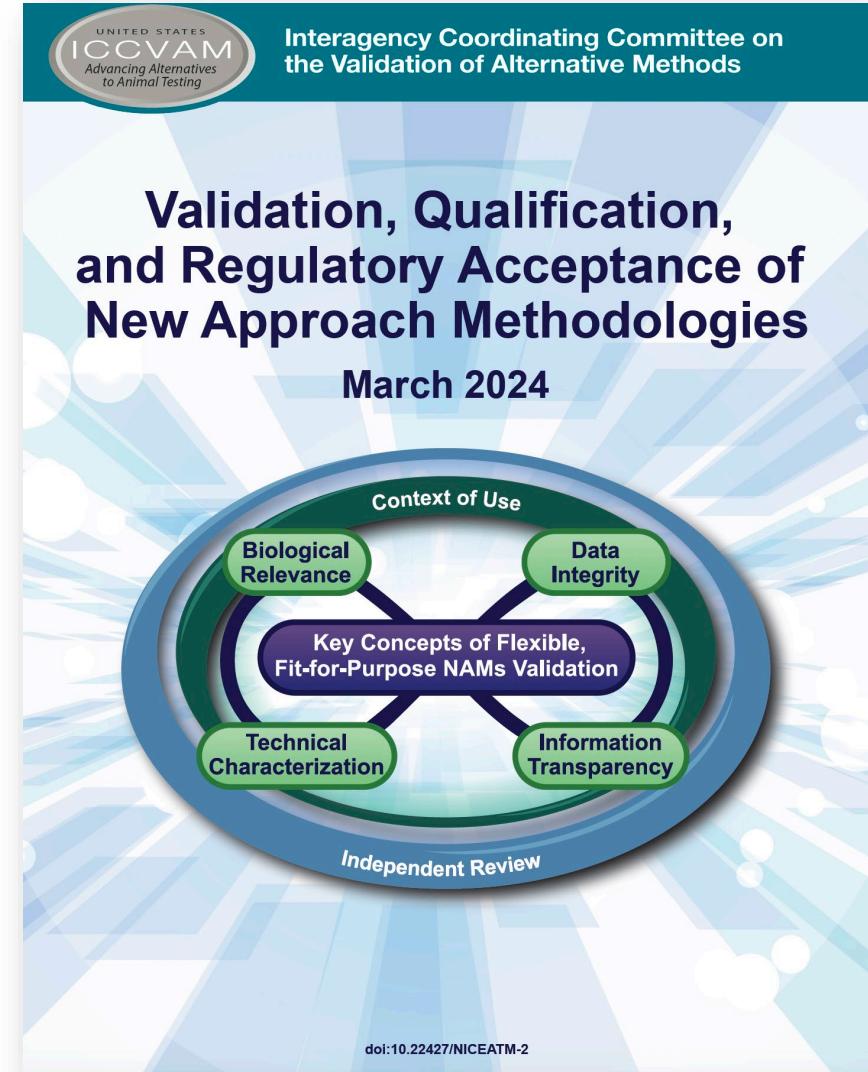
Establish new validation approaches that are more flexible and efficient



Ensure adoption and use of new methods by both regulators and industry



<https://ntp.niehs.nih.gov/go/natl-strategy>



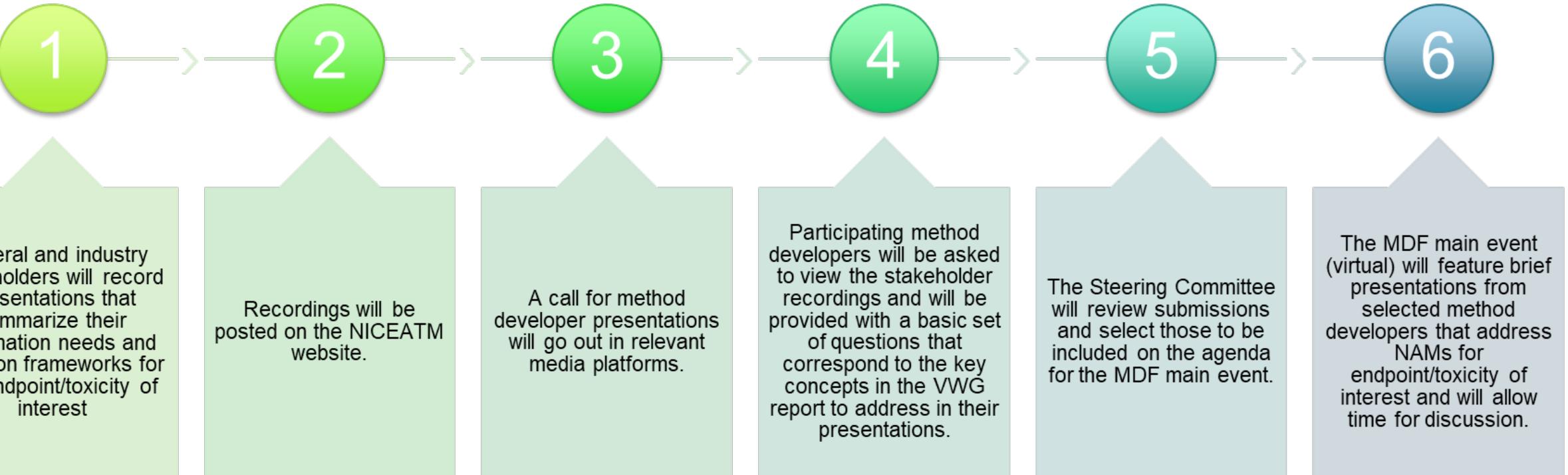
<https://ntp.niehs.nih.gov/go/ICCVAM-submit>

## Method Developers Forum (MDF)

- A proactive effort to highlight and implement the recommendations detailed within the VWG report and provide an opportunity for NAMs developers to interact with stakeholders around regulatory issues.
- Anticipate holding approximately 2-3 MDFs per year.
- Each iteration will focus on a specific endpoint/toxicity.
  - First MDF focused on carcinogenicity (August 21-22, 2024).
  - ICCVAM agencies and industry stakeholders summarize their information needs for carcinogenicity and potential contexts of use for NAMs.
  - Developers demonstrate how their methods address the topic of interest and consider the key concepts from the VWG report in a webinar.
  - 10 submissions were reviewed and approved for presentation
- Future topics include cardiovascular toxicity, developmental toxicity, reproductive toxicity, neurotoxicity, systemic toxicity, specific target organ toxicity (e.g., liver).

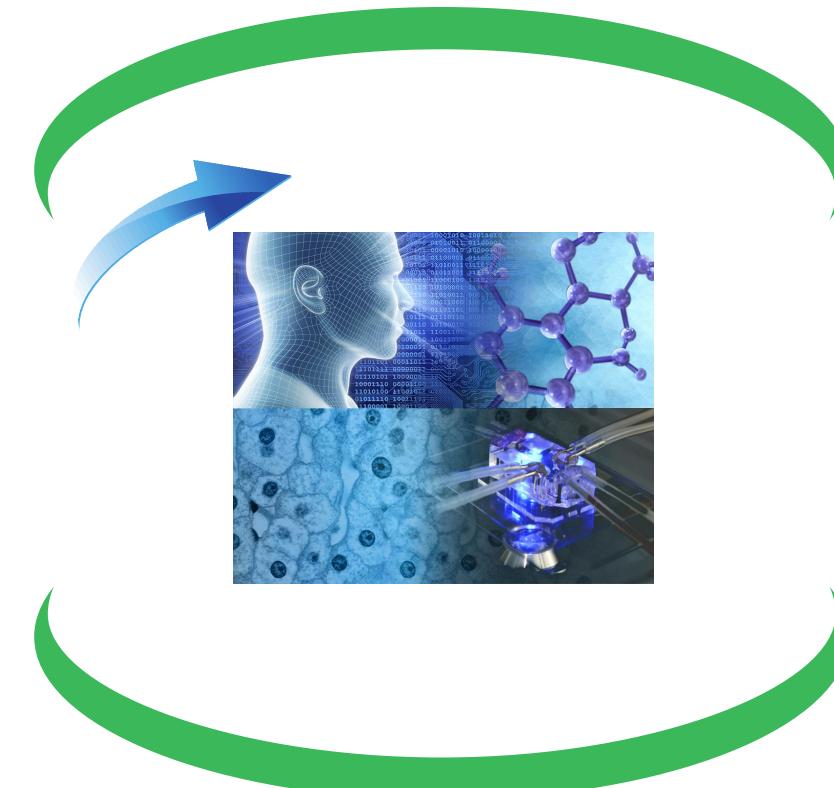


# MDF Process



# Ongoing NICEATM and ICCVAM Projects

- Integrated Chemical Environment
- OPERA (QSAR/QSPR)
- Computational Chemistry
- Quantitative IVIVE
- Reference data curation
- Variability of in vivo data
- Acute Systemic Toxicity
- Dermal absorption
- Eye and skin irritation
- Skin sensitization
- Ecotoxicology
- Carcinogenesis
- Cardiovascular Toxicity
- Developmental Toxicity
- DNT Testing Battery
- Zebrafish models
- Animal-free affinity reagents
- Microphysiological Systems
- Evolving Process of Validation

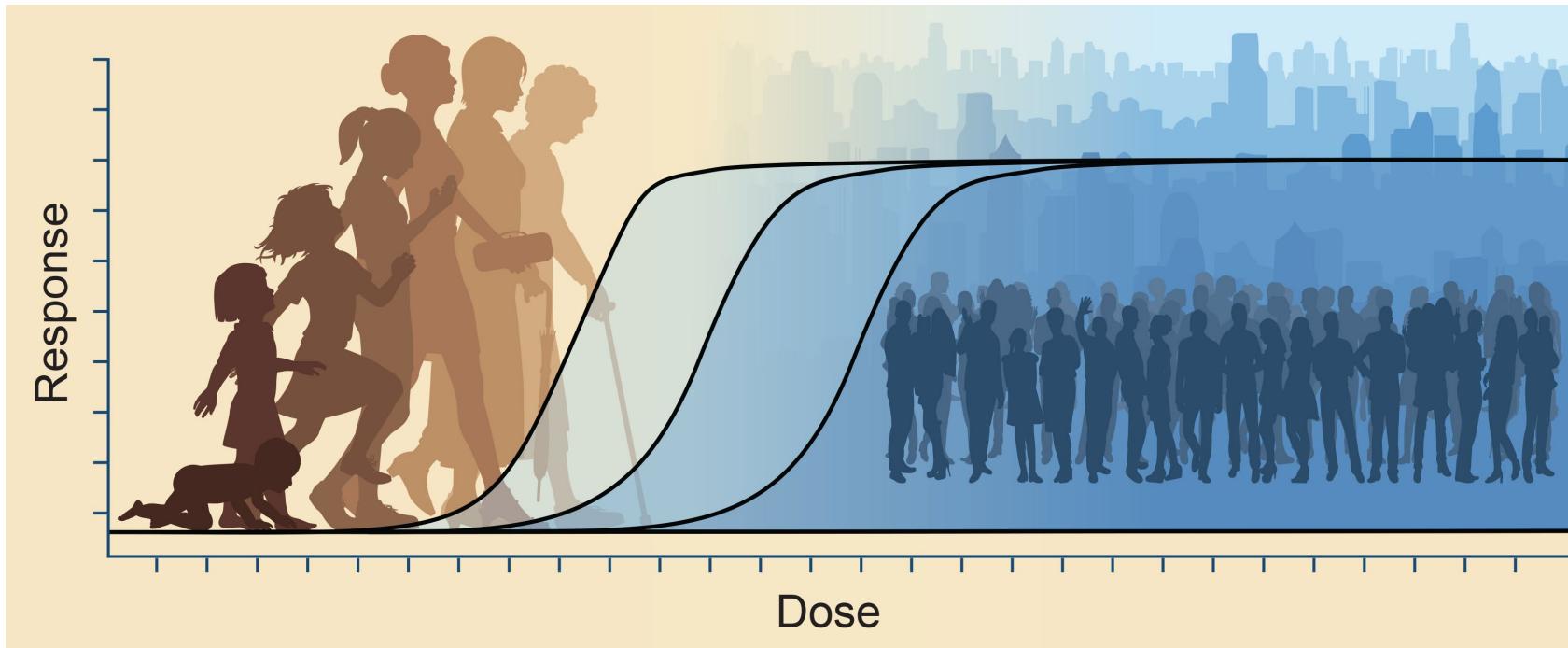


**Integrated  
Chemical  
Environment**



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<https://ntp.niehs.nih.gov/go/niceatm>

## Using New Approach Methodologies to Address Variability and Susceptibility Across Populations



<https://ntp.niehs.nih.gov/go/popvar>

Workshop report in prep to be submitted to Human Genomics shortly



Human Genomics

SPRINGER NATURE GROUP  
SDG Programme  
supporting the Sustainable Development Goals



### Human Genomics Call for Papers

### New Approach Methodologies to Address Population Variability and Susceptibility in Human Risk Assessment

Guest Editors: Helena Hogberg, PhD; Nicole Kleinstreuer, PhD; Kim To, PhD

Submission Status: Open

Read more about the collection

<https://www.biomedcentral.com/collections/NAMAPVS>



Physchem properties		Chemicals	Version
BP	Boiling Point	7860	2.9
HL	Henry's Law Constant	2233	2.9
LogP	Octanol-water Partition Coefficient	18154	2.9
MP	Melting Point	22554	2.9
VP	Vapor Pressure	6764	2.9
WS	Water Solubility	9943	2.9
pKa	Acid Dissociation Constant	6503	2.6
KOA	Octanol/Air Partition Coefficient	270	2.6

Environmental fate		Chemicals	Version
AOH	Atmospheric Hydroxylation Rate	692	2.6
BCF	Bioconcentration Factor	626	2.6
BioHL	Biodegradation Half-life	150	2.6
RB	Ready Biodegradability	1603	2.6
KM	Fish Biotransformation Half-life	541	2.6
KOC	Soil Adsorption Coefficient	728	2.6

ADME properties		Chemicals	Version
FUB	Fraction unbound	3229	2.8
Clint	Intrinsic clearance	1346	2.8
CACO2	Caco-2 permeability	4601	2.8

Toxicity endpoints		Chemicals	Version
ER	Estrogen Receptor Activity	32464	2.6
AR	Androgen Receptor Activity	47673	2.6
AcuteTox	Acute Oral Systemic Toxicity	50660	2.6

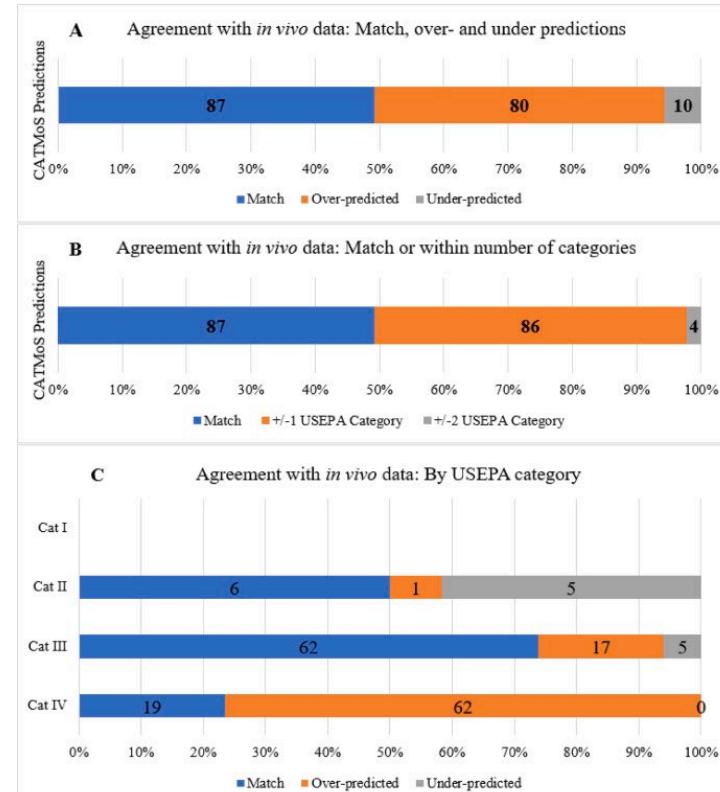
Future models	
Inhalation	Acute Inhalation Systemic Toxicity
SixPack	Acute Toxicity Six-Pack Endpoints
UGT	Glucuronidation: substrate selectivity
SULT	Sulfation: substrate selectivity



# Case Study with EPA Environmental Fate and Effects Division

- Comparative analysis of 177 pesticides with LD<sub>50</sub> data between CaTMOS and EPA database

Toxicity Category based on CaTMos Prediction	Number of predictions	Toxicity Category based on Empirical <i>In Vivo</i> Test Data			
		I	II	III	IV
I (<50 mg/kg)	2	-	1	1	-
II (50-500 mg/kg)	25	-	6	16	3
III (>500-5,000 mg/kg)	126	-	5	62	59
IV (>5,000 mg/kg)	24	-	-	5	19
III and IV combined	150	-	5	145	



Regulatory Toxicology and Pharmacology 149 (2024) 105614



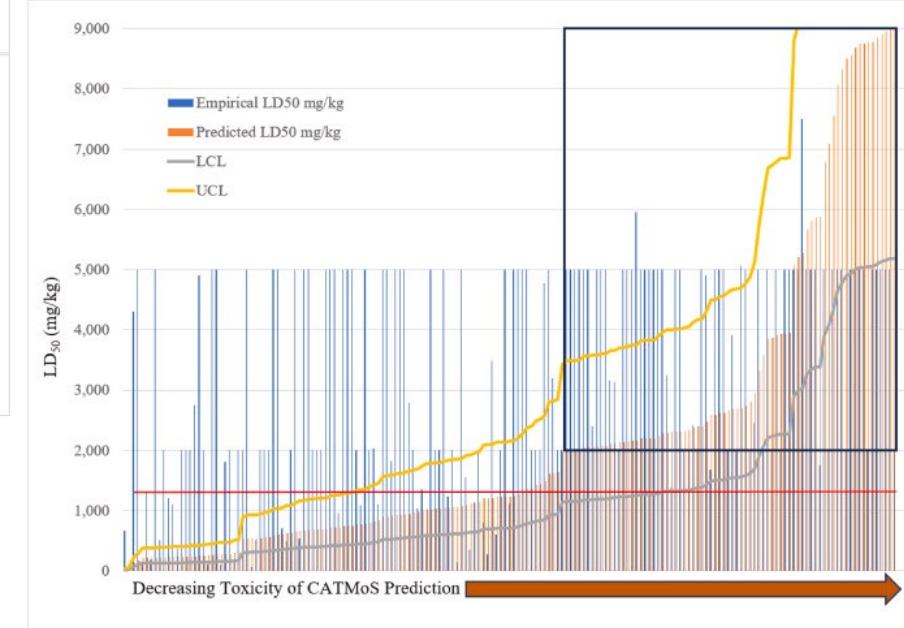
Contents lists available at ScienceDirect

Regulatory Toxicology and Pharmacology

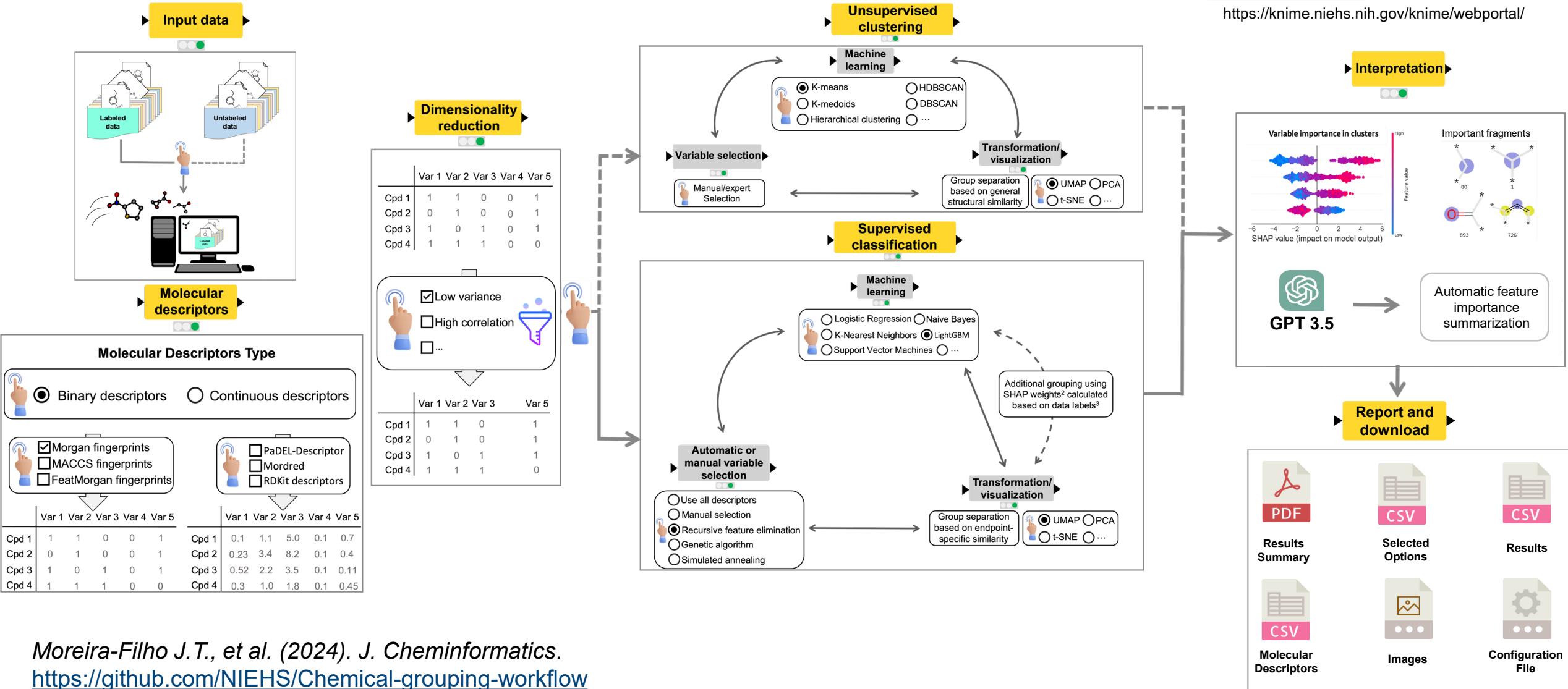
journal homepage: [www.elsevier.com/locate/rtph](http://www.elsevier.com/locate/rtph)

Evaluation of *in silico* model predictions for mammalian acute oral toxicity and regulatory application in pesticide hazard and risk assessment

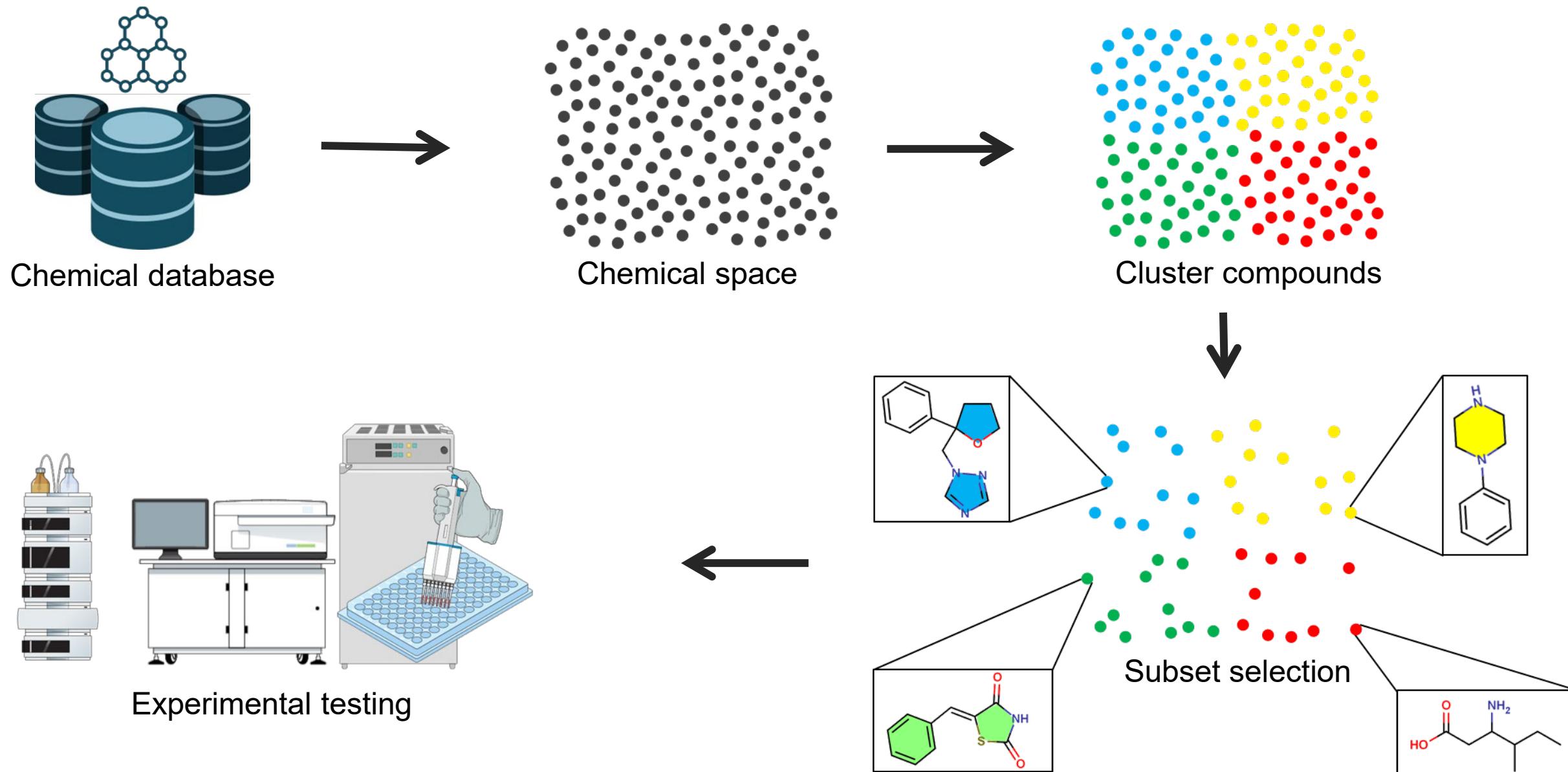
Patricia L. Bishop <sup>a,\*</sup>, Kamel Mansouri <sup>b</sup>, William P. Eckel <sup>c</sup>, Michael B. Lowit <sup>c</sup>, David Allen <sup>d,1</sup>, Amy Blankinship <sup>c</sup>, Anna B. Lowit <sup>e</sup>, D. Ethan Harwood <sup>c</sup>, Tamara Johnson <sup>c</sup>, Nicole C. Kleinstreuer <sup>b</sup>



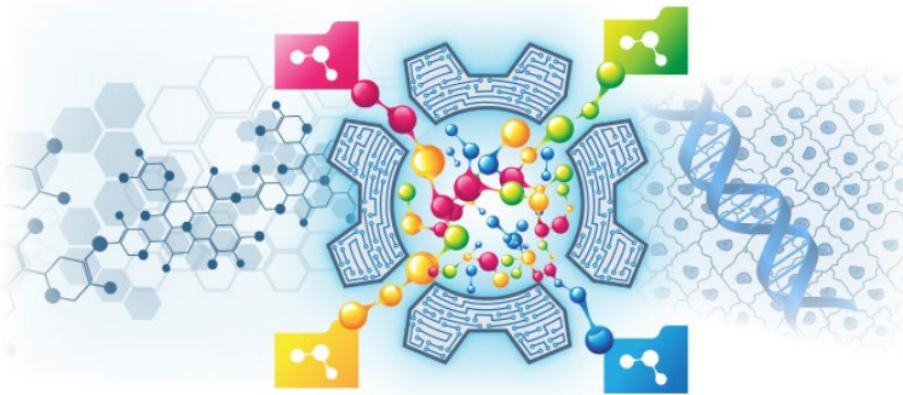
## Chemical Grouping Workflow



# Application – DTT HTS initiative



# Clustering and Classification Workshop



<https://www.niehs.nih.gov/news/events/pastmtg/2022/nams2022/index.cfm>

Convened international experts to discuss methods, their applications to guide toxicology research and inform hazard and risk assessment.

## Accomplishments:

- Defined the concept similarity for supervised and unsupervised approaches
- Introduced different approaches, corrected some misconceptions
- Involved both NAM developers and users
- Established a consortium and a community for increasing communication and collaboration across sectors
- Ongoing and future:* develop and share new ideas/concepts (best practices & innovation)

Mansouri K., et al. (2024). *Env Health Persp*

# Machine Automating Study Data Curation

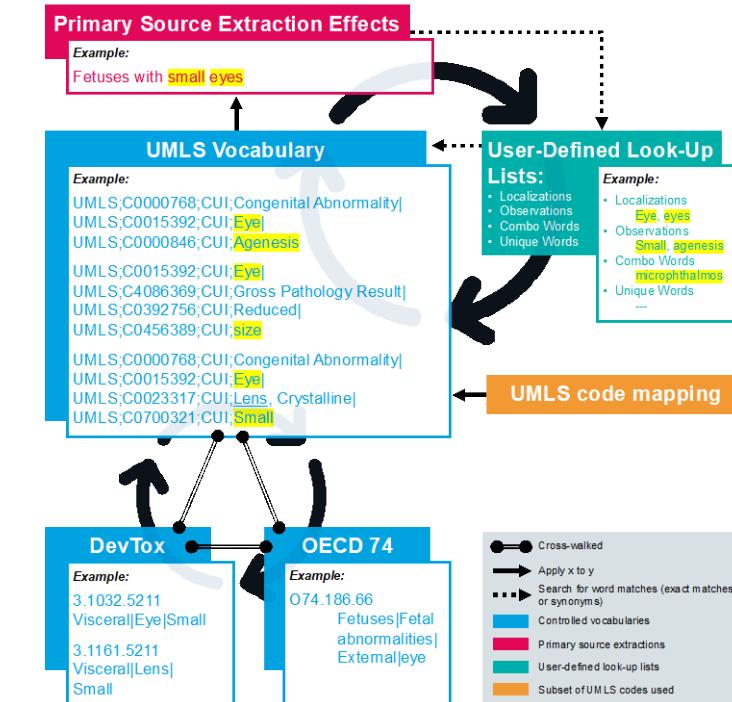
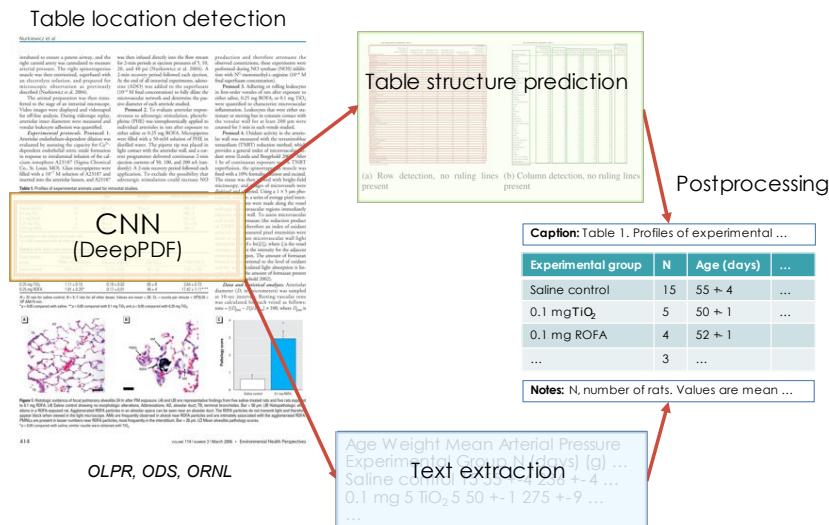
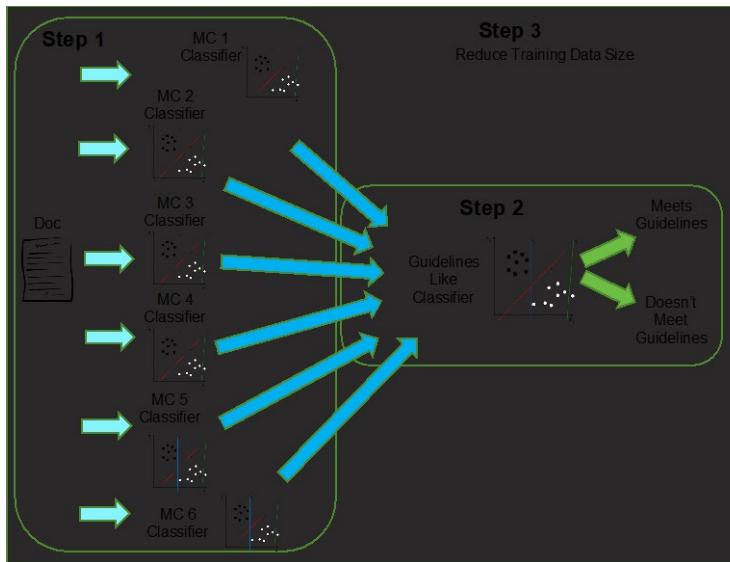
## Identification



## Extraction

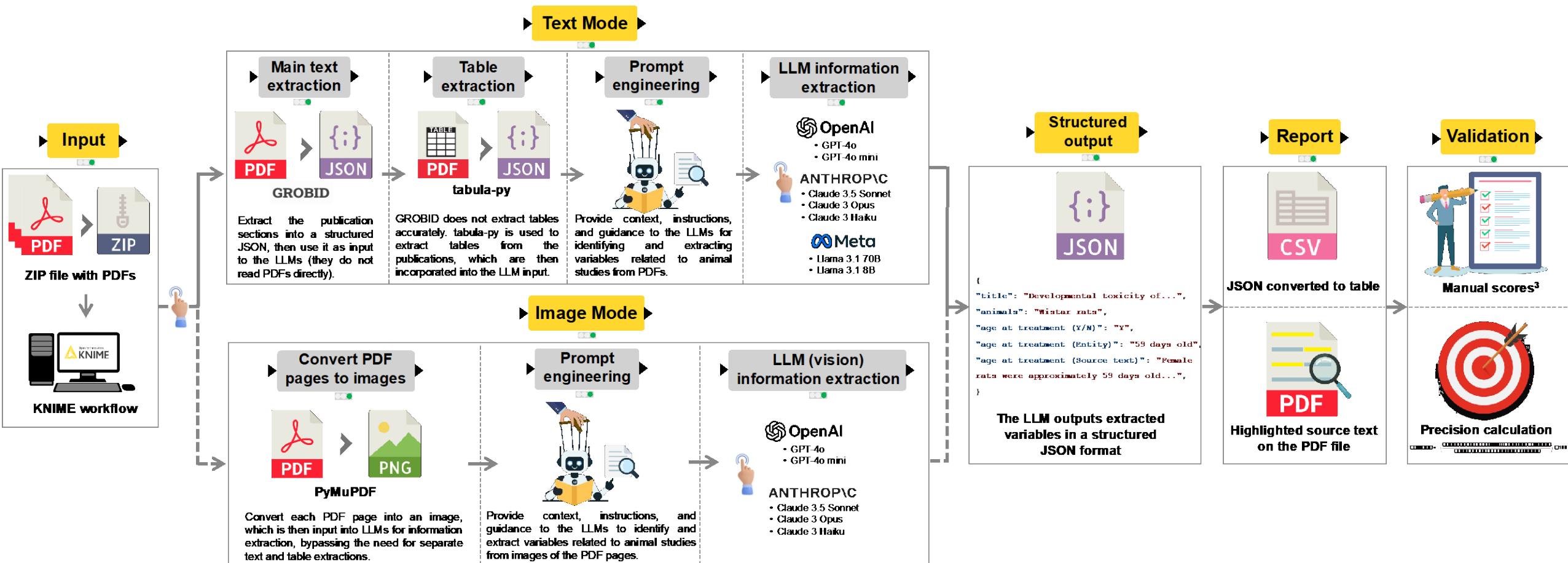


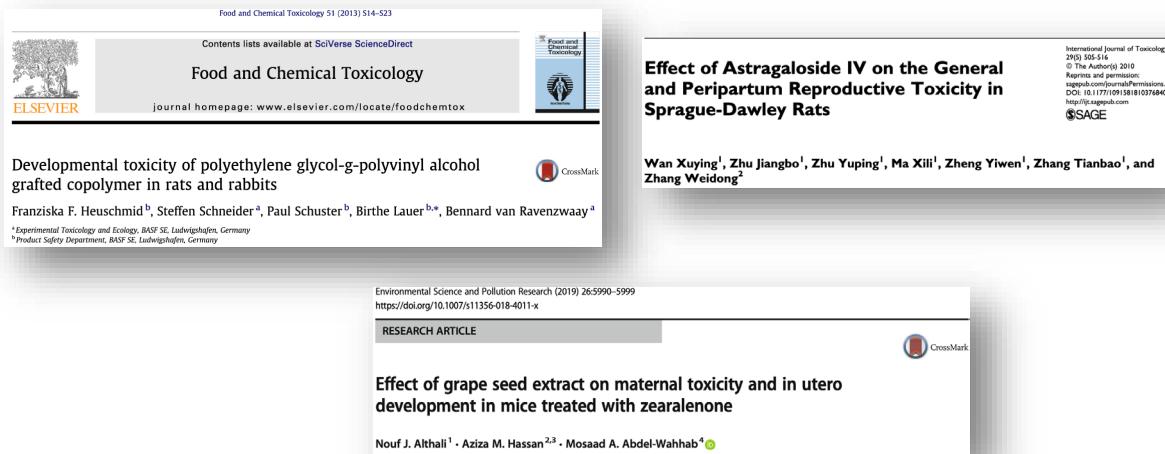
## Annotation



- Important for leveraging high-quality studies in the published literature
- Applications in systematic review of chemical effects
- Establishing reference datasets for validating new methods

# Extraction workflow

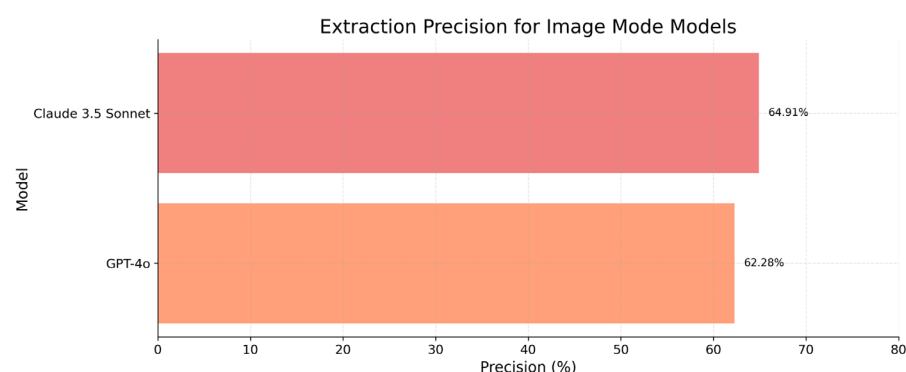
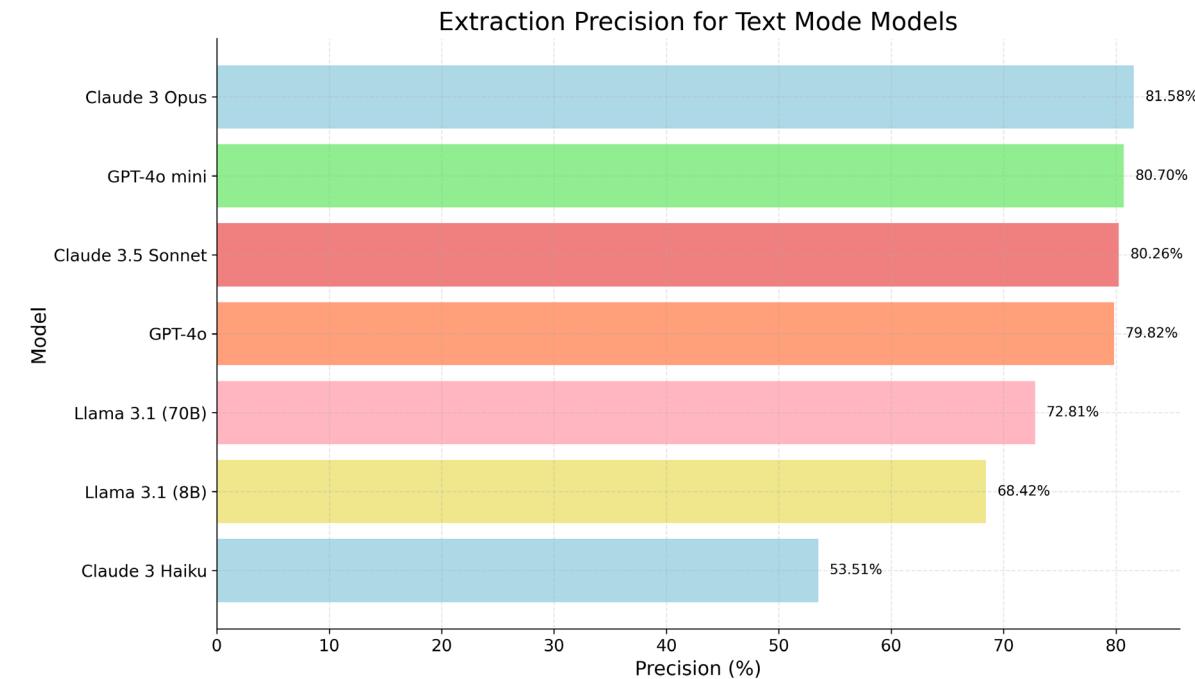




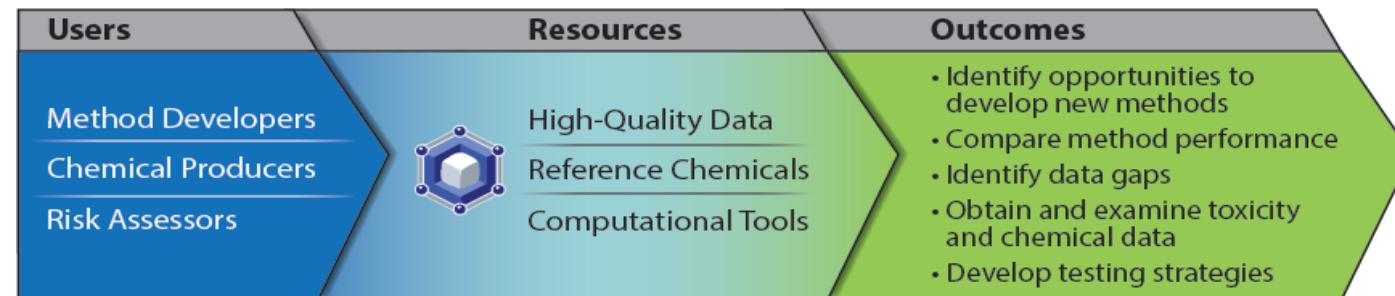
❖ Extracted if the following variables are present (Y/N), the entity, and the source text:

- Title
- Animal
- Age at treatment
- Body weight at treatment
- Number of animals per treatment group
- Route of administration
- Dose
- Daily dosing
- Dose day
- Animal checks during treatment
- Body weights during treatment
- Food consumption during treatment
- Sacrifice
- Maternal body weight at sacrifice
- Fetal body weight at sacrifice, individual
- Fetal body weight at sacrifice, combined
- Uterine weight
- Organ weights
- Pregnancy status
- Number of Live fetuses
- Number of Dead fetuses
- Fetal sex
- Number of Implantation sites
- Number of Corpora lutea
- Number of Resorptions
- Placental evaluation
- Fetal exam, any type
- Fetal external examination
- Fetal visceral examination
- Fetal skeletal examination

# Validation - precision



# ICE: The Integrated Chemical Environment



**Integrated  
Chemical  
Environment**



**F**indable



**A**ccessible



**I**nteroperable



**R**Reusable



Bell et al. 2017 *EHP*

Bell et al. 2020 *Tox In Vitro*

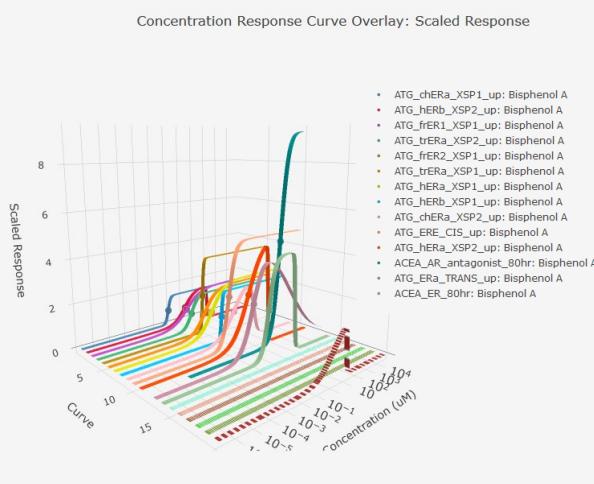
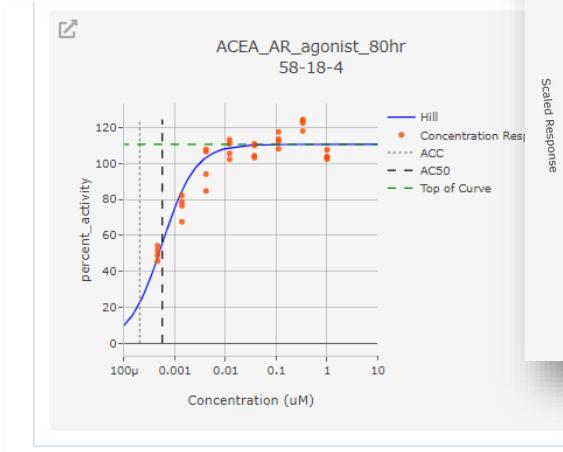
Abedini et al. 2021 *Comp Tox*

Daniel et al. 2022 *Front Toxicol*

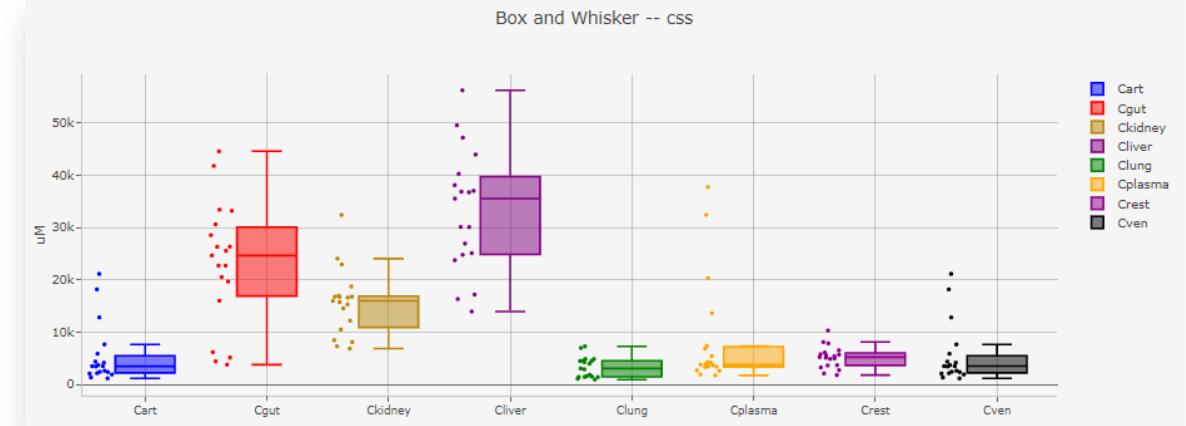
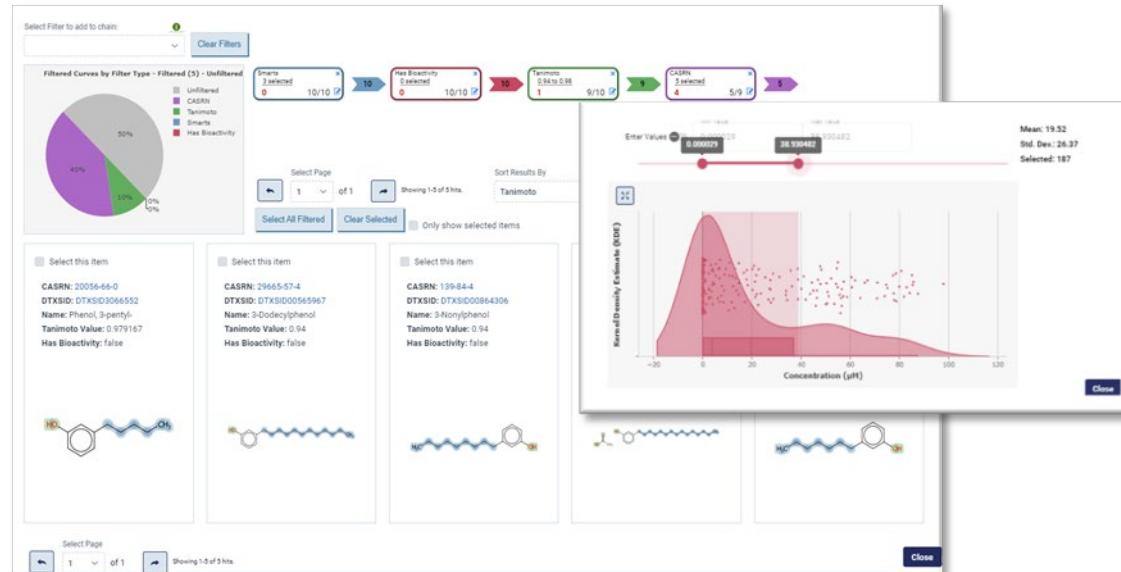
<https://ice.ntp.niehs.nih.gov/>



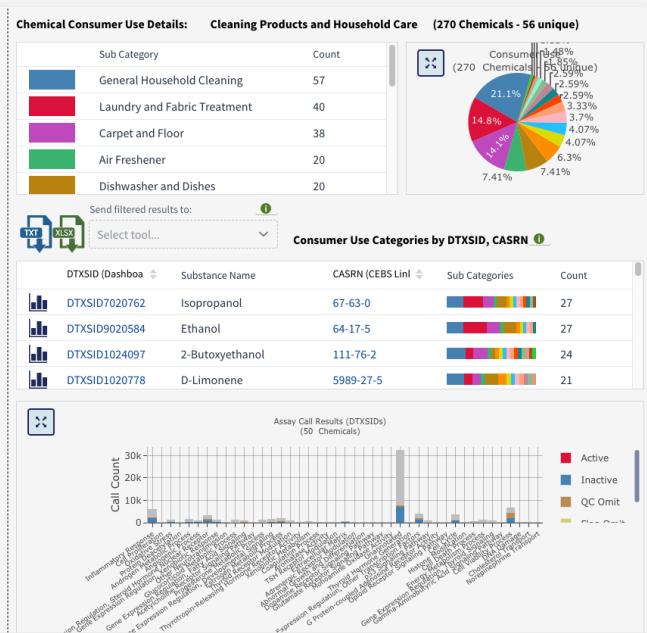
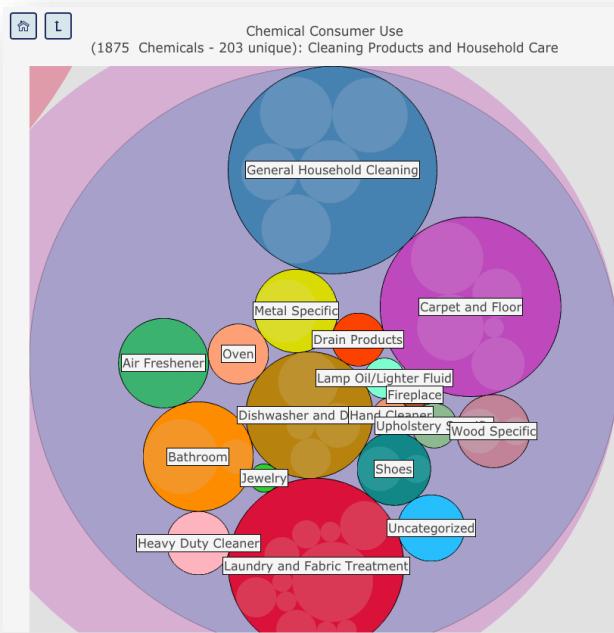
# HTS Data Exploration



## Chemical Similarity

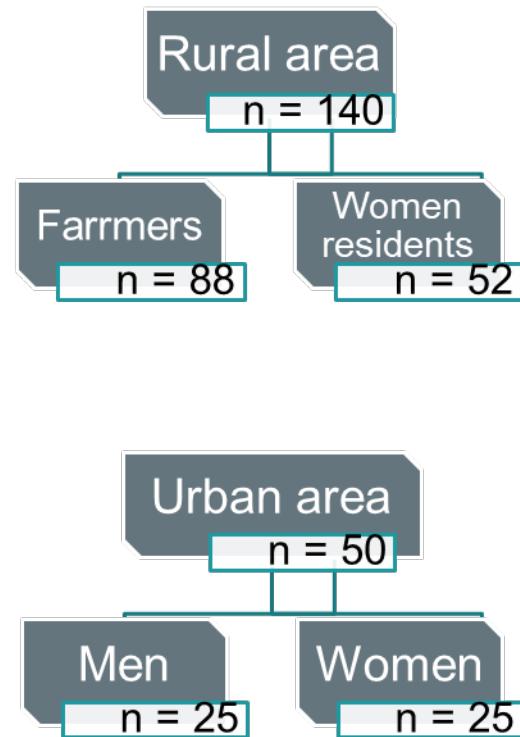


## Predicting Chemical Exposure: Body Tissues, Consumer Products

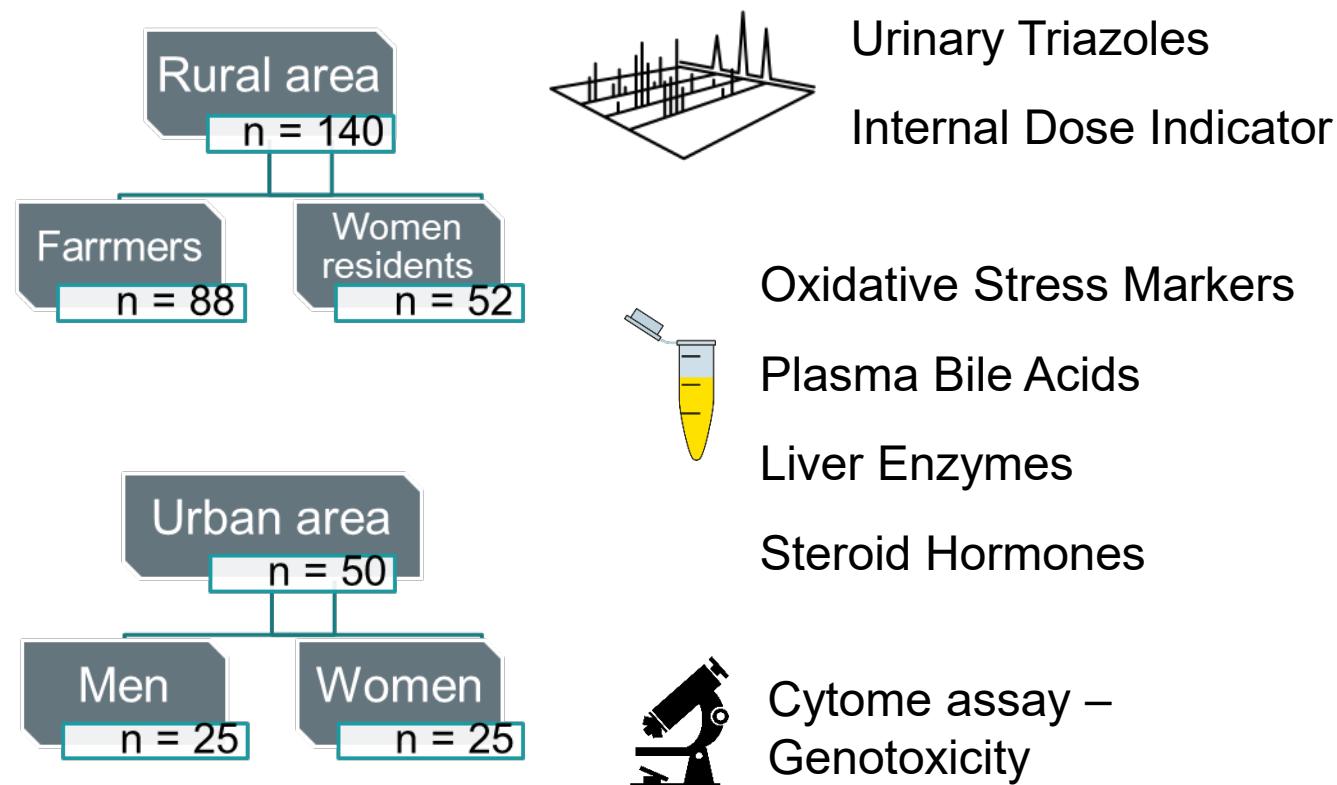


# Risk Characterization of Triazole Fungicides using Human Biomonitoring and Mechanistic Data

## Sampling



## Biomarkers



## Risk Calculations

HQ Calculation at the highest quantified value:

EDI = 6.31 µg/kg-bw/day  
 HQ = 2.1  
Farmers

EDI = 8.77 µg/kg-bw/day  
 HQ = 2.9  
Rural Women Residents

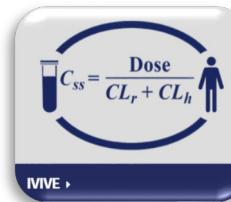


# Association of *in vitro* molecular targets and human biomarker alterations



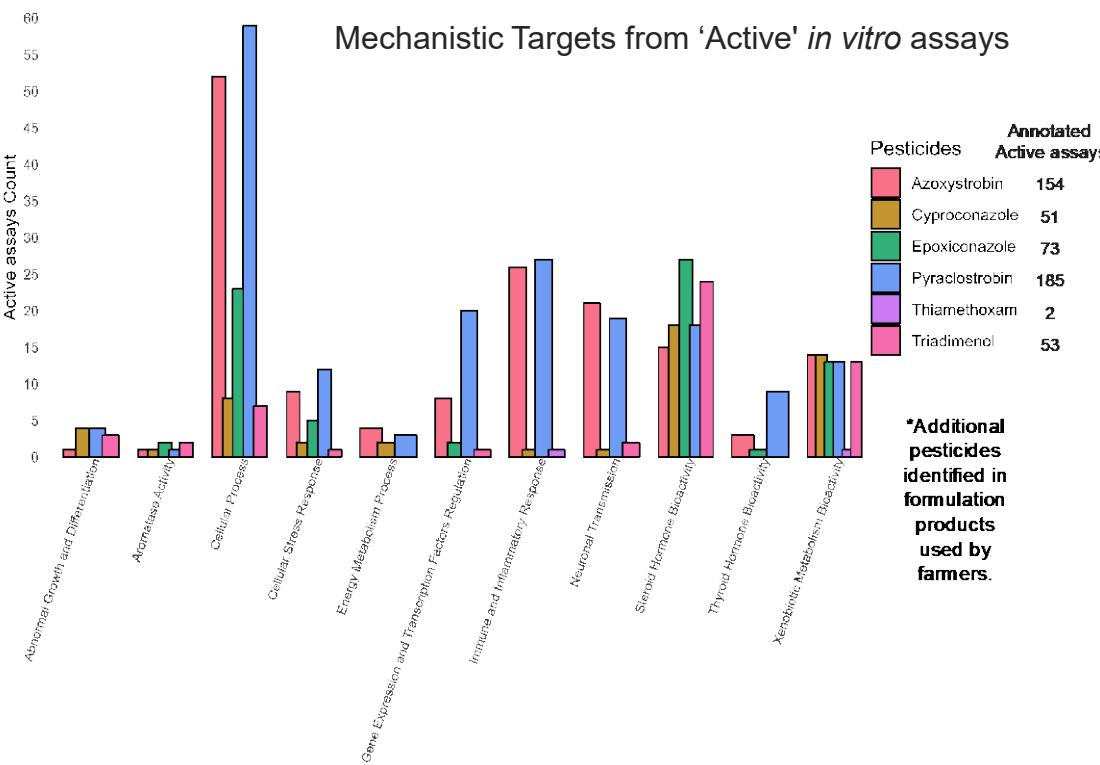
## Concentration-Response Curves

- cHTS data from Tox21/ToxCast

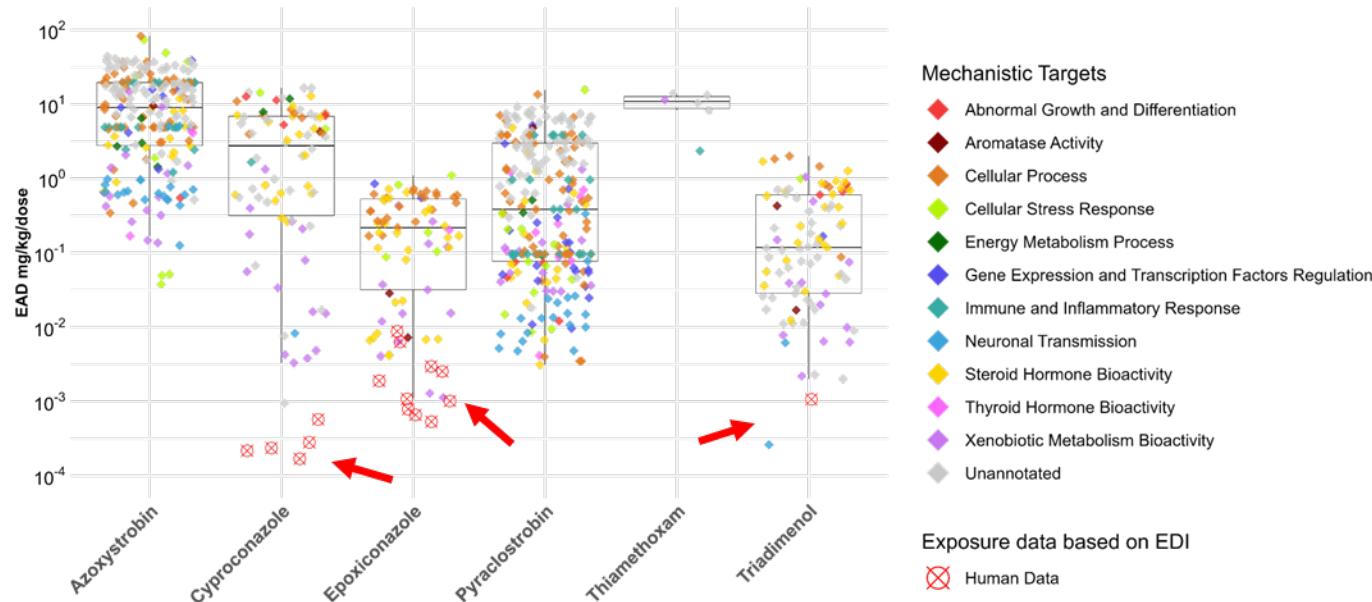


## *In Vitro* to *In Vivo* Extrapolation

- Calculate equivalent doses from cHTS data
- Comparison with human exposure

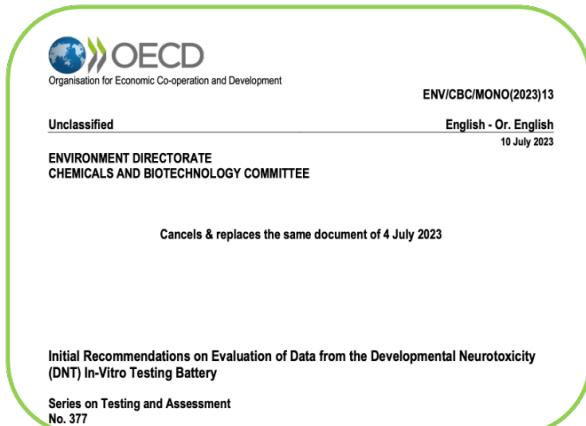


## Solve\_oral\_pbtk model



# Integrated Approach to Testing and Assessment (IATA) DNT Case Study for Prioritization

Guidance document to inform on the DNT IVB, its usage and interpretation



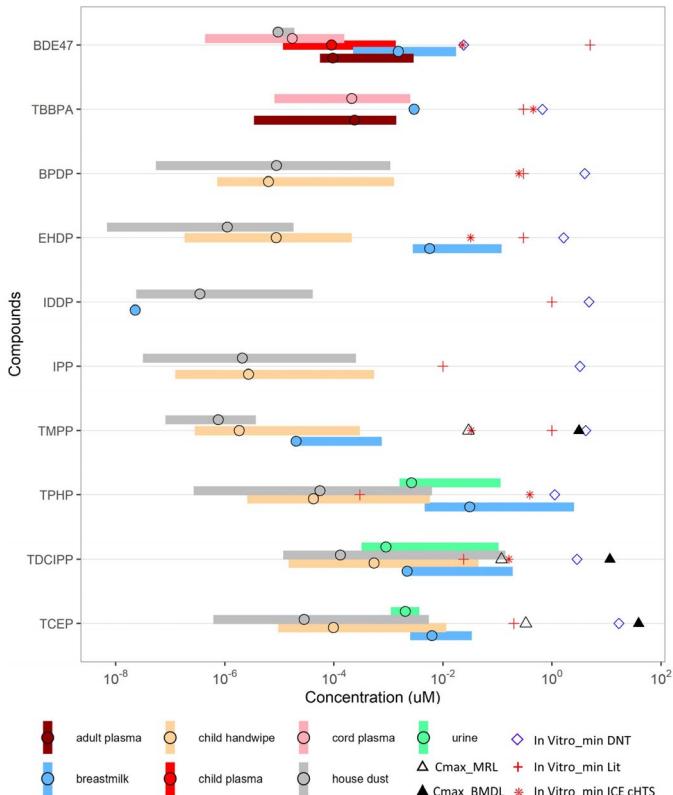
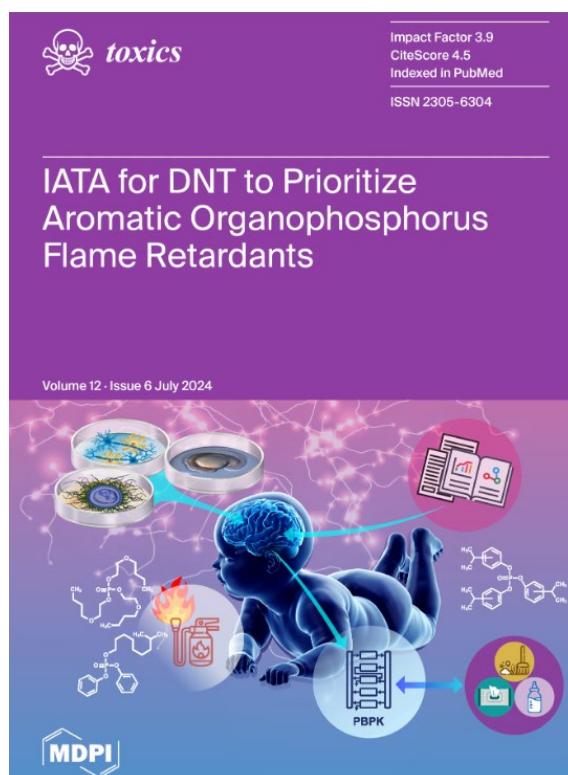
IATA case studies to exemplify different regulatory needs



Case study on the use of Integrated Approaches for Testing and Assessment for DNT to prioritize a class of Organophosphorus flame retardants

Series on Testing and Assessment  
No. 364

Updated IATA case study for prioritization



Kreutz A et al. (2024)  
Toxics 12(6):437



# Catalyze the Development and Use of NAMs

- Identify the types of alternative methods and assess their general strengths and weaknesses for studying human biology, circuits, systems, and disease states
- Characterize the types of research, condition, or disease for which NAMs are most applicable or beneficial
- Articulate high-priority areas for NIH investment in the **use and development with human applicability** to:
  - Advance progress into understanding specific biological processes or states
  - Augment the tools and capabilities for biomedical research to complement and/or potentially replace traditional models

# Implementing the ACD NAMs WG recommendations

## RECOMMENDATIONS TO CATALYZE THE DEVELOPMENT AND USE OF NAMs

Rectangular Snip

Recommendation 1.	Prioritize the development and use of combinatorial NAMs.
Recommendation 2.	Establish resources, infrastructure, and collaborations to promote the use of interoperable, reliable, and well curated/high quality datasets produced from research using NAMs.
Recommendation 3.	Promote effective dissemination and interconnection of NAMs technologies.
Recommendation 4.	Invest in comprehensive training to bolster continuous advances in NAMs development and use.
Recommendation 5.	Facilitate multidisciplinary teams with expertise across technologies and the lifecycle of NAMs development and use.
Recommendation 6.	Promote social responsibility in both the creation and deployment of NAMs across the research lifecycle.
Recommendation 7.	Support and maintain coordinated infrastructure to catalyze effective and responsible NAM development and use.

## New interactive database of validated/qualified NAMs



# Acknowledgments

## The NICEATM Group



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**Integrated  
Chemical  
Environment**

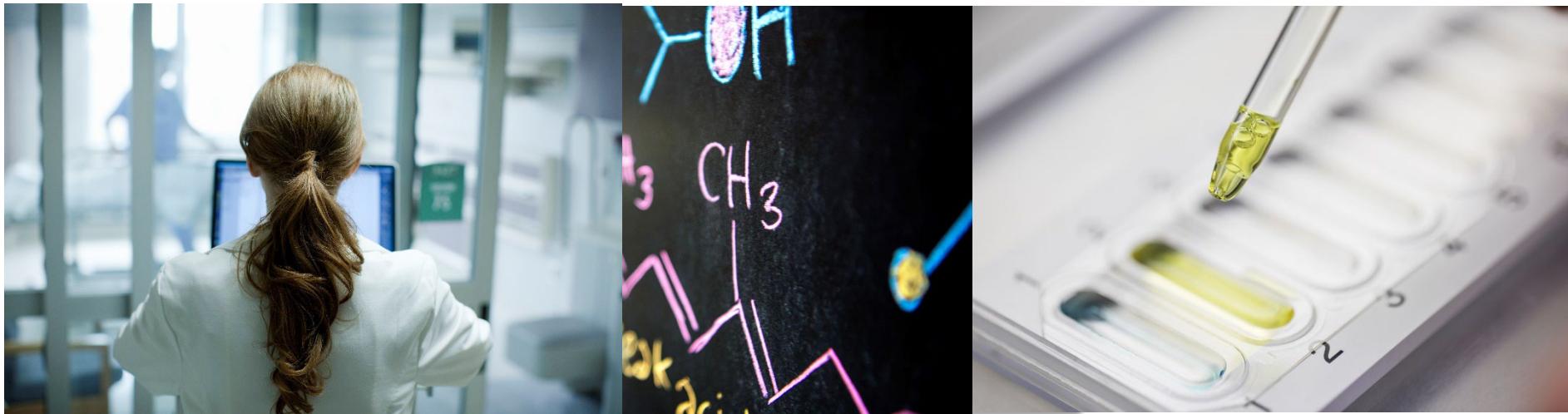


**Now Available:  
2022 – 2023 ICCVAM  
Biennial Report**

# **Building Confidence in NAMs via Validation Standard Setting in a Revised OECD GD 34**

**Alison Harrill, PhD**

Associate Director for Toxicology, Center for Computational Toxicology and Exposure



The views expressed in this presentation are those of the presenter and do not represent the views or policies of the U.S. EPA

# OECD Guidance Document 34

Unclassified

ENV/JM/MONO(2005)14

18-Aug-2005

English - Or. English

Organisation de Coopération et de Développement Economiques  
Organisation for Economic Co-operation and Development

ENVIRONMENT DIRECTORATE  
JOINT MEETING OF THE CHEMICALS COMMITTEE AND  
THE WORKING PARTY ON CHEMICALS, PESTICIDES AND BIOTECHNOLOGY

OECD SERIES ON TESTING AND ASSESSMENT  
Number 34

GUIDANCE DOCUMENT ON THE VALIDATION AND INTERNATIONAL ACCEPTANCE OF NEW  
OR UPDATED TEST METHODS FOR HAZARD ASSESSMENT

- Expert group working to update
- Pre-dates development of many in vitro NAM and in silico approaches
- Pre-dates the institution of the Defined Approach
- Best practices have changed somewhat

ENV/JM/MONO(2005)14  
Unclassified

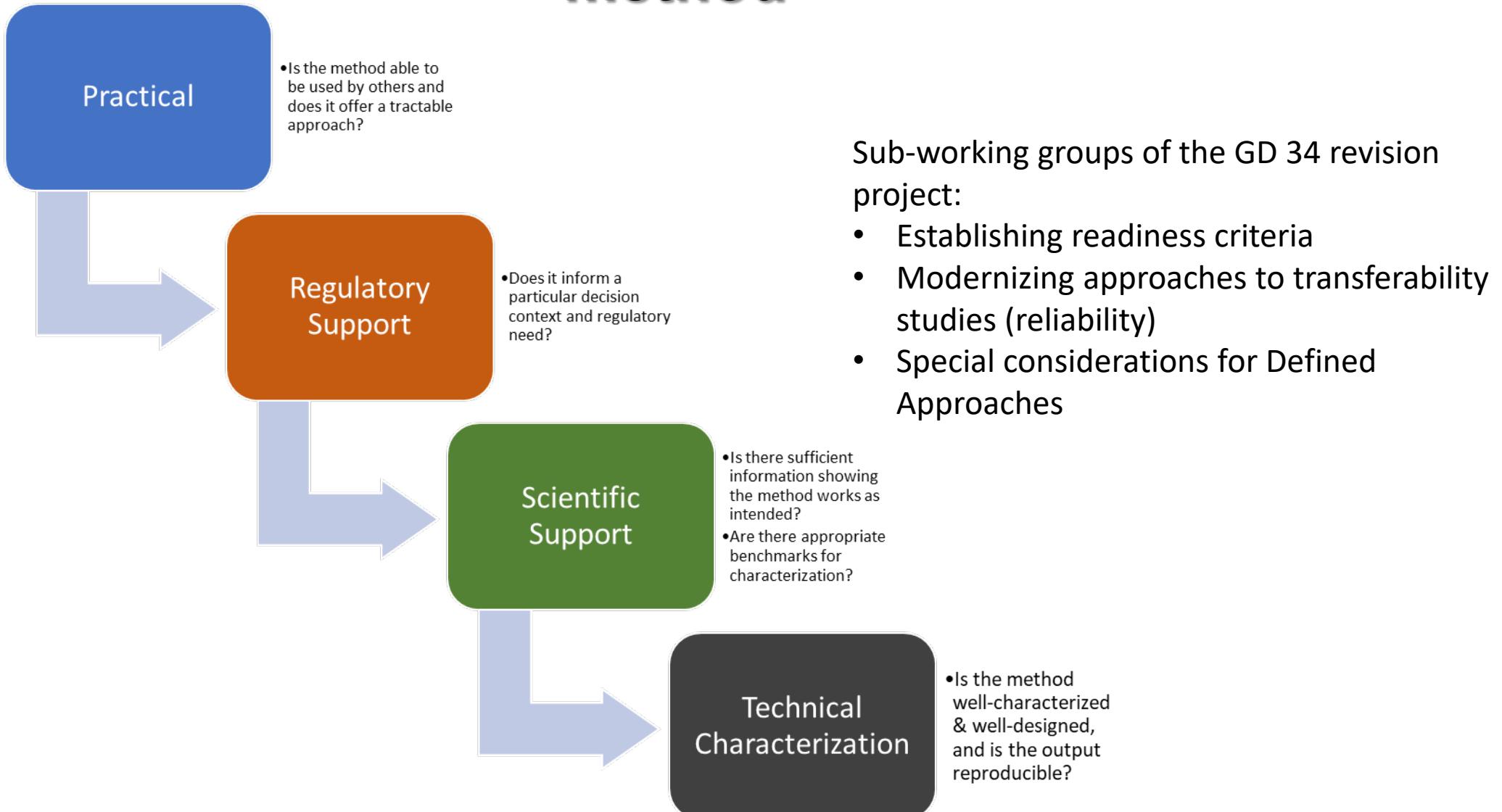
# Definitions and Recommendations from Existing GD34 Guidance

- OECD definition of “validation” as a “*process based on scientifically sound principles (5)(6) by which the reliability and relevance of a particular test, approach, method, or process are established for a specific purpose.*”
- Reliability is defined as “*the extent of reproducibility of results from a test within and among laboratories over time, when performed using the same standardised protocol.*”
- The relevance of a test method describes “*the relationship between the test and the effect in the target species and whether the test method is meaningful and useful for a defined purpose, with the limitations identified.*”
- Other recommendations...
  - “*the validation process should be flexible and adaptable*”,
  - performance must be “*demonstrated using a series of reference chemicals*”, and
  - “*evaluated in relation to existing relevant toxicity data.*”

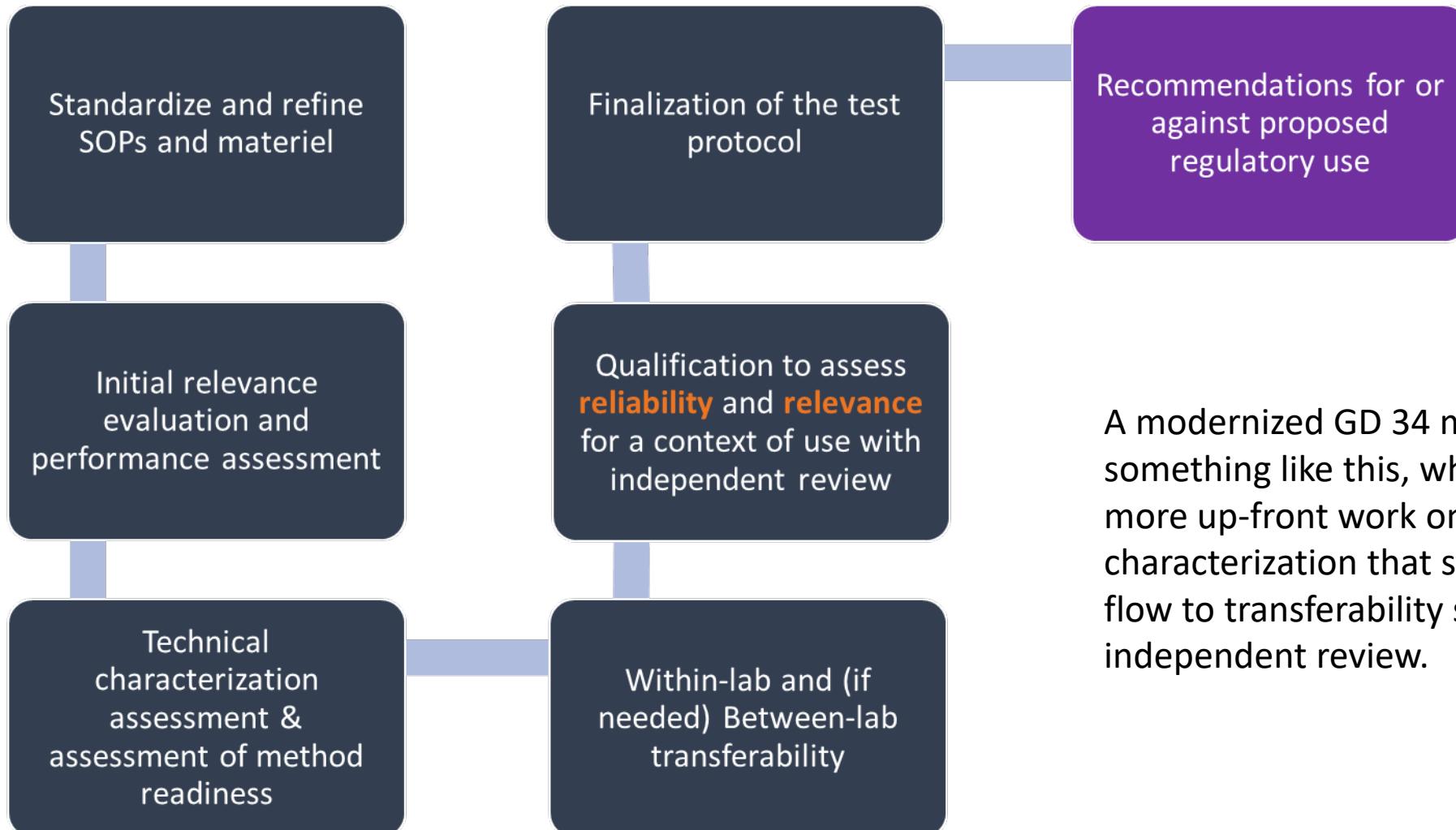
Mutual Acceptance of Data (MAD) system dictates data to be accepted from OECD TGs by all participating countries.

The foundation of the MAD system is the approximately 150 OECD Test Guidelines (methods).

# Building international scientific support for a new method



# Envisioning a more streamlined GD 34 process

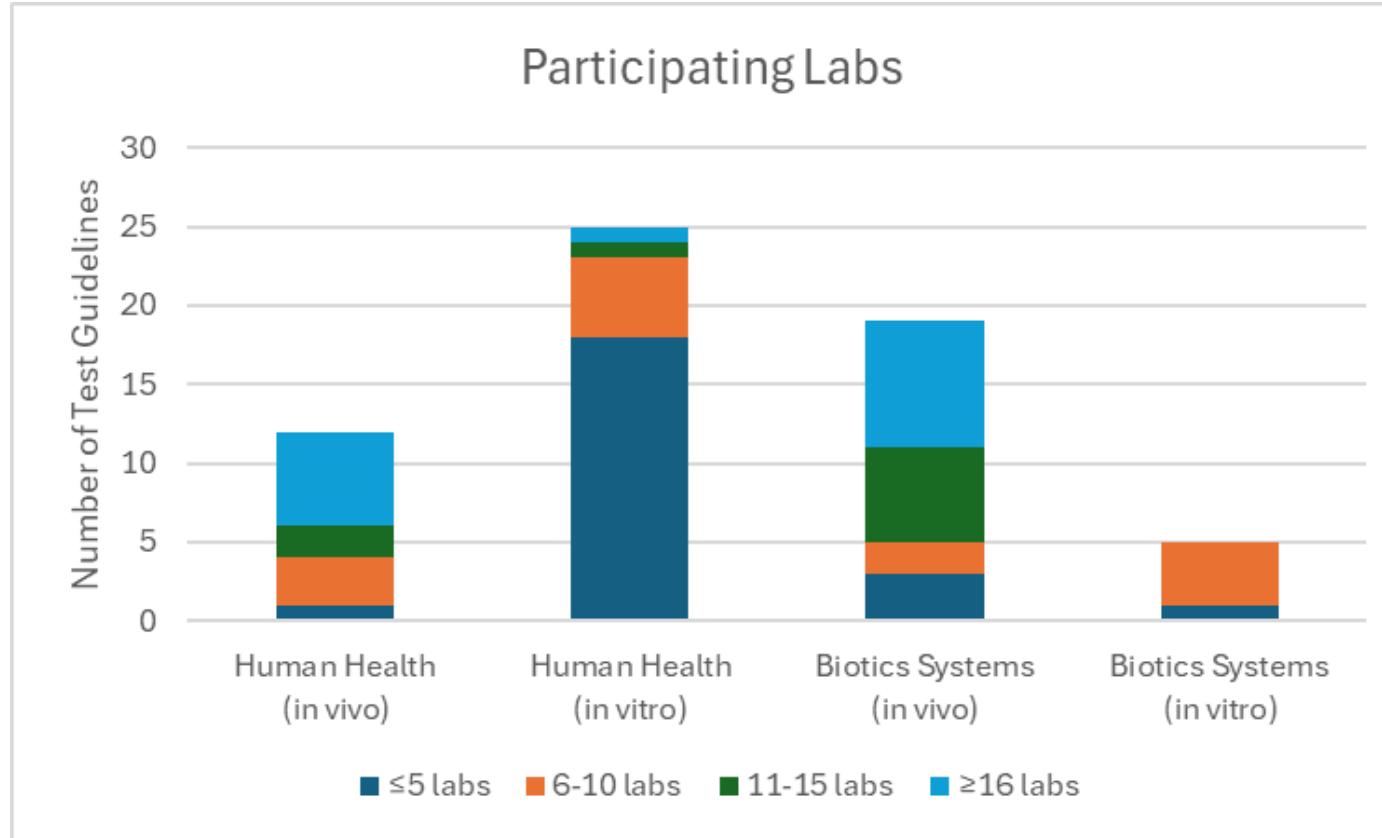


A modernized GD 34 might look something like this, where there is more up-front work on technical characterization that streamlines flow to transferability studies and independent review.

# What do validation efforts typically entail for OECD Test Guidelines?

- Transferability studies are typically performed across multiple labs to assess the variability of point estimates of the readout across labs (how reproducible is the result?)
- While all of the OECD Test Guidelines have been *standardized* into a formal method, a subset of these have been formally ‘validated’ using between-lab transferability studies
- To assess assay reliability in a validation study, performance is frequently assessed in both quantitative terms (variability around a point estimate) AND qualitative terms (how well the assay predicts the endpoint)
- To validate an assay, lists of reference chemicals with association (or lack thereof) to the measured effect are used to assess assay performance and reproducibility (reliability)

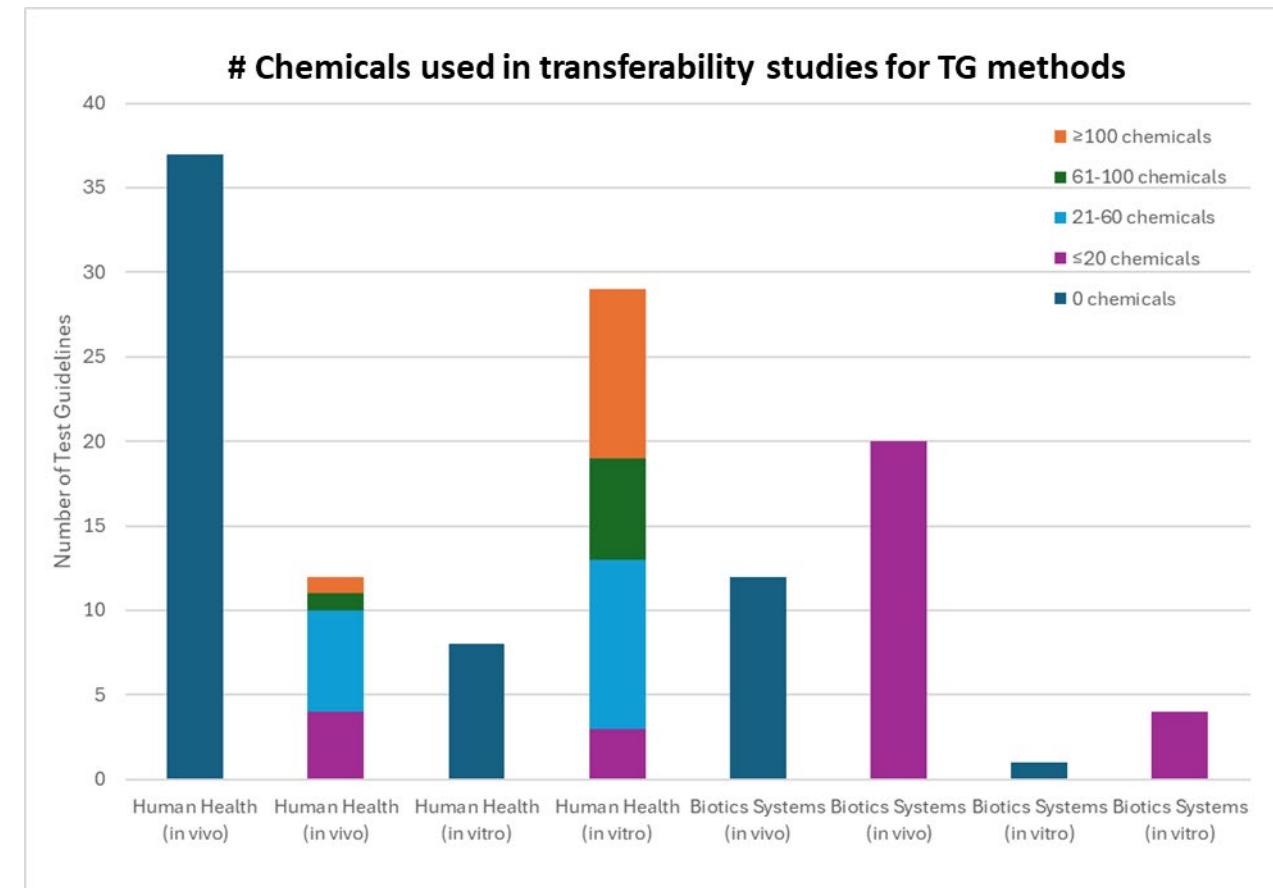
# Numbers of labs involved in transferability and validation studies that led to OECD TGs



- Not all OECD records of transferability studies had accessible data on the numbers of participating labs
- Participating lab numbers were based on the particular phase of validation that had greatest number of participating labs

# Not all OECD TGs have undergone transferability studies to assess method reliability

Test Guideline Type	Total TGs	Total with Transferability	% with Transferability Studies
Human Health (in vivo)	49	12	24%
Human Health (in vitro)	36	29	81%
Biotics Systems (in vivo)	34	21	62%
Biotics Systems (in vitro)	6	5	83%



# Expectations for the size of reference chemical sets for NAMs exceeds those of *in vivo* TGs

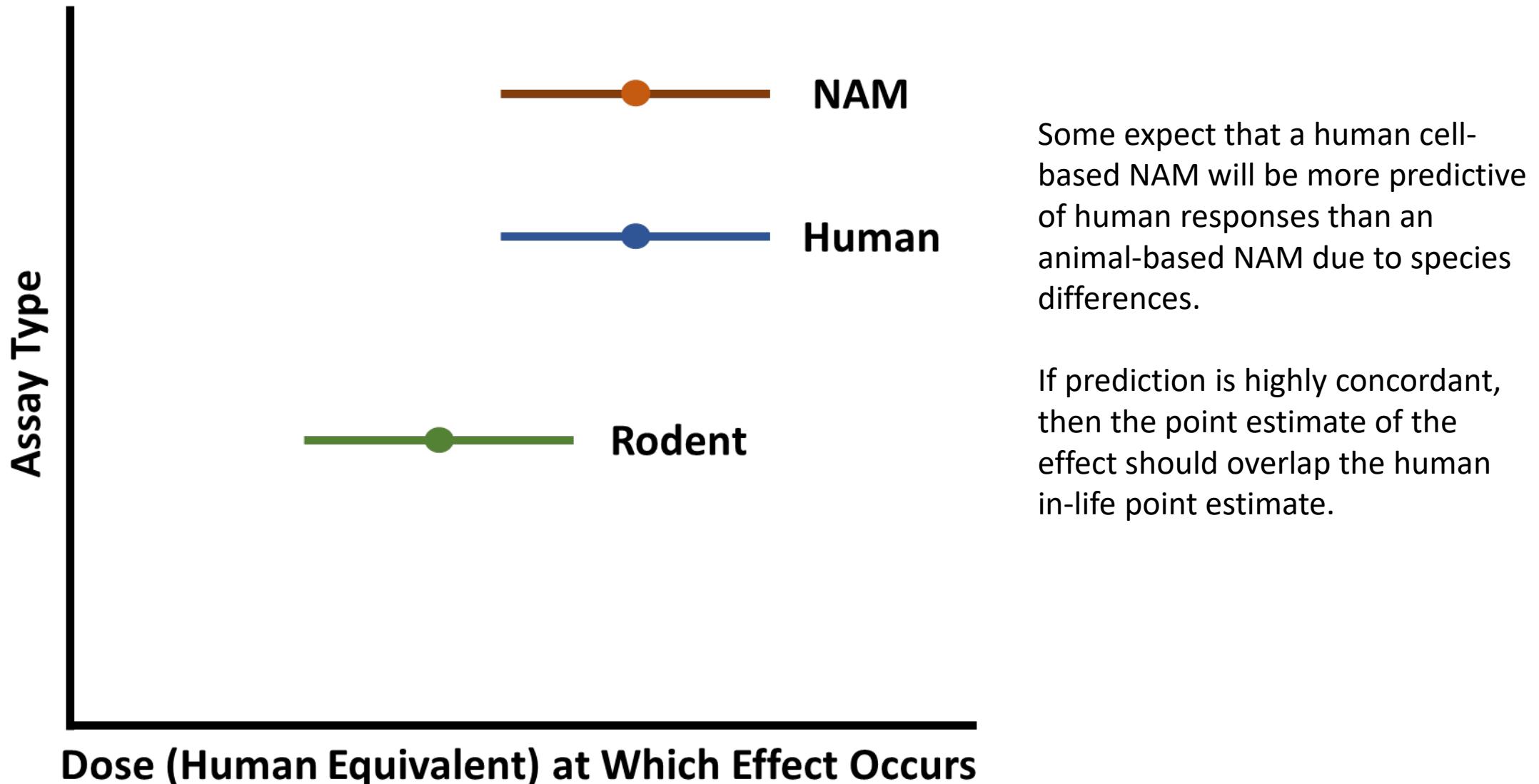
- Data were extracted from 125 TGs covering vertebrate and invertebrate species and associated in vitro alternatives
- *In vitro* methods for both human health and biotic systems had a larger proportion of TGs that had been formally validated (81 and 83%) and had larger numbers of chemicals associated with the validation studies overall
  - 55% of validated *in vitro* human health TGs > 61 chemicals, 34% had greater than 100 chemicals
  - 16% of human health *in vivo* studies with >60 chemicals, only 8% had greater than 100 chemicals
  - Majority of all biotic systems validation studies had 20 or fewer chemicals (1 *in vitro* with 61-100 chemicals)
- *In vitro* test methods overall have larger chemicals sets for transferability and greater numbers of validation studies, although *in vivo* human health appeared to have larger numbers of participating labs
- One reason for the increase in numbers of test chemicals being used is that assessing reliability and relevance may be challenging, and doing so requires appropriate benchmarks.

*Data are preliminary and analysis is ongoing*

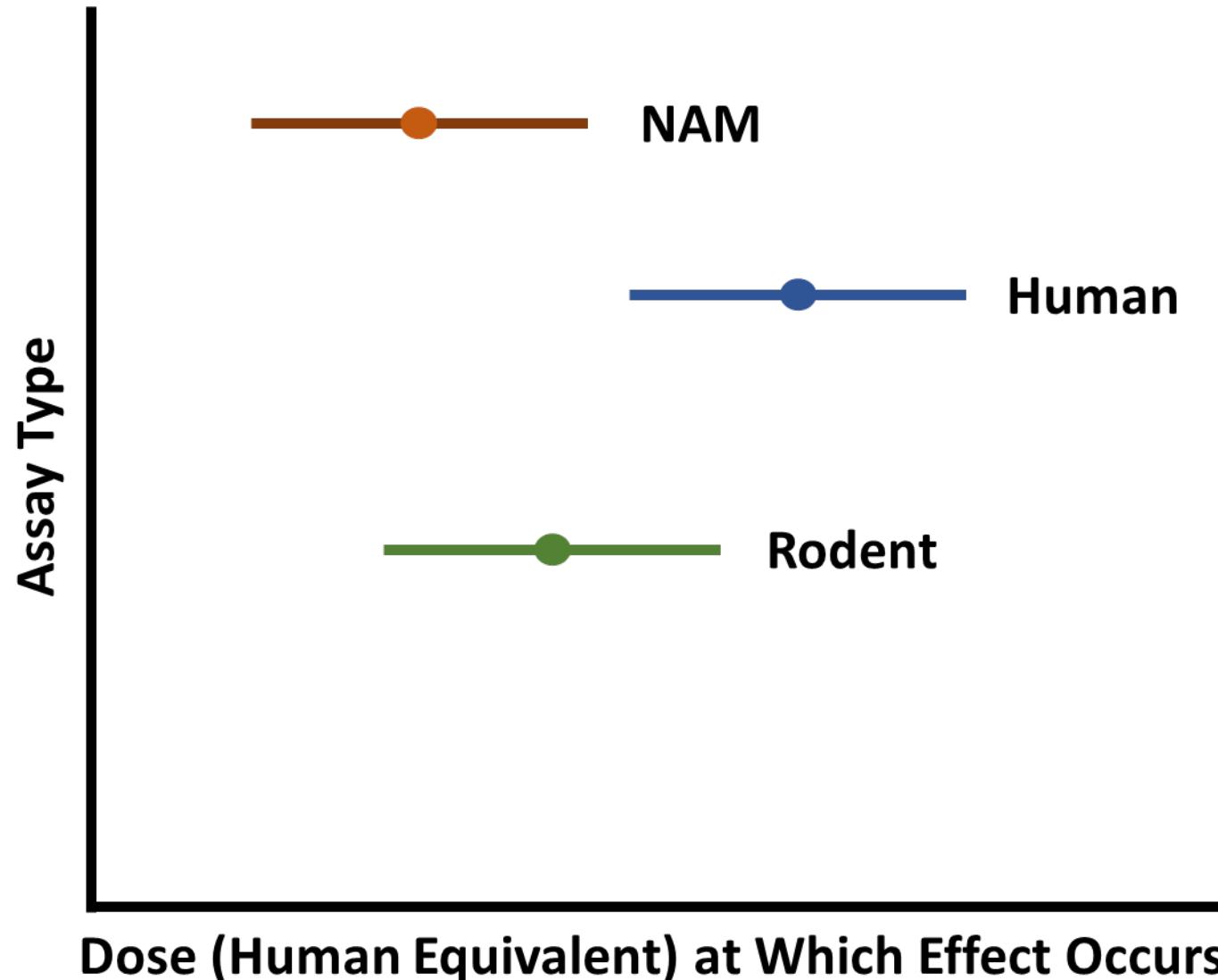
# **EPA efforts may inform scientific support elements that relate to reliability and relevance**

- In what contexts do we need to require inter-laboratory validation versus ensuring the method is standardized?
- How does the human relevance of the new approach compare to the “gold standard” or traditional test (if a comparator assay exists)?
- Is the assay result reliable when compared to repeated studies using traditional approaches?
- How do we contextualize a result from a new method, particularly if the result has more uncertainty than the traditional approach?

# Relevance: Cross-species concordance



# Relevance: Cross-species concordance



However, because NAMs are often MOA-based, the sensitivity may be greater, which means that the dose-effect linkage is **less concordant** with human, but potentially may be **more protective** of human responses.

Most frameworks for confidence building require predictions to be “as good or better” than current models.

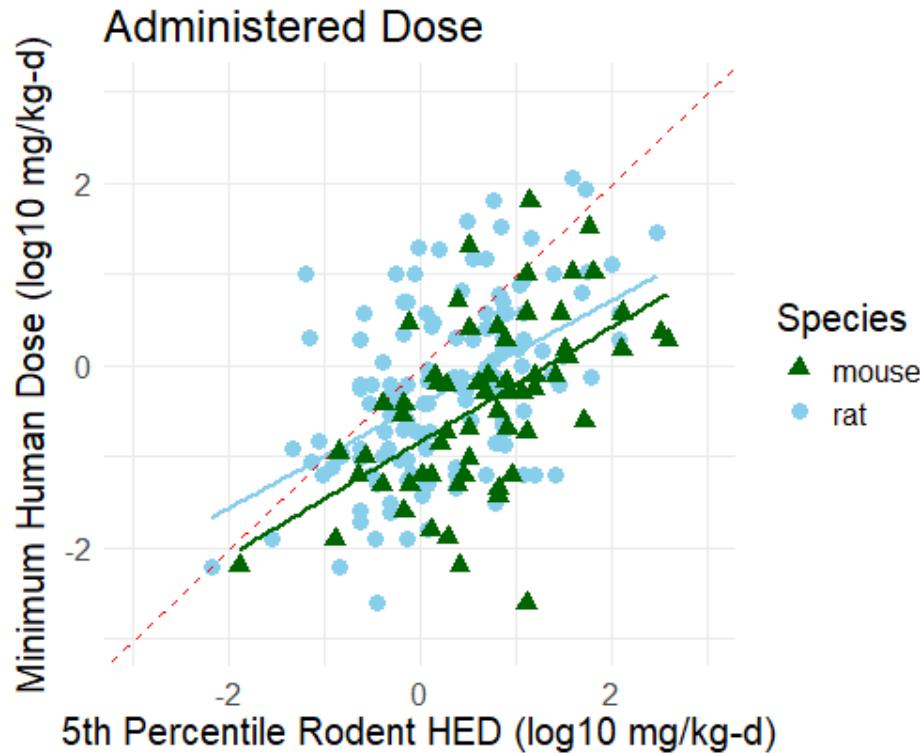
A good starting place would be to assess concordance between doses and adverse effects that occur in both humans and animal models for the same chemicals.

# Benchmarks for assessing relevance in validation studies

- We can leverage pharmaceutical data to assess benchmarks for
  - *Quantitative* concordance – dose matching across species
  - *Qualitative* concordance – hazard matching across species
- Pharmaceutical data have the benefit of providing both human (clinical) and rodent (non-clinical) dose-effect linkage data

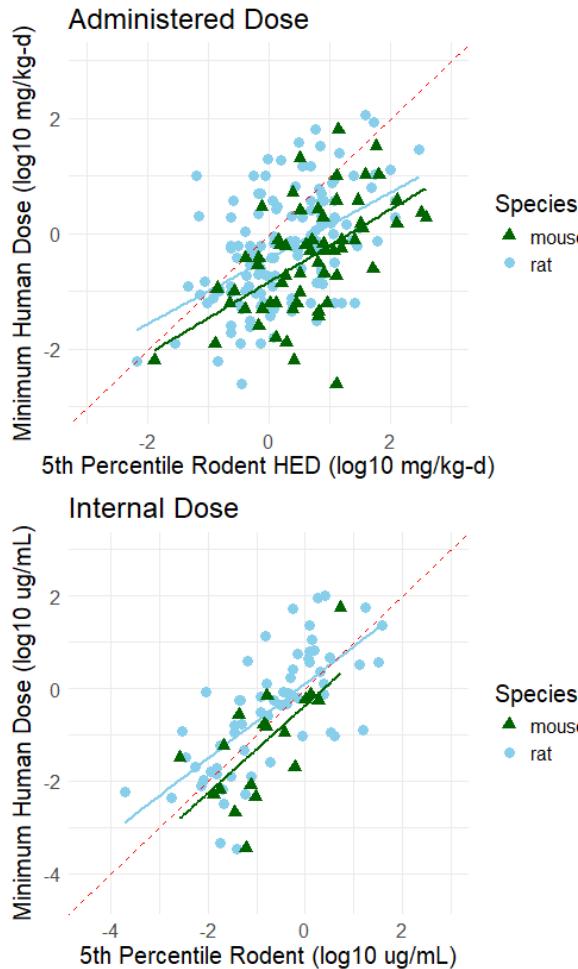


# Benchmarks for human relevance – dose concordance in pharmaceuticals



- Data derived from extracting relevant information from new drug applications (NDA) to the FDA
- Rodent doses adjusted to human equivalent dose
- Multiple studies submitted on single compounds – 5<sup>th</sup> percentile of the distribution of POD values for each species used as a conservative lower bound estimate
- Human administered dose associated with adverse event (AE) is not strongly concordant with mouse or rat, with mouse potentially more protective than rat

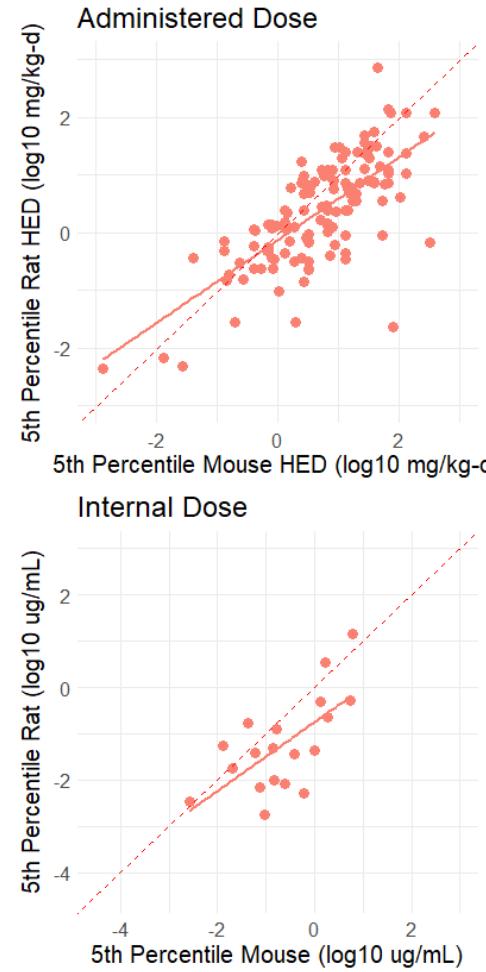
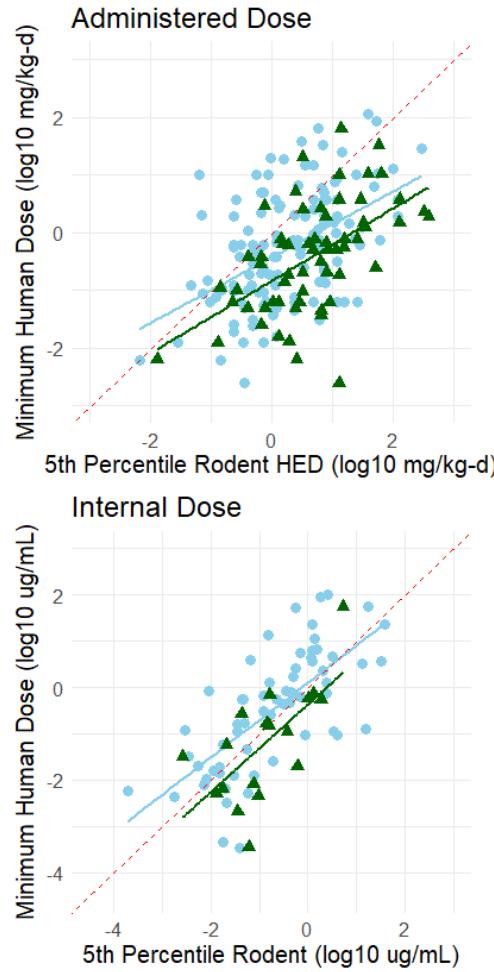
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- Human administered dose associated with adverse event (AE) is not strongly concordant with mouse or rat, with mouse potentially more protective than rat
- For drugs where we have internal dose (PK) information – internal dose for human associated with AE is more concordant with rodent

# Benchmarks for human relevance – dose concordance in pharmaceuticals



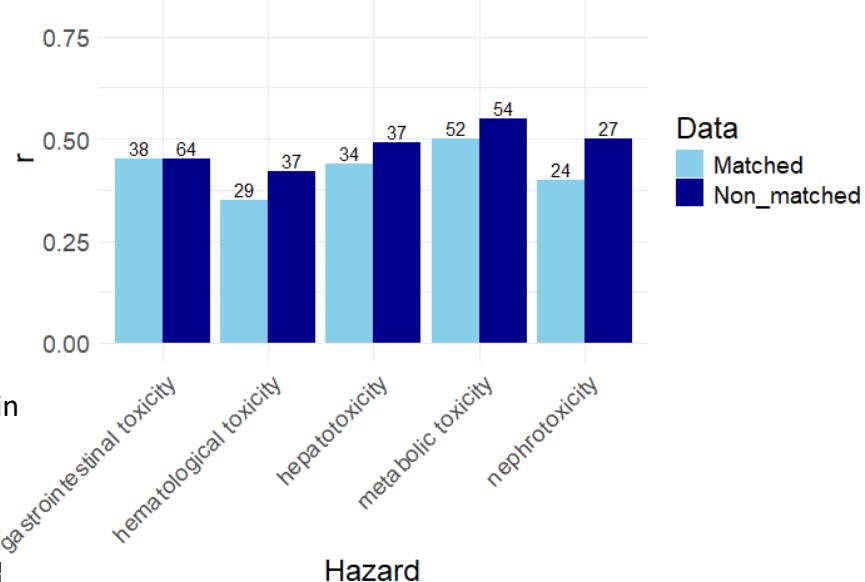
- Rat tends to be more concordant with mouse than either rodent species is with human
- ToxCast assay-> human is similar to human->rodent
- Provides a starting point for assessing where dose concordance should fall for NAM->human concordance

	Correlation statistics				Regression statistics		
	N	r	MAD	Mean bias	RMSD	R <sup>2</sup>	RMSE
<b>Comparison with rodent human equivalent dose effect levels</b>							
Human – Rat	134	0.49	0.85	-0.52	1.1	0.24	0.84
Human –	61	0.56	1.13	-1.1	1.4	0.31	0.79
Mouse							
Rat – Mouse	133	0.72	0.57	-0.35	0.71	0.52	0.6
<b>Comparison with internal dose adjusted effect levels</b>							
Human – Rat	64	0.7	0.79	0.26	0.97	0.49	0.91
Human –	18	0.67	0.75	-0.31	0.94	0.44	0.89
Mouse							
Rat – Mouse	19	0.65	0.8	-0.57	0.98	0.43	0.75

# Benchmarks for human relevance – qualitative concordance for particular adverse events

- Nonclinical and clinical trial data have been analyzed from the perspective of *qualitative* concordance (presence/absence of effects)
- Monticello *et al.* 2016
  - Low PPV (~30%) but high NPV (~86%)

Predictive concordance: NDA



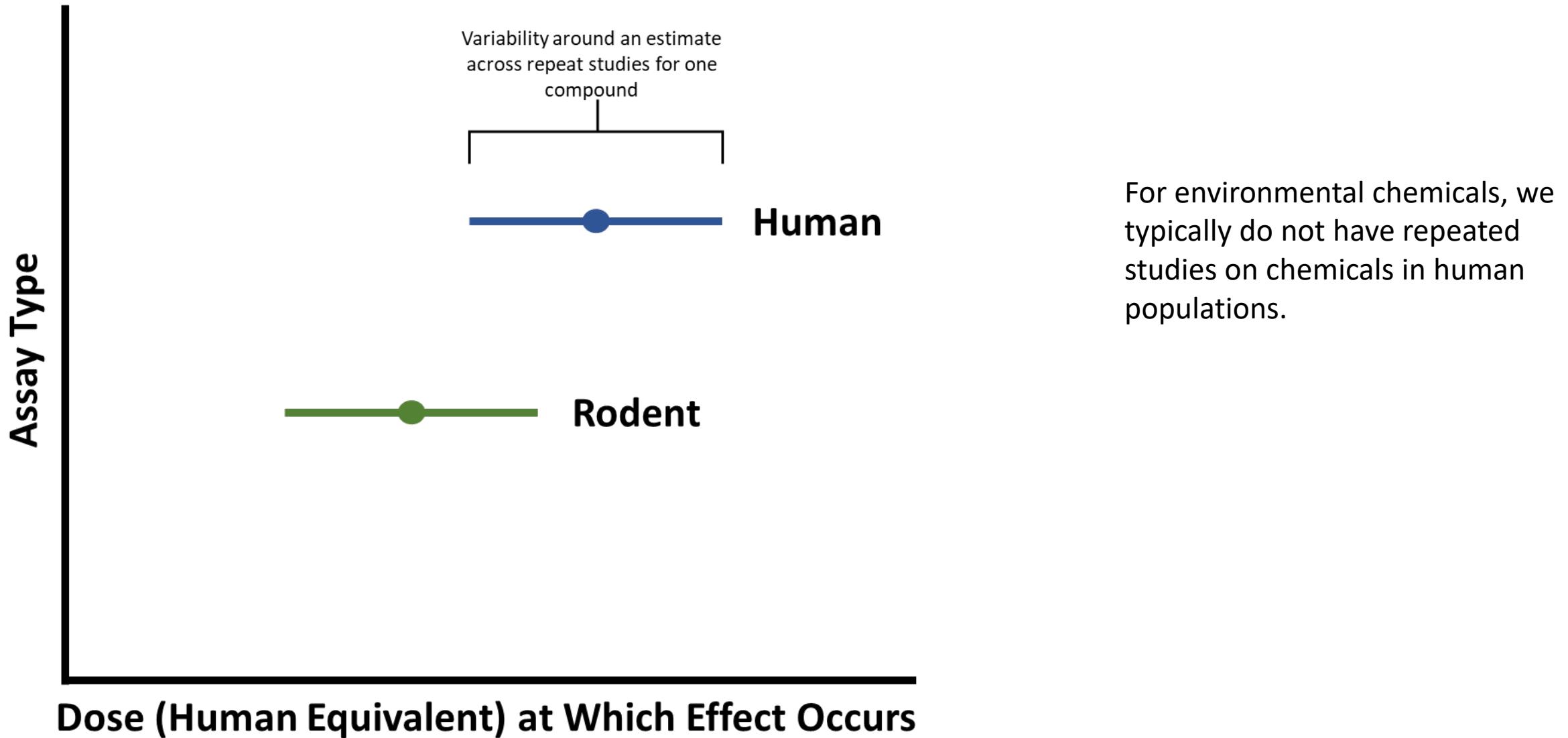
Current nonclinical testing paradigm enables safe entry to First-In-Human clinical trials: The IQ consortium nonclinical to clinical translational database

Thomas M. Monticello<sup>a,\*</sup>, Thomas W. Jones<sup>b</sup>, Donna M. Dambach<sup>c</sup>, David M. Potter<sup>d</sup>, Michael W. Bolt<sup>e</sup>, Maggie Liu<sup>f</sup>, Douglas A. Keller<sup>g</sup>, Timothy K. Hart<sup>h</sup>, Vivek J. Kadambi<sup>i</sup>

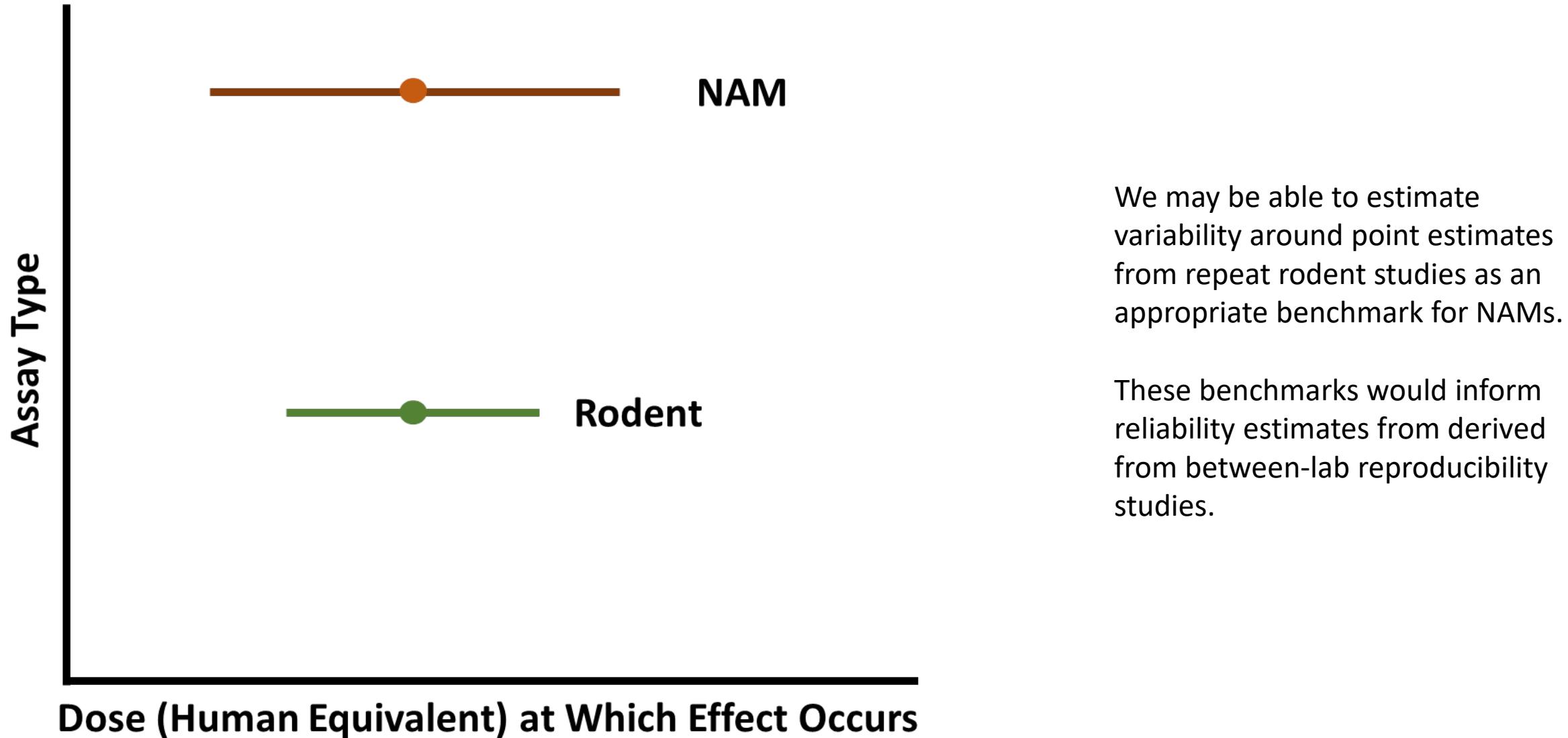
There is a low concordance between rodent and human both when the effects are matched (hepatohepatic) and when the effects are unmatched (hepato-any other adverse effect).

We can use these concordance estimates as potential benchmarks when assessing expectations for *in vitro* systems to predict effects in the human population.

# Reliability: What is an acceptable amount of variability around the assay estimate across studies?



# Reliability: Benchmarks for the amount of variability around the assay estimate



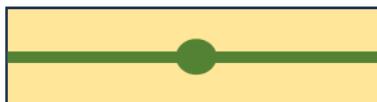
# Reliability: Benchmarks for the amount of variability around the assay estimate

## Assay Type



**NAM**

May inform what this variability should ideally be



**Rodent**

Understanding what this variability normally is

What is an expected level of variance around a point estimate?

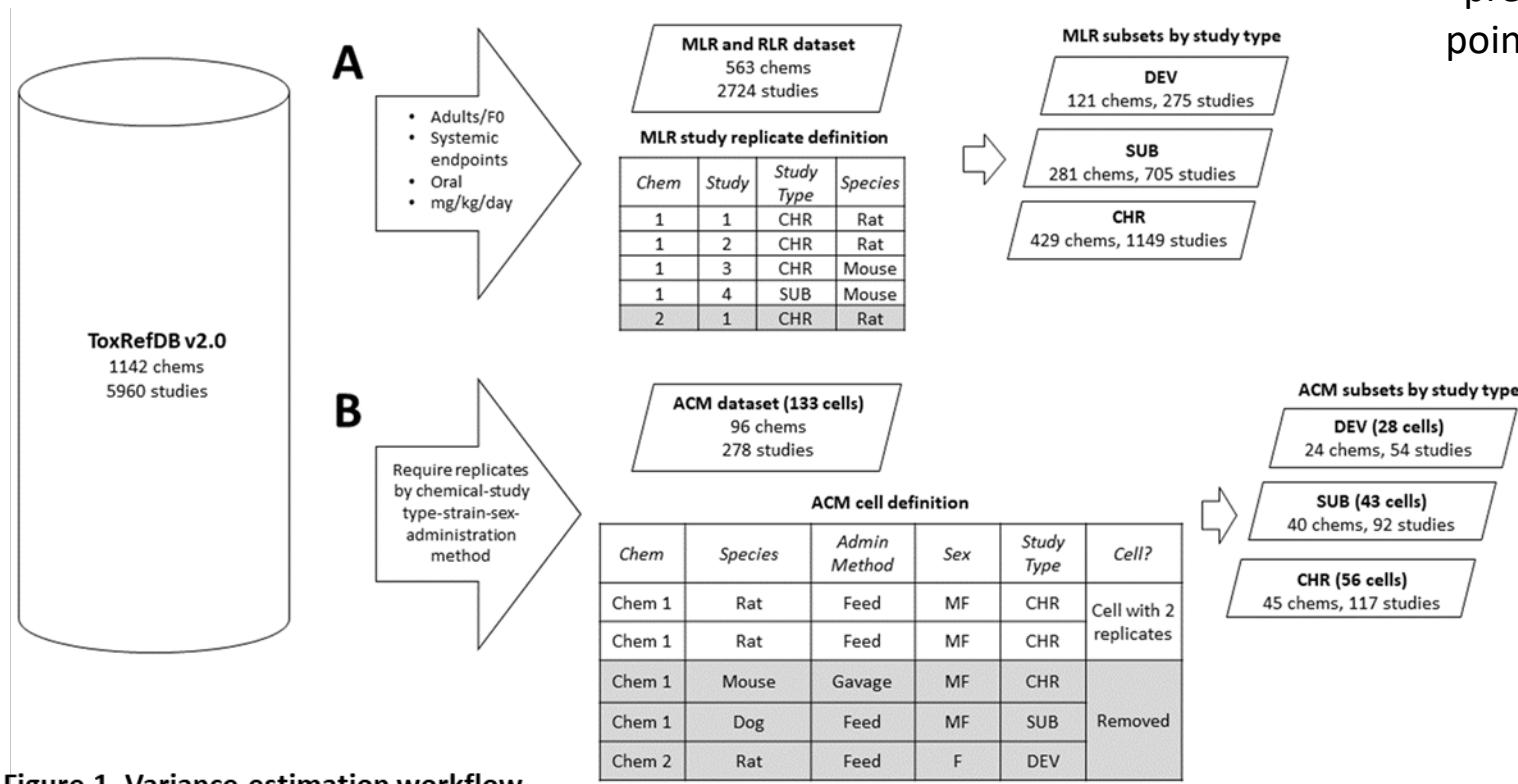
A potential benchmark is to examine variability around a POD using estimates from multiple animal studies on a chemical as a measure of reliability.

To do this, we can leverage ToxValDB, EPA's largest repository of published in-life study data.

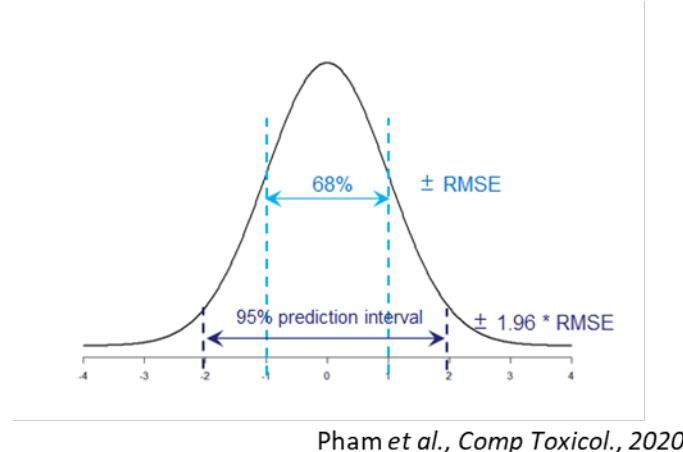
**Dose (Human Equivalent) at Which Effect Occurs**

# Reliability: Benchmarks for variability around the POD in repeated studies

28 different statistical models to approximate total variance, unexplained variance, and the spread of the residuals from statistical models of study-level points-of-departure in adult animals.



The variance, as approximated by RMSE, is **0.45-0.56 log<sub>10</sub>-mg/kg-bw/day**. This helps us estimate a minimum prediction interval for a new estimation of study-level point-of-departure and to set a benchmark for NAMs to predict these values.



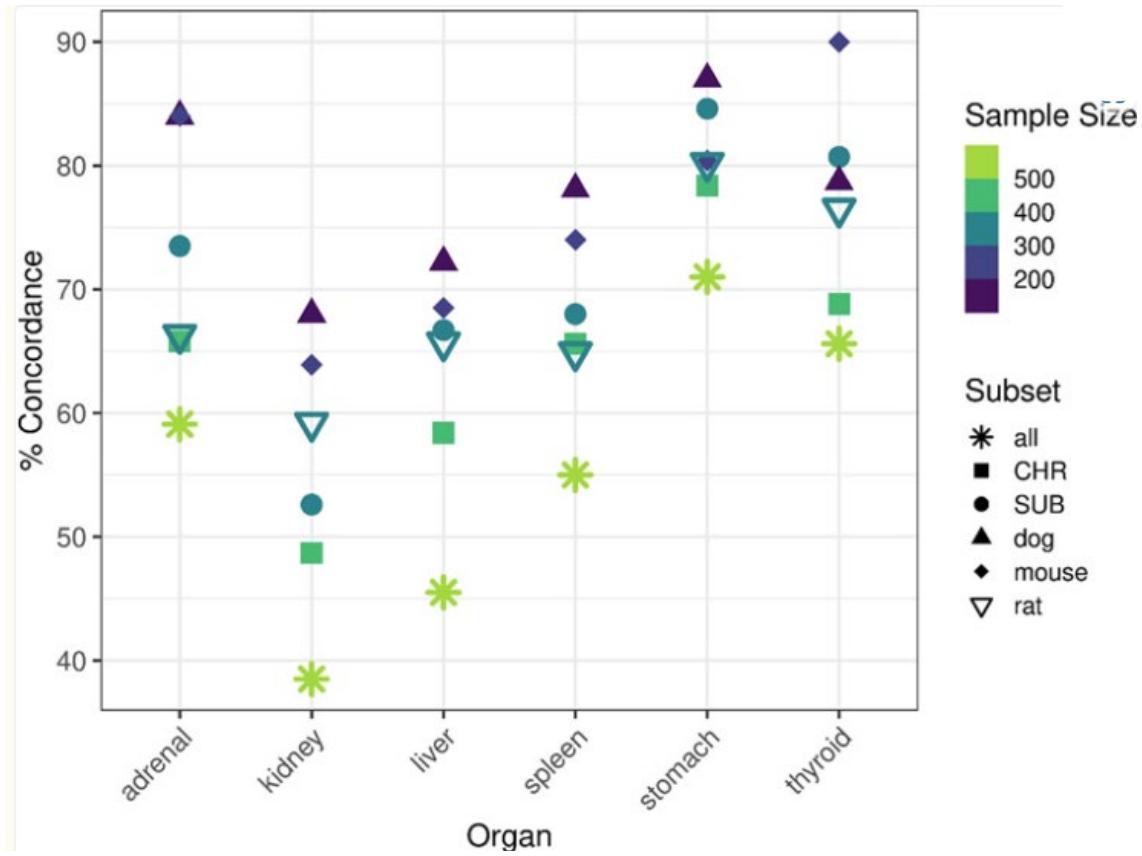
Pham et al., Comp Toxicol., 2020

Figure 1. Variance estimation workflow.

CHR = chronic; DEV = developmental (adults only); SUB = subchronic; cells are defined by the factor of all categorical variables; MF = males and females; F = females; MLR = multilinear regression; POD = point of departure; RLR = robust linear regression; ACM = augmented cell means.

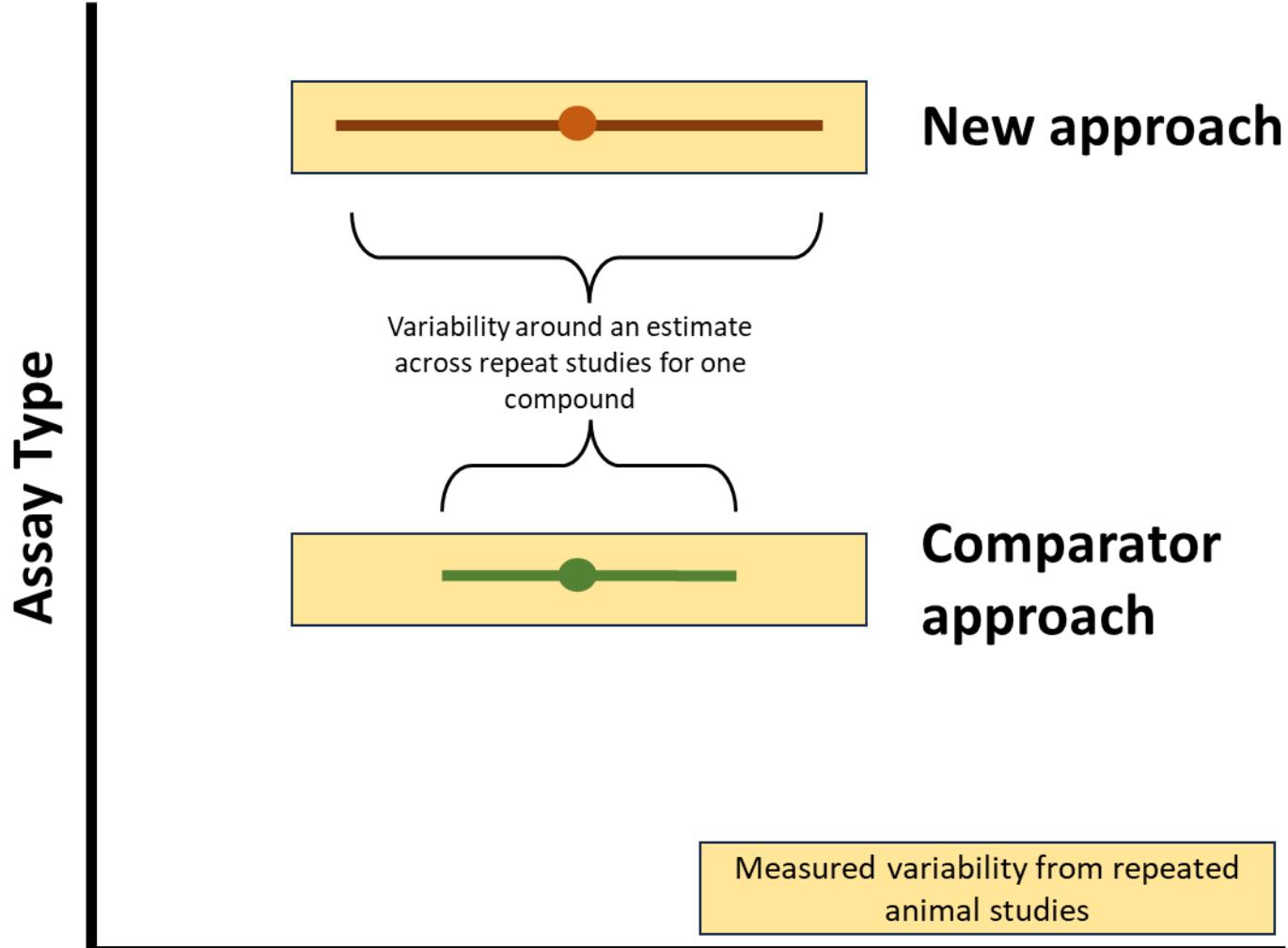
# Reproducibility of organ-level effects in repeat dose animal studies

Qualitative reproducibility of organ-level effect observations  
in repeat dose studies of adult animals



- Data for 538 chemicals across 2284 studies
- Study replicates considered at the chemical level
- Liver and kidney were associated with the greatest number of studies with positive reporting, yet had the lowest concordance of findings
- Can be used as potential benchmark for qualitative concordance of endpoint effects

# Reliability: Contextualizing Variability of a New Method



If reproducibility in the new method is within the limits of historical in vivo data, but the variability of the new method is still greater than a comparator test, is there value in using the new approach?

Value of Information (VOI) can be used as a decision-making framework to contextualize trade-offs in uncertainty around the POD, costs, and time to decision between a choice of methods than can lead to informing a decision.

**Dose (Human Equivalent) at Which Effect Occurs**

# Value of Information: EPA-developed framework

DOI: 10.1111/rina.13931

ORIGINAL ARTICLE

## A value of information framework for assessing the trade-offs associated with uncertainty, duration, and cost of chemical toxicity testing

Shintaro Hagiwara<sup>1,2</sup> | Greg M. Paoli<sup>1</sup> | Paul S. Price<sup>3</sup> | Maureen R. Gwinn<sup>4</sup> | Annette Guiseppi-Elie<sup>3</sup> | Patrick J. Farrell<sup>2</sup> | Bryan J. Hubbell<sup>5</sup> | Daniel Krewski<sup>1,6</sup> | Russell S. Thomas<sup>3</sup>

<sup>1</sup>Risk Sciences International, Ottawa, Canada  
<sup>2</sup>School of Mathematics and Statistics, Carleton University, Ottawa, Canada  
<sup>3</sup>Center for Computational Toxicology and Exposure, Office of Research and Development, US Environmental Protection Agency, Research Triangle Park, North Carolina, USA  
<sup>4</sup>Office of Research and Development, US Environmental Protection Agency, Research Triangle Park, North Carolina, USA  
<sup>5</sup>Air, Climate, and Energy Research Program, Office of Research and Development, US Environmental Protection Agency, Research Triangle Park, North Carolina, USA  
<sup>6</sup>McLaughlin Centre for Population Health Risk Assessment, University of Ottawa, Ottawa, Canada

**Abstract**  
A number of investigators have explored the use of value of information (VOI) analysis to evaluate alternative information collection procedures in diverse decision-making contexts. This paper presents an analytic framework for determining the value of toxicity information used in risk-based decision making. The framework is specifically designed to explore the trade-offs between cost, timeliness, and uncertainty reduction associated with different toxicity-testing methodologies. The use of the proposed framework is demonstrated by two illustrative applications which, although based on simplified assumptions, show the insights that can be obtained through the use of VOI analysis. Specifically, these results suggest that timeliness of information collection has a significant impact on estimates of the VOI of chemical toxicity tests, even in the presence of smaller reductions in uncertainty. The framework introduces the concept of the expected value of delayed sample information, as an extension to the usual expected value of sample information, to accommodate the reductions in value resulting from delayed decision making. Our analysis also suggests that lower cost and higher throughput testing also may be beneficial in terms of public health benefits by increasing the number of substances that can be evaluated within a given budget. When the relative value is expressed in terms of return-on-investment per testing strategy, the differences can be substantial.

**KEY WORDS**  
cost of delay, return on investment, risk decision making, social cost, toxicity testing, value of information

### 1 | INTRODUCTION

Evidence-based risk assessment has become a cornerstone of public and population health risk decision making, integrating evidence on toxicity and exposure from multiple evidence streams. When the available evidence is insufficient to allow a decision to be made with confidence, consideration can be given to gathering additional evidence to strengthen the evidence base. The present paper focuses on the use of value of information (VOI) analysis to evaluate the utility of gathering additional evidence on the toxicity of chemicals. Specifically, we present a VOI analytic framework that builds on previous methodological work in this field, explicitly incorporating the value of additional test data resulting from reductions in the uncertainty in estimates of a chemical's toxicity, the cost of delay in decision making that results

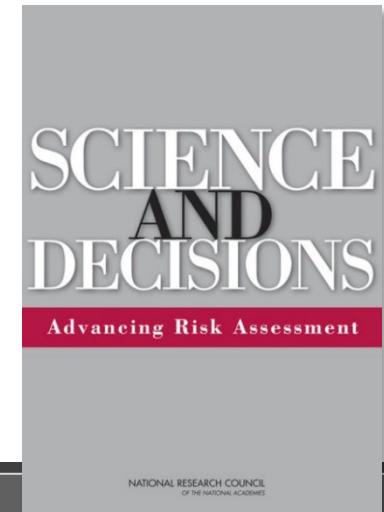
This article is licensed under the terms of the [Creative Commons Attribution](#) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.  
© 2023 Risk Sciences International. *Risk Analysis* published by Wiley Periodicals LLC on behalf of Society for Risk Analysis. This article has been contributed to by U.S. Government employees and their work is in the public domain in the USA.

*Risk Analysis*, 2023, 1–18. [wileyonlinelibrary.com/journal/rina](http://wileyonlinelibrary.com/journal/rina) | 1

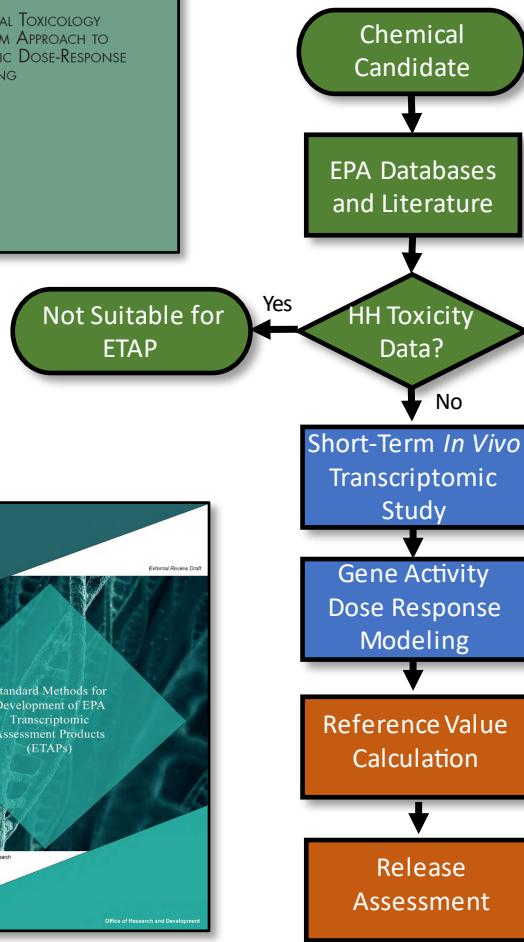
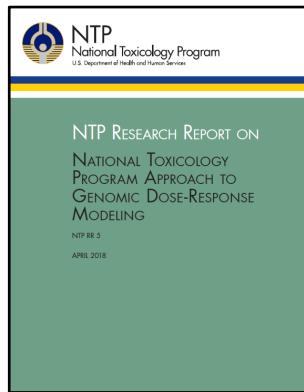
Utilize the EPA-developed VOI framework that is ground-breaking because it explicitly considers the impact of delay in decision-making.

The framework takes into account:

- Amount of **uncertainty** reduced
- **Cost** of additional toxicity testing
- **Delay** in obtaining and evaluating toxicity testing data



# Case Study: Value of Information associated with ETAP



The VOI analysis in this study aimed to answer the following question: ***given that additional toxicity testing data may be beneficial, which toxicity testing methodology and assessment process provides the most value?***

Case study compared chronic 2-year rodent toxicity test & assessment to shorter-duration transcriptomic study (ETAP)

Variability around the POD for ETAP was within estimates from Pham *et al.* 2020.

	Transcriptomics Study and Human Health Assessment	Traditional Toxicity Testing and Human Health Assessment
Time Required	<1 year	8 years
Quantitative uncertainty	Modestly greater	Modestly less
Costs	~\$200,000	~\$4 million

# VOI considers socioeconomic factors and public health benefit to assess return on investment

- **Not testing a chemical may have a cost** borne by the public in terms of healthcare costs arising from exposure to a chemical
  - Economists think in terms of annualized health costs for a variety of outcomes, in terms of healthcare costs, lost productivity, and direct non-medical costs such as education or transportation
  - Annual economic values for a variety of conditions have been estimated
    - Ex: autism spectrum disorder (\$69,530/year), asthma (\$36,500/yr), pervasive developmental disorders (\$10,538/yr), EPA economic guidance estimates fatality at \$110,000/yr, considering a value of statistical life (VSL) of \$8.8 mil and an 80-year life span
- **Delay has a cost** – Annualized healthcare costs accumulate over time if the exposure is not mitigated and are multiplicative based on the size of the affected population
  - 100,000 people exposed for 5 years prior to mitigation with a \$10k annual healthcare cost (total health cost is \$5 billion)
    - Mitigating exposure after 2 years saves the public \$3 billion
  - For VOI, we consider a time horizon over which benefits of a particular testing strategy may be realized, economists typically use a 20-year time horizon

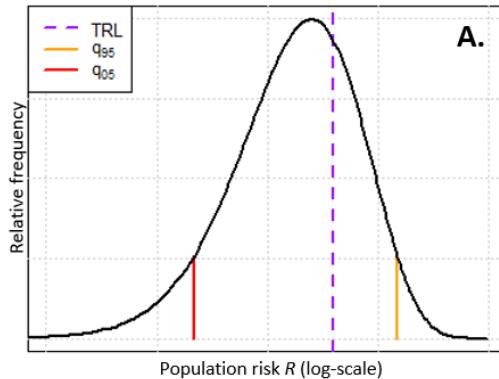
# Additional socioeconomic cost to consider

- **Another cost to be considered once a regulatory action is finalized – cost of control**
  - Variety of actions that can be taken – ex. reducing emissions, incorporating water treatment/purification modalities, excavating and moving soil, substituting one chemical in a product formulation for an alternative
  - Under REACH (2021), annualized control costs had a mean of \$50.6M and a median of \$5.7M

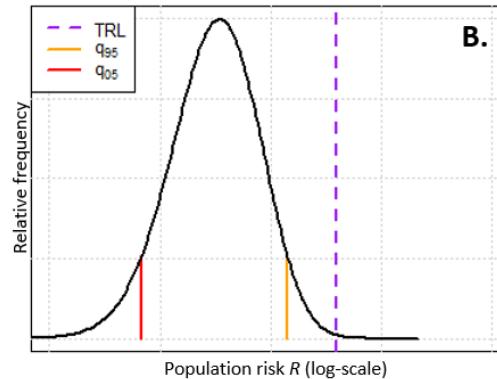


# Two idealized decision makers in case study

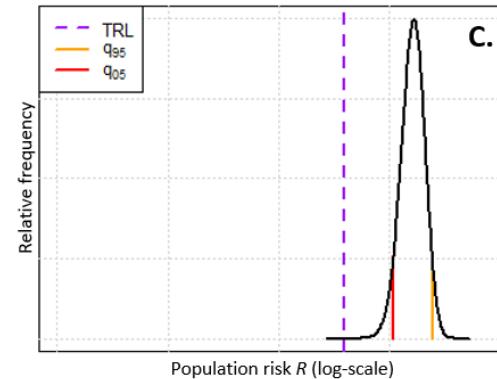
- Benefit-Risk Decision Maker (BRDM): Chooses to regulate a chemical if the reduction in health cost (or increased health benefit) outweighs the associated cost of control
- Target-Risk Decision Maker (TRDM): Chooses to regulate a chemical if the (lower quantile of) risk exceeds the pre-specified target risk level



TRDM would need additional evidence to make a decision



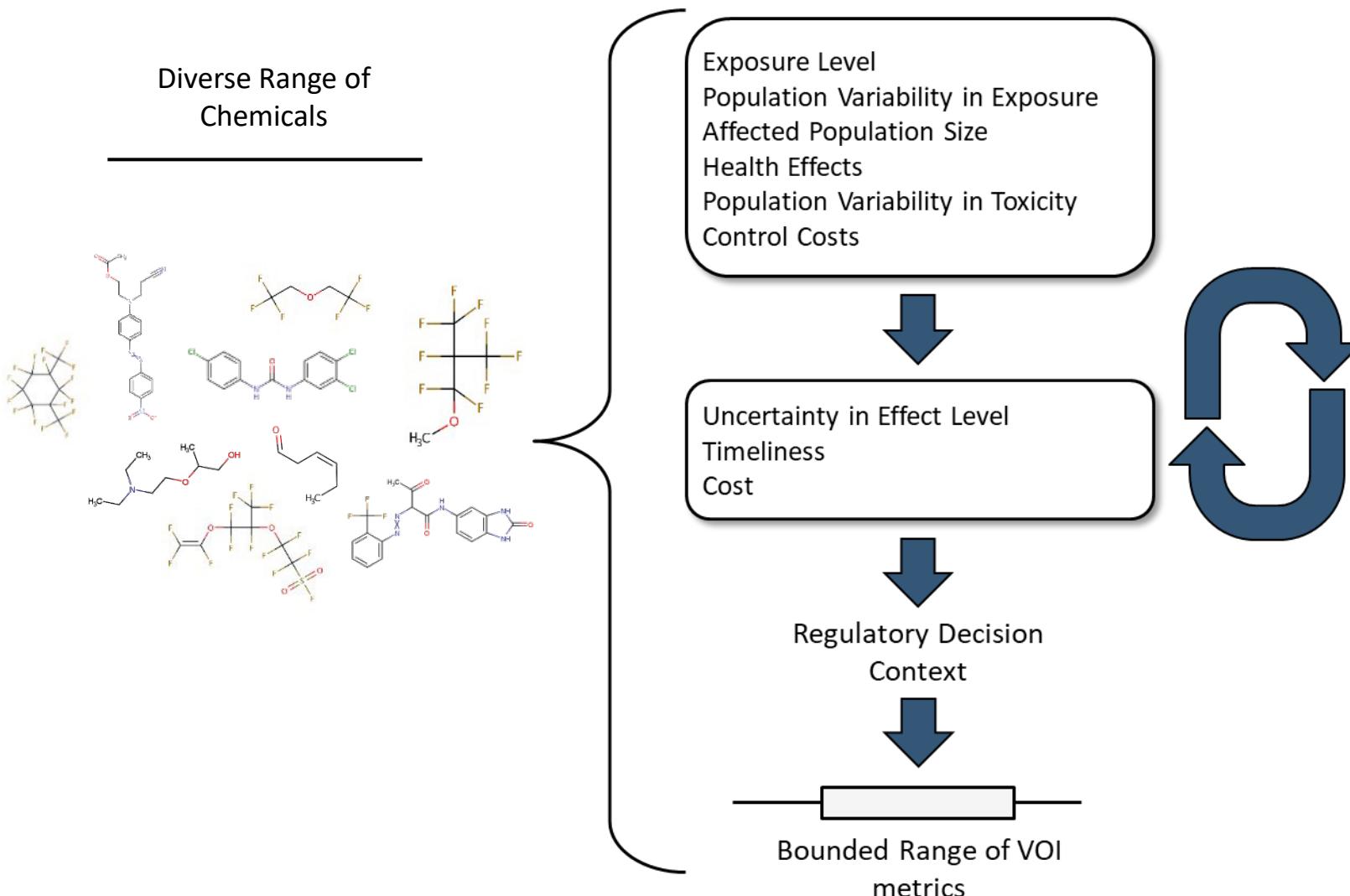
Target risk level is greater than the uncertainty distribution, no regulatory action required



Target risk level is below the 5<sup>th</sup> percentile of uncertainty distribution, regulatory action is required

TRL: Prespecified Target Risk Level

# Contextualizing reliability of two assays with VOI analysis



360 Data Driven Scenarios Examined  
Comparing ETAP vs Traditional HHA Process

- SHEDS-HT exposure tertiles
- Population sizes (US)
  - 33, 165, **330 million** (10, 50, **100%**)
- Time horizons
  - **20**, 40, 75 years
- Testing costs
  - THHA \$1M or **\$4M** (M=million)
  - ETAP **\$200K** or \$250K (K=thousand)
- Time from testing start to assessment finish
  - THHA **6, 8, 14** years
  - ETAP **0.5**, 1, 2 years
- Control costs
  - \$50M or **\$23.1B** for 25% reduction
- Annualized health costs
  - \$1K, **\$10K**, \$110K
- Discount rate: 3, **5**, 7%
- Uncertainty around the point-of-departure
  - SD about the mean for each assay from empirical measurements
  - Additional uncertainty added to ETAP

# **New approach was preferred over the traditional approach in most scenarios, despite greater uncertainty in estimates derived from repeated studies**

- The VOI Case study evaluated 360 scenarios
  - For each decision context, 9 baseline and 171 sensitivity scenarios
- Benefit-Risk Decision Maker (180 scenarios)
  - In 82% of scenarios, ETAP was preferred with favorable ROI & ENBS
  - 18% - no testing preferred
  - Average benefit was \$44 billion for BRDM
- Target-Risk Decision Maker (180 scenarios)
  - ETAP was preferred in 89% of scenarios (ENBS) and 99% of scenarios (ROI)
  - 7.2% - no testing preferred
  - Average benefit was \$81 billion for TRDM

# Conclusions

- Understanding the validation efforts to date may help inform optimization of the numbers of participating labs and reference chemicals required for building confidence in a method's reliability
- Retrospective analysis of available *in vivo* datasets allows for understanding of appropriate qualitative and quantitative benchmarks for assessing relevance and reliability of NAMs
- Value of Information frameworks can assist with contextualizing relative value in socioeconomic terms of using a more uncertain versus less uncertain assay or method
- These efforts inform updates to confidence building frameworks, including the GD 34 revision

# Thank you

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Scott Auerbach, Tara Barton Maclaren, Suzy Fitzpatrick, Alexandra Long, Anna Lowit, Roman Mezencev, Monique Perron, Pilar Prieto-Peraita, Mike Rasenberg, Michael Santillo, Ulla Simanainen, Doris Smith, Maurice Whelan, Paul White, Krystle Yozzo  
ECHA, FDA, Health Canada, EU-JRC





# Development and Application of Exposure New Approach Methodologies in EPA's ExpoCast Project



*Kristin Isaacs*

*Center for Computational Toxicology and Exposure  
Office of Research and Development  
United States Environmental Protection Agency*



*4<sup>th</sup> US EPA Conference on the State of  
Science on Development and Use of  
NAMs for Chemical Safety Testing  
November 5, 2024*

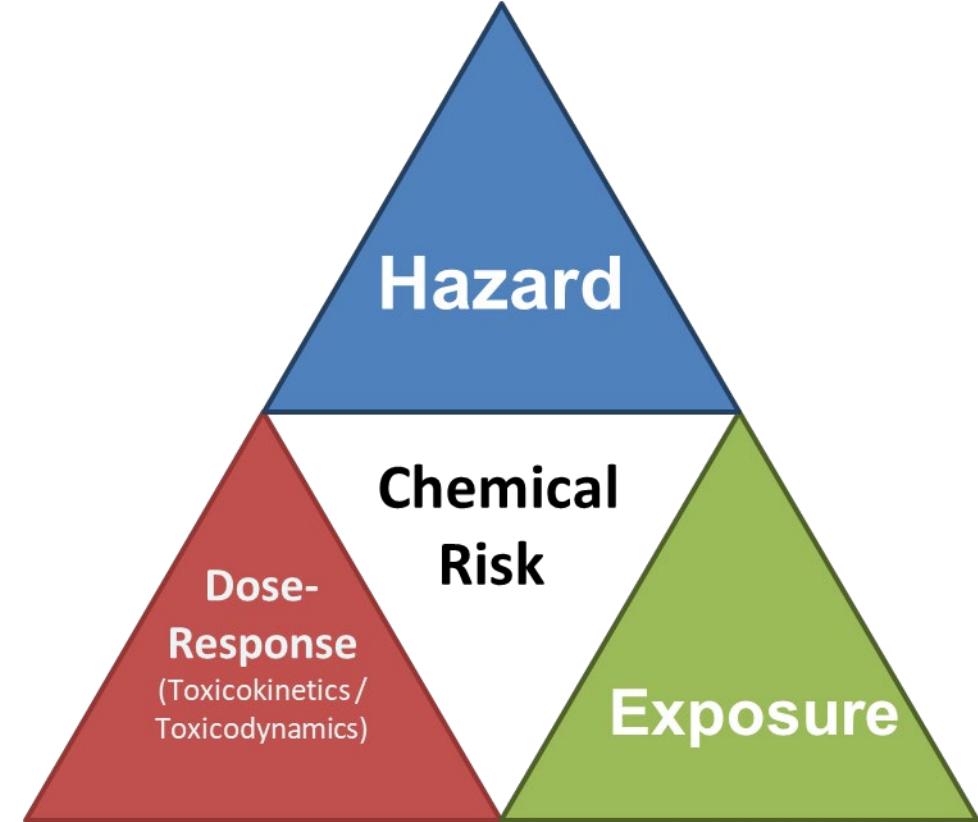
## Disclaimer

The views expressed in this presentation are those of the authors and do not necessarily reflect the views or policies of the U.S. EPA.

**USING  
21ST CENTURY  
SCIENCE  
TO IMPROVE  
RISK-RELATED  
EVALUATIONS**

NASEM (2017)

“Recent advances in high throughput toxicity assessment, notably the **ToxCast and Tox21** programs... and in high throughput computational exposure assessment [**ExpoCast**] have enabled first-tier risk-based rankings of chemicals on the basis of **margins of exposure**”



# NAMs and Chemical Risk

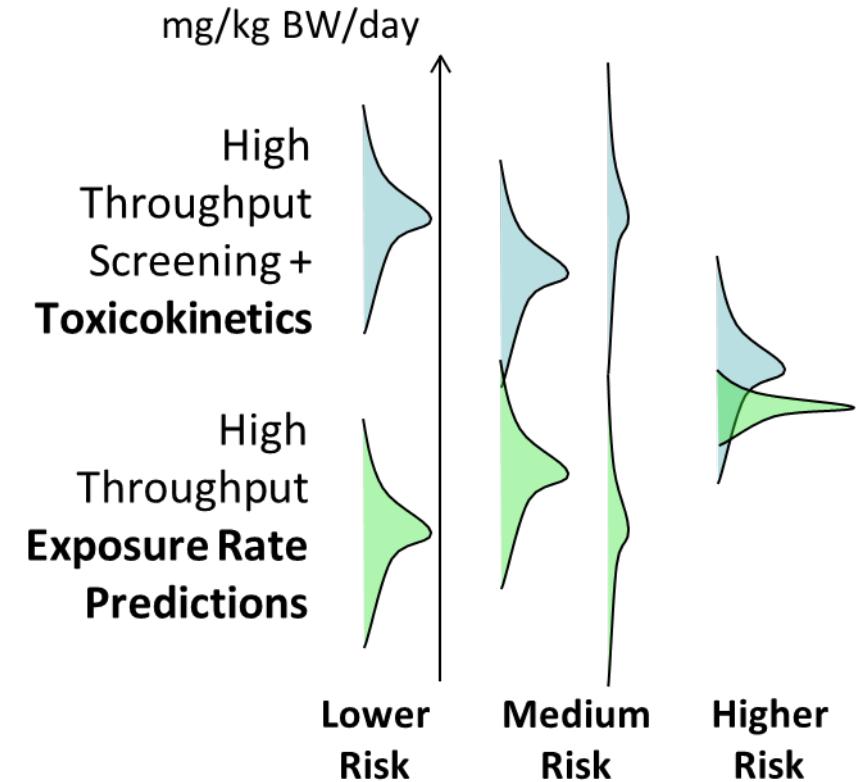
## USING 21ST CENTURY SCIENCE TO IMPROVE RISK-RELATED EVALUATIONS

NASEM (2017)

“Recent advances in high throughput toxicity assessment, notably the **ToxCast and Tox21** programs... and in high throughput computational exposure assessment [**ExpoCast**] have enabled first-tier risk-based rankings of chemicals on the basis of **margins of exposure**”

*Exposure that results in effect*

*Exposure that actually occurs*

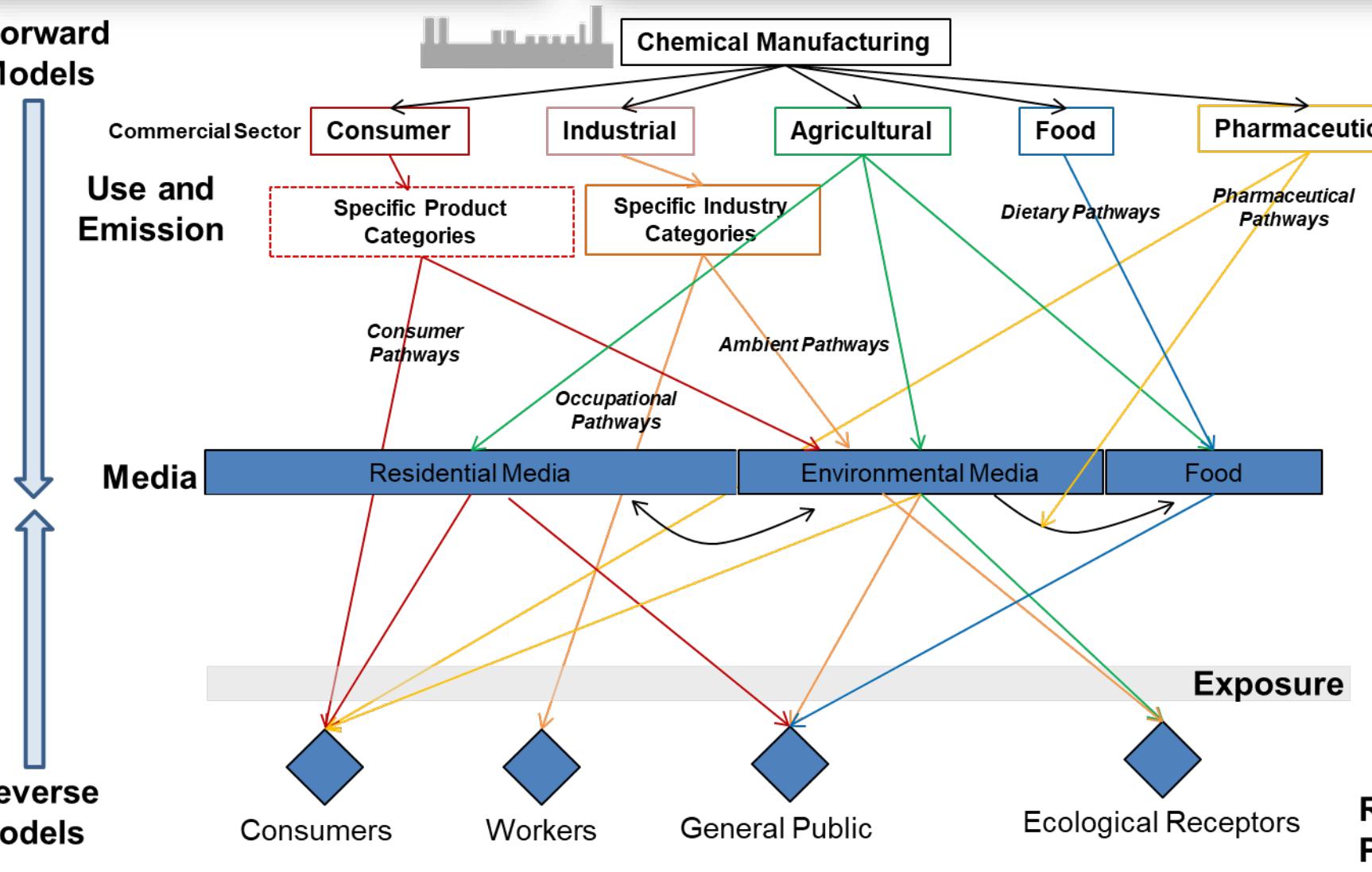


**New approach methodologies (NAMs)** enable risk assessors to more rapidly address public health challenges and chemical regulation

# Exposure Pathways from Source to Receptor



## Forward Models



## Critical Exposure-Relevant Domains

- **Chemical use and emission.** Provides critical information for identifying chemical sources, exposure pathways, and relevant models for a given chemical.
- **Media occurrence, environmental surveillance, and biomonitoring.** Provides exposure data for evaluating predictive models.
- **Toxicokinetics.** Provides real-world exposure context to *in vitro* high-throughput screening data and biological receptor monitoring information.

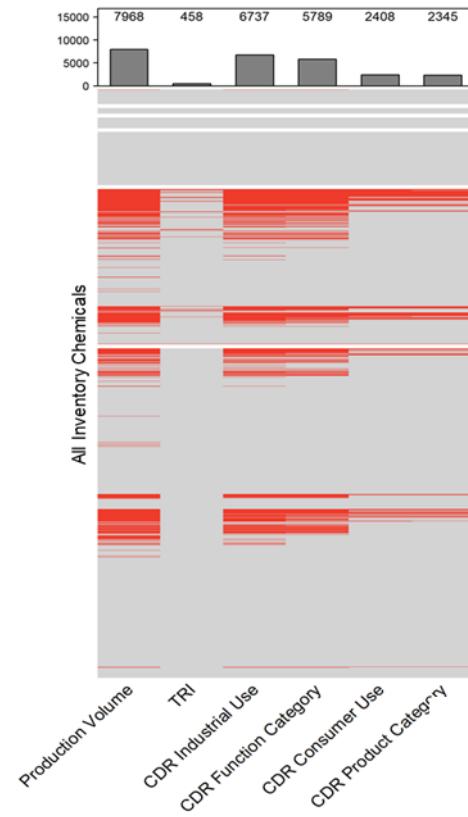
# Traditional Exposure Data are Scarce

Accelerating the Pace of Chemical Risk Assessment

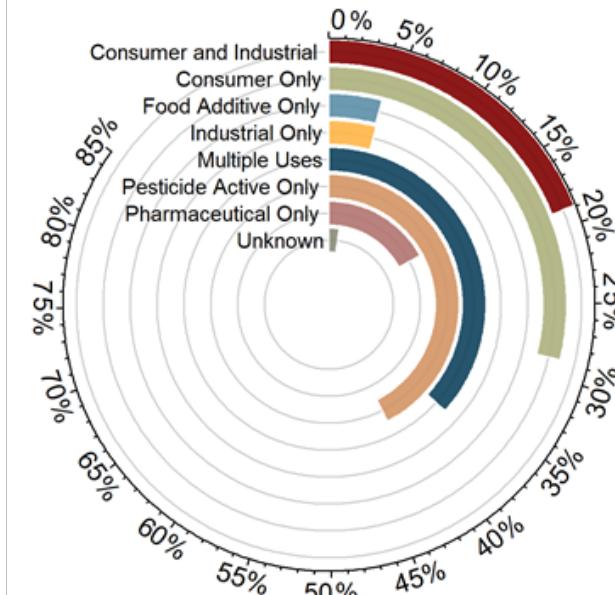


- Examined combined list of 38,715 chemicals from government regulatory inventories

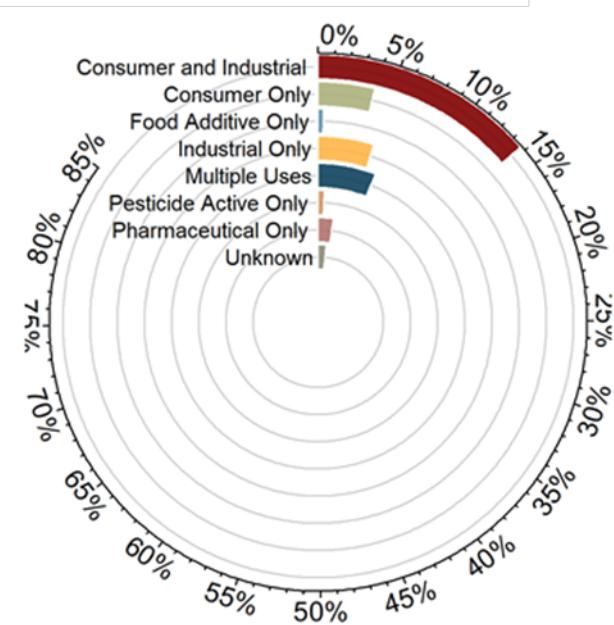
Available Use and Emission Data



Percent of Commercial Sector with Traditional Monitoring Data



Percent of Commercial Sector with Exposure Estimates



Isaacs et al., J Exp. Sci. Env. Epidem. (2022)

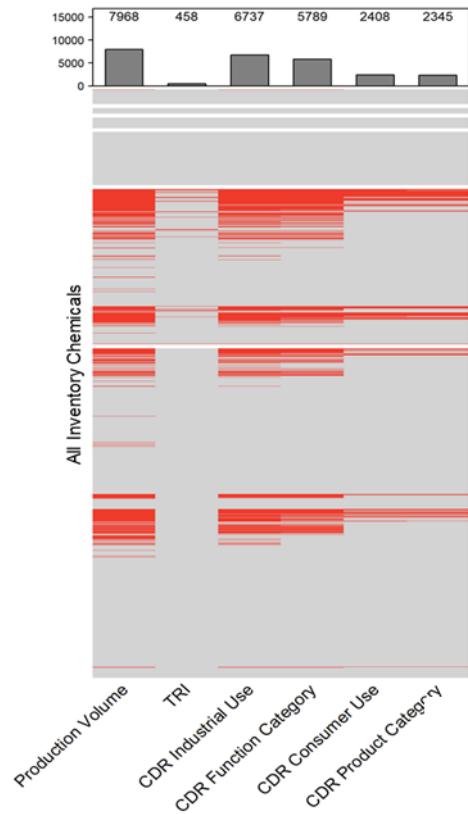
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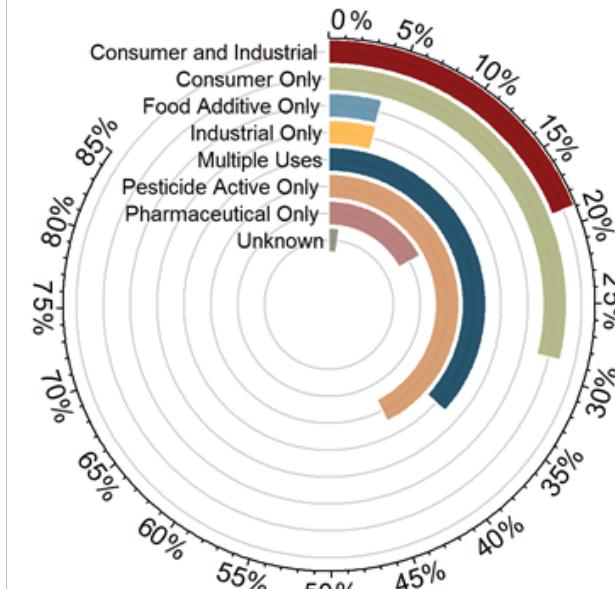


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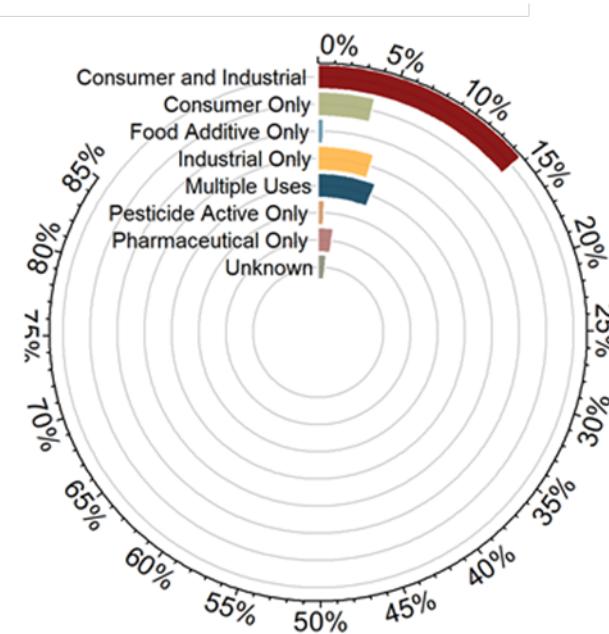
Available Use and Emission Data



Percent of Commercial Sector with Traditional Monitoring Data



Percent of Commercial Sector with Exposure Estimates



Isaacs et al., J Exp. Sci. Env. Epidemiol. (2022)

*The ExpoCast project and its collaborators are working to fill gaps in exposure data for 1000s of chemicals using high-throughput **new approach methods (NAMs)** for exposure*



ELSEVIER

## Current Opinion in Toxicology

Available online 31 July 2019

In Press, Journal Pre-proof 



## New Approach Methodologies for Exposure Science

John F. Wambaugh<sup>1</sup> , Jane C. Bare<sup>2</sup>, Courtney C. Carignan<sup>3</sup>, Kathie L. Dionisio<sup>4</sup>, Robin E. Dodson<sup>5, 6</sup>, Olivier Jolliet<sup>7</sup>, Xiaoyu Liu<sup>8</sup>, David E. Meyer<sup>2</sup>, Seth R. Newton<sup>4</sup>, Katherine A. Phillips<sup>4</sup>, Paul S. Price<sup>4</sup>, Caroline L. Ring<sup>9</sup>, Hyeong-Moo Shin<sup>10</sup>, Jon R. Sobus<sup>4</sup>, Tamara Tal<sup>11</sup>, Elin M. Ulrich<sup>4</sup>, Daniel A. Vallero<sup>4</sup>, Barbara A. Wetmore<sup>4</sup>, Kristin K. Isaacs<sup>4</sup>

***Defined 7 classes of Exposure NAMs oriented toward high-throughput application: suitable for dealing with the thousands of chemicals in commerce with limited sources of chemical exposure information***

# Exposure NAMs in the ExpoCast Paradigm

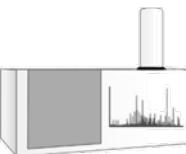
**Chemical descriptors** that provide information on chemicals in an exposure context (e.g., how chemicals are used or released)



**Machine-learning approaches** that use these descriptors to fill gaps in existing data

**Collect**  
relevant data

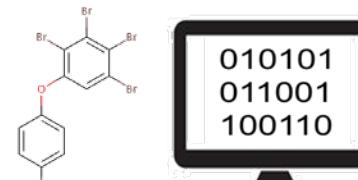
**High-throughput measurements** to fill gaps in monitoring data



**High-throughput toxicokinetics** approaches for measuring and predicting chemical fate *in vivo*

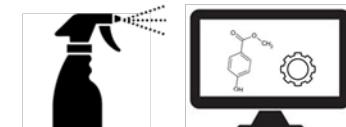


**Extrapolate**  
to fill data gaps

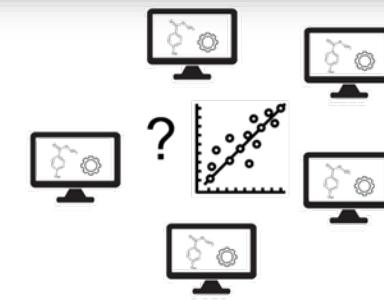


**High-throughput exposure models** for various exposure pathways

**Predict**  
exposures for specific pathways



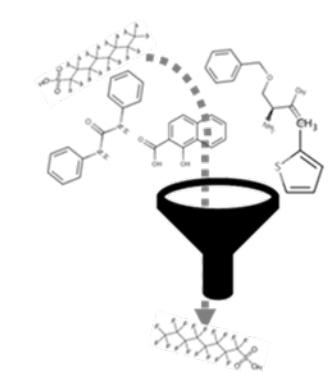
**Forecast**  
exposures for 1000s of chemicals using consensus approaches



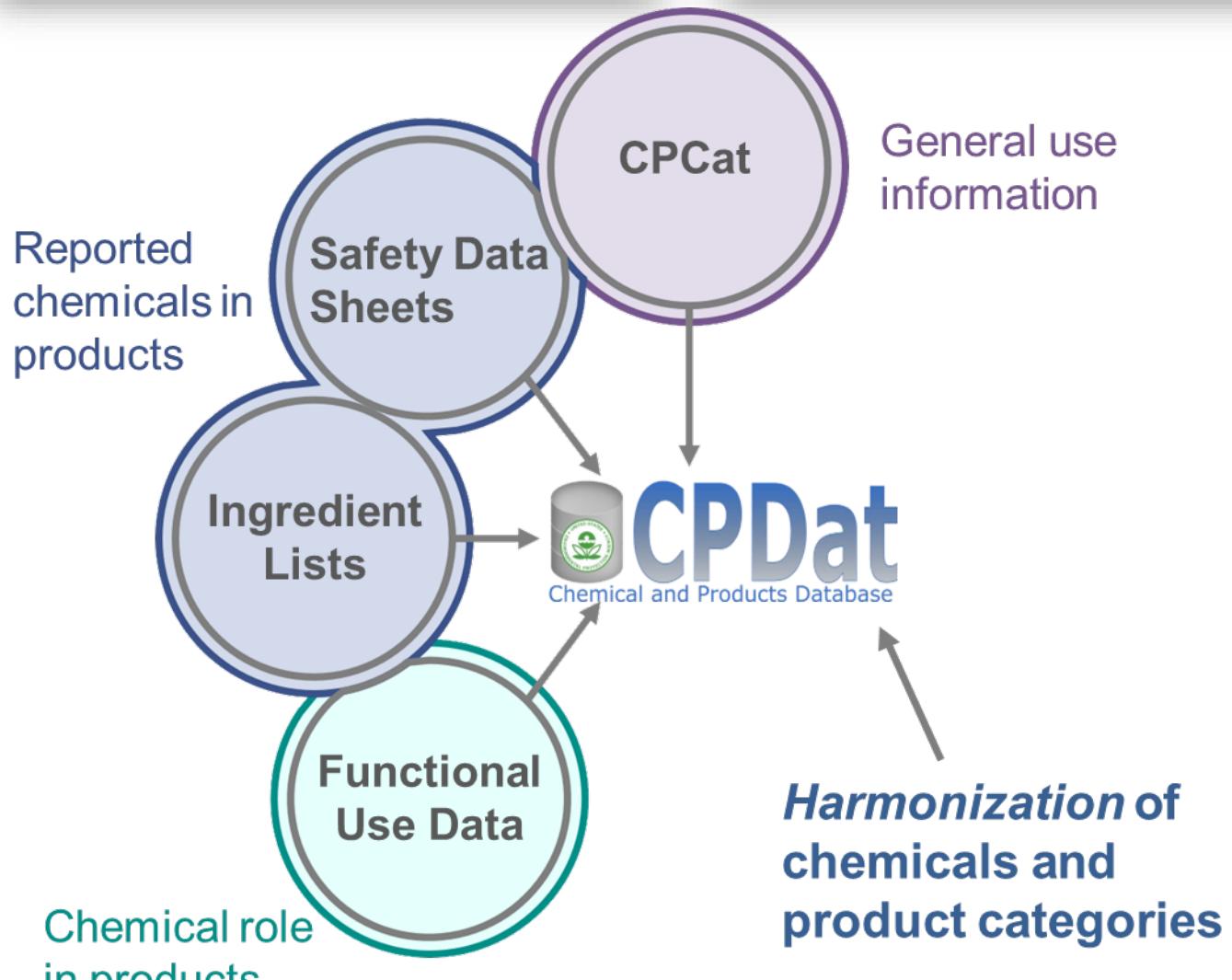
**Statistical frameworks** for integrating models for multiple pathways with exposure data to provide calibrated exposure predictions

Integration of hazard and exposure NAMs for high-throughput **chemical prioritization**

**Prioritize**  
chemicals for further evaluation



# Curation of Chemical Descriptor NAMs



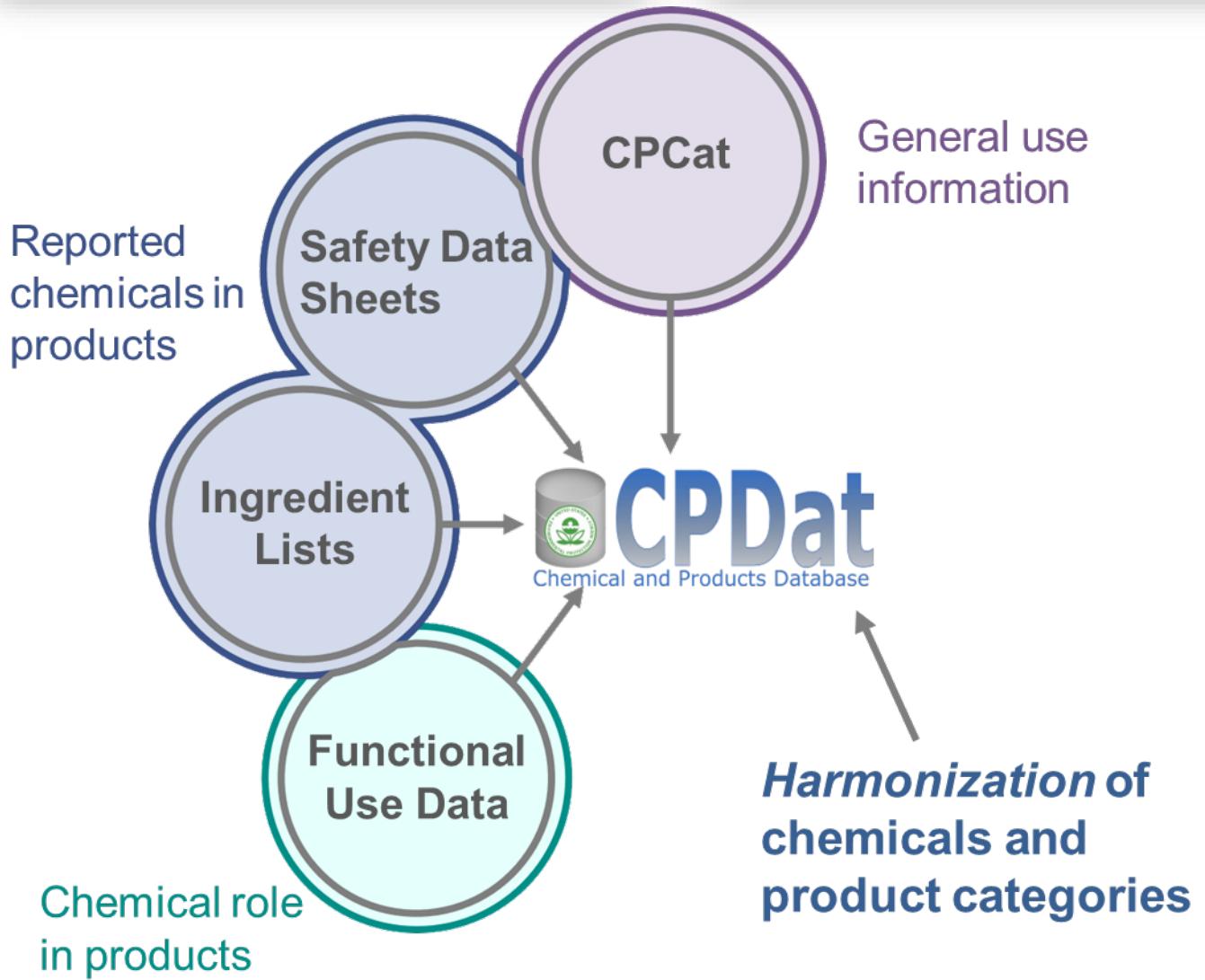

**Food and Chemical Toxicology**  
 journal homepage: [www.elsevier.com/locate/foodchemtox](http://www.elsevier.com/locate/foodchemtox)

Development of a consumer product ingredient database for chemical exposure screening and prioritization  
 M.-R. Goldsmith <sup>a,\*</sup>, C.M. Grulke <sup>a</sup>, R.D. Brooks <sup>b</sup>, T.R. Transue <sup>c</sup>, Y.M. Tan <sup>a</sup>, A. Frame <sup>a,e</sup>, P.P. Egeghy <sup>a</sup>, R. Edwards <sup>d</sup>, D.T. Chang <sup>a</sup>, R. Tornero-Velez <sup>a</sup>, K. Isaacs <sup>a</sup>, A. Wang <sup>a,c</sup>, I. Johnson <sup>a</sup>, K. Holm <sup>a</sup>, M. Reich <sup>f</sup>  
 Toxicology Reports  
 journal homepage: [www.elsevier.com/locate/toxrep](http://www.elsevier.com/locate/toxrep)

Exploring consumer exposure pathways and patterns of use for chemicals in the environment  
 Kathie L. Dionisio <sup>a</sup>, Alicia M. Frame <sup>b,1</sup>, Michael-Rock Goldsmith <sup>a,2</sup>, John F. Wambach <sup>b</sup>, Alan Liddell <sup>c,3</sup>, Tommy Cathey <sup>d</sup>, Doris Smith <sup>b</sup>, James V. Kristin K.  
 ORIGINAL ARTICLE  
 Consumer product chemical weight fractions from ingredient lists  
 Kristin K. Isaacs <sup>1</sup>, Katherine A. Phillips <sup>1</sup>, Derya Biryol <sup>1,2</sup>, Kathie L. Dionisio <sup>1</sup> and Paul S. Price <sup>1</sup>  
 Journal of Exposure Science & Environmental Epidemiology (2020) 30:171–183  
<https://doi.org/10.1038/s41370-019-0187-5>

**ARTICLE**  
 Establishing a system of consumer product use categories to support rapid modeling of human exposure  
 Kristin K.

**SCIENTIFIC DATA**  
 OPEN  
 Data Descriptor: The Chemical and Products Database, a resource for exposure-relevant data on chemicals in consumer products  
 Received: 16 October 2017  
 Accepted: 30 April 2018  
 Published: 10 July 2018  
 Kathie L. Dionisio <sup>1</sup>, Katherine Phillips <sup>1</sup>, Paul S. Price <sup>1</sup>, Christopher M. Grulke <sup>2</sup>, Antony Williams <sup>2</sup>, Derya Biryol <sup>1,3</sup>, Tao Hong <sup>4</sup> & Kristin K. Isaacs <sup>1</sup>

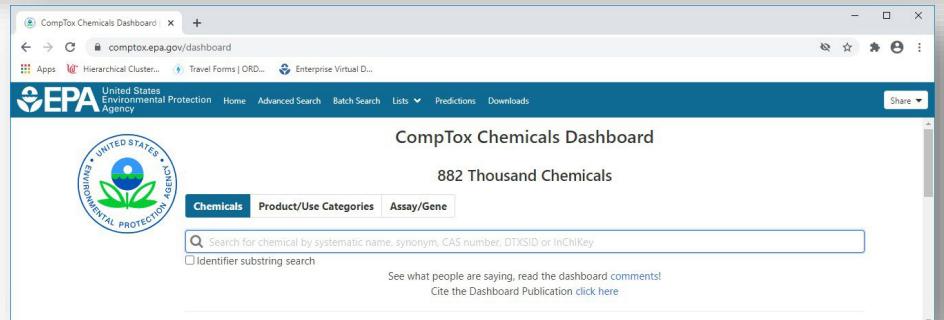


<https://comptox.epa.gov/chemexpo/>

## Welcome to ChemExpo Knowledgebase

ChemExpo is an interactive web application for exploring chemical data, curated from public documents, relevant to exposure assessment. ChemExpo currently surfaces data collected by EPA about how chemicals are used in commerce and how they occur in consumer and industrial products; these data are collectively known as the Chemicals and Products Database (CPDat). ChemExpo provides tools for exploring and downloading these data, which include consumer product composition, chemical functional use, and general chemical use information. The ChemExpo team actively works to curate these data to specific consumer and occupational product categories, to chemical functional uses, and to substance identifiers (DTXIDs) used by EPA and the CompTox Chemicals Dashboard.

This beta version of ChemExpo (v0.1) is currently undergoing review and feedback from stakeholders. Its data and functionality are subject to change.



<https://comptox.epa.gov/dashboard/>

- Other curation efforts have focused on media occurrence of chemicals
- 63 million+ chemical records from 20 sources mapped to harmonized chemical identifiers and ~30 media categories
- 3271 unique chemicals

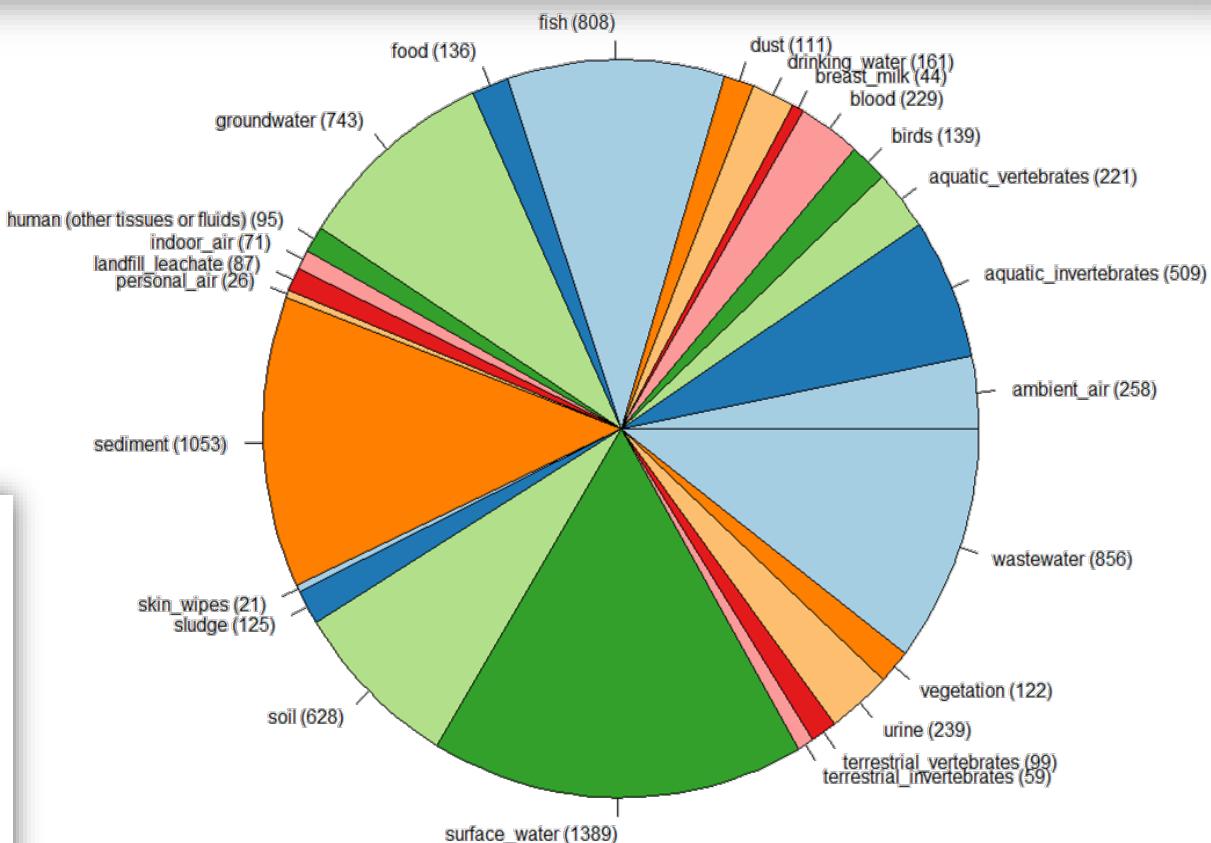
Data Descriptor | Open Access | Published: 16 June 2022

## A harmonized chemical monitoring database for support of exposure assessments

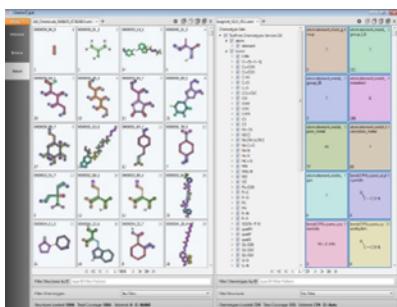
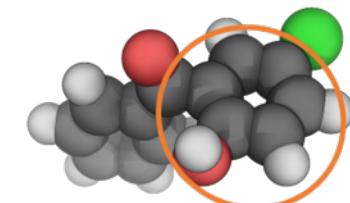
[Kristin K. Isaacs](#) , [Jonathan T. Wall](#), [Ashley R. Williams](#), [Kevin A. Hobbie](#), [Jon R. Sobus](#), [Elin Ulrich](#), [David Lyons](#), [Kathie L. Dionisio](#), [Antony J. Williams](#), [Christopher Grulke](#), [Caroline A. Foster](#), [Josiah McCoy](#) & [Charles Bevington](#)

[Scientific Data](#) 9, Article number: 314 (2022) | [Cite this article](#)

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# Machine Learning NAMs



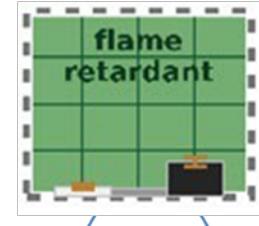
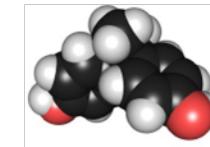
**CPDat**  
Chemical and Products Database



Machine Learning Based Classification Models

Training Sets of Chemical Descriptor NAMs

Prediction of Potential Alternatives from Chemical Libraries



YES      NO

Green Chemistry

PAPER



Cite this: *Green Chem.*, 2017, 19, 1063

High-throughput screening of chemicals as functional substitutes using structure-based classification models†



[View Article Online](#)

[View Journal](#)

[View Issue](#)

# High-throughput Measurement NAMs

- “Targeted” Analysis:
  - We know exactly what were looking for
  - 10s – 100s of chemicals
- “Non-Targeted” Analysis (NTA):
  - We have no preconceived lists
  - 1,000s – 10,000s of chemical
- NTA research is being performed both in-house by EPA investigators and via contract
- Key focus on developing **reproducible and defensible** NTA methods and results
- Ultimate goal is to develop tools, databases, and workflows for rapid analysis of any sample for chemicals of interest, i.e. ***exposure forensics***



Targeted Analysis



NTA



Material from Jon Sobus

# High-throughput Measurement NAMs

## Source and Release

### Pilot: 20 Consumer Product Categories



Phillips et al., Env. Sci. Tech. 2018

### Recycled Consumer Materials



Lowe et al., Env. Sci. Tech. 2018

### Consumer Product Emissions from Different Substrates



Watson et al., Env. Sci. Tech. 2024 (in press)

## Media Occurrence

### Residential Air



### Residential Dust



Rager et al., Env. Int., 2016

### Drinking Water



Newton et al., Env. Pollut., 2019

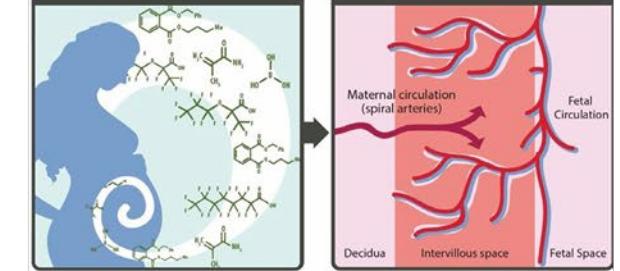
## Exposure

### Pooled Human Blood



Phillips et al., 2023

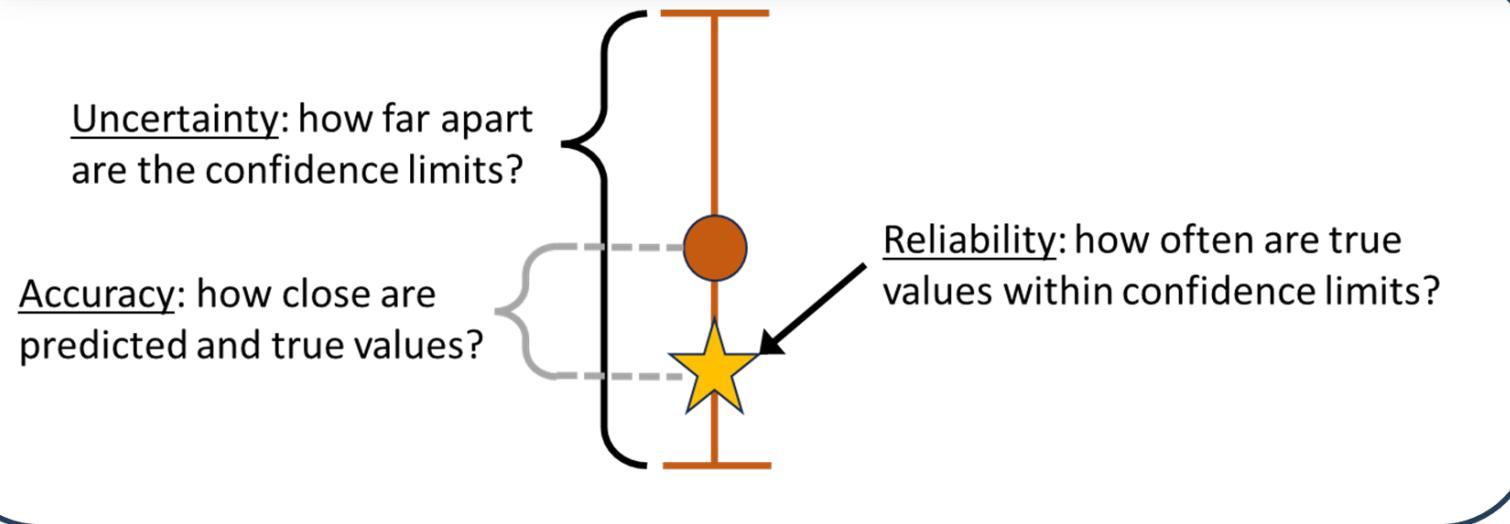
### Human Placenta



Rager et al., Repro. Tox., 2020  
Chao et al., Env. Int., 2022

**Current Focus and Challenge:** How can we quantify concentrations of chemicals in media using NTA?

Methods are being developed for selecting appropriate calibration surrogates and characterizing qNTA performance



= true conc.



= estimated conc.



= estimated confidence interval

Analytical and Bioanalytical Chemistry (2022) 414:4919–4933  
<https://doi.org/10.1007/s00216-022-04118-z>

RESEARCH PAPER

Uncertainty estimation strategies for quantitative non-targeted analysis

Louis C. Groff II<sup>1,2</sup> · Jarod N. Grossman<sup>2,3</sup> · Anneli Kruve<sup>4</sup> · Jeffrey M. Minucci<sup>1</sup> · Charles N. Lowe<sup>1</sup> · James P. McCord<sup>1</sup> · Dustin F. Kapraun<sup>1</sup> · Katherine A. Phillips<sup>1</sup> · S. Thomas Purucker<sup>1</sup> · Alex Chao<sup>1</sup> · Caroline L. Ring<sup>1</sup> · Antony J. Williams<sup>1</sup> · Jon R. Sobus<sup>1</sup>



Quantitative non-targeted analysis: Bridging the gap between contaminant discovery and risk characterization

James P. McCord<sup>a,\*</sup>, Louis C. Groff II<sup>b,c</sup>, Jon R. Sobus<sup>b</sup>

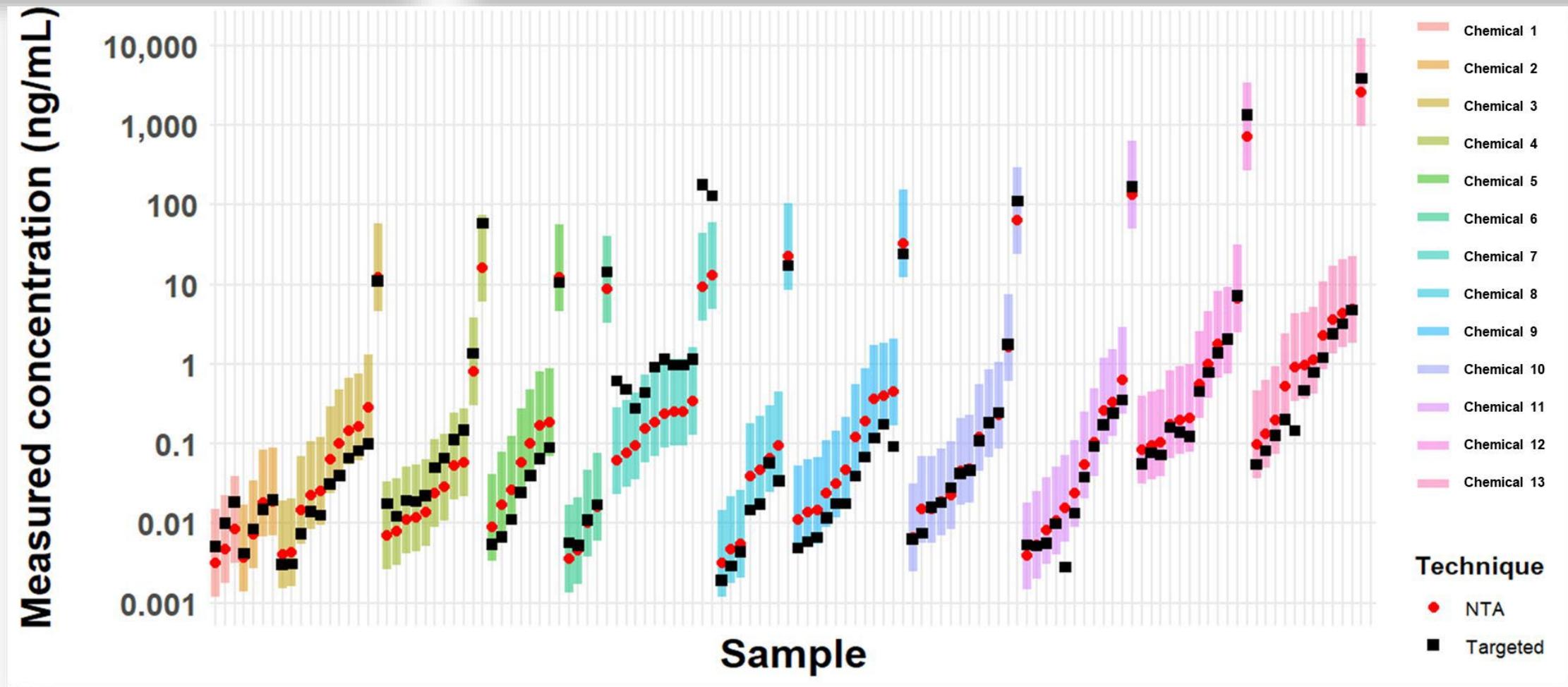
Analytical and Bioanalytical Chemistry (2024) 416:1249–1267  
<https://doi.org/10.1007/s00216-023-05117-4>

RESEARCH PAPER

Establishing performance metrics for quantitative non-targeted analysis: a demonstration using per- and polyfluoroalkyl substances

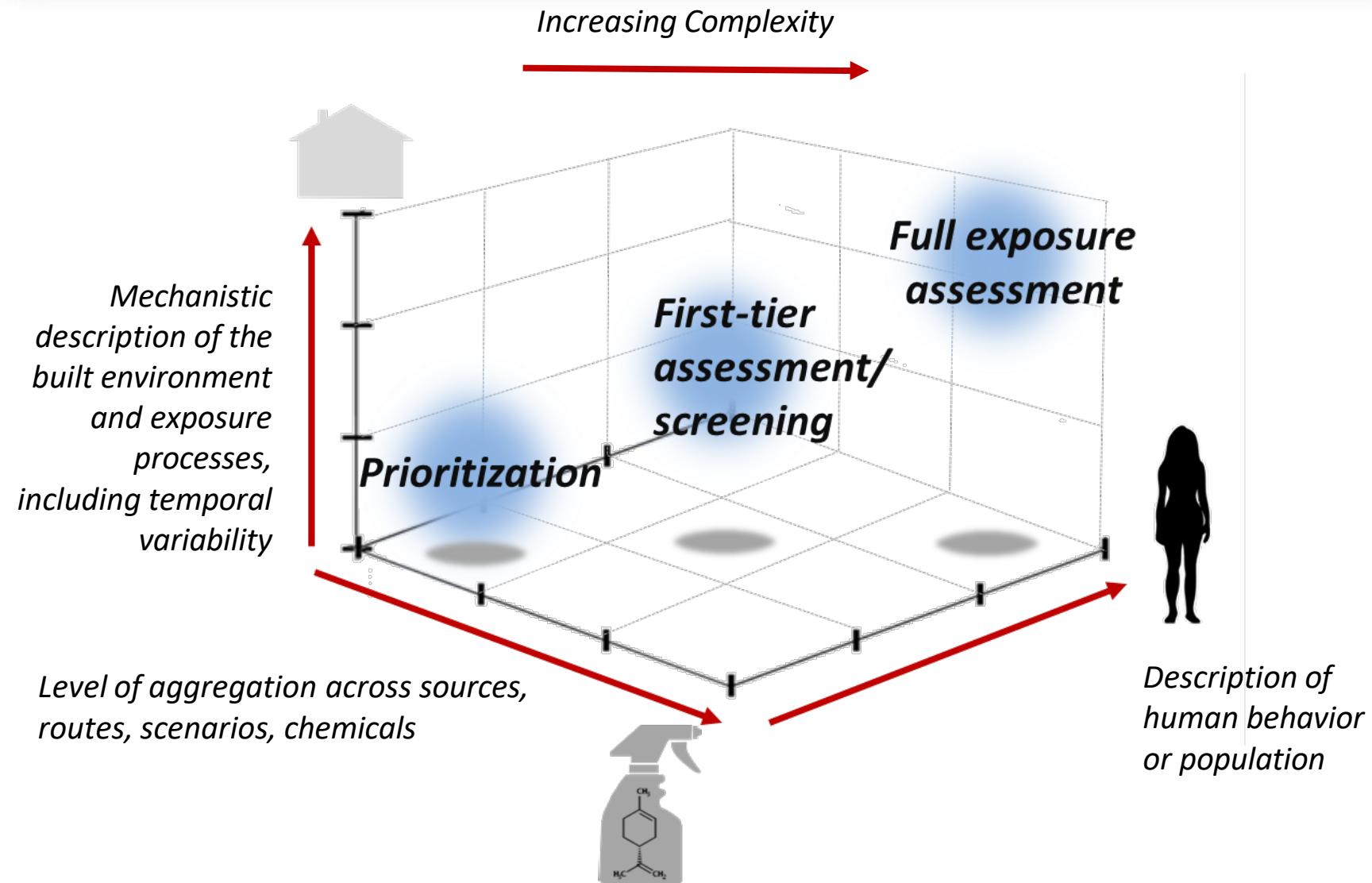
Shirley Pu<sup>1,2</sup> · James P. McCord<sup>3</sup> · Jacqueline Bangma<sup>3</sup> · Jon R. Sobus<sup>1</sup>

# Towards Quantitative NTA (qNTA)

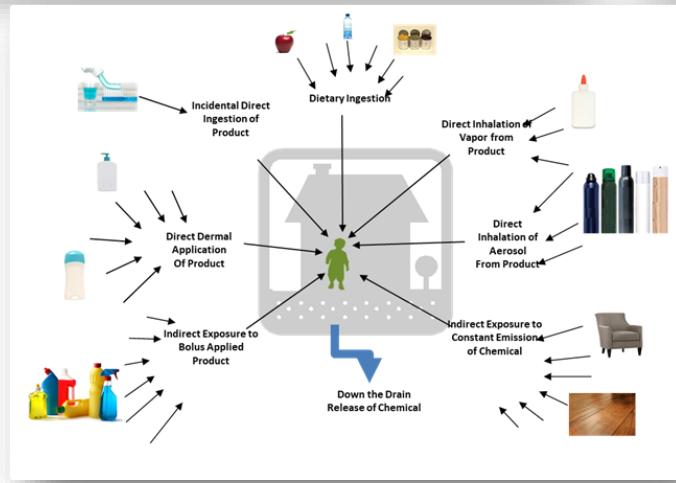


# High Throughput Exposure Model NAMs

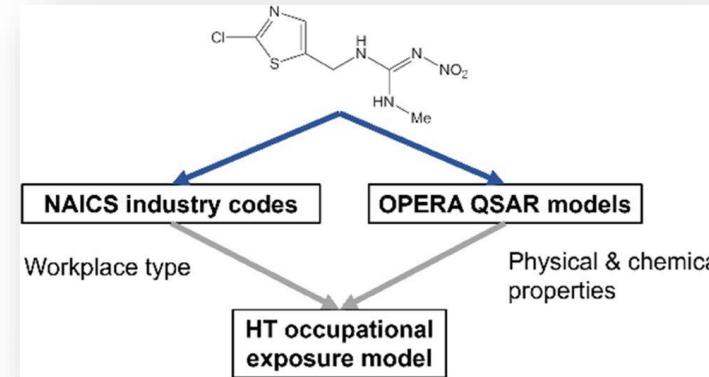
- All models vary in complexity and data needed to describe chemical exposure
- **High throughput exposure (HTE) models** can handle many chemicals with minimal descriptive information
- HTE models can provide rough but quantitative estimates of exposure



# High Throughput Exposure Model NAMs

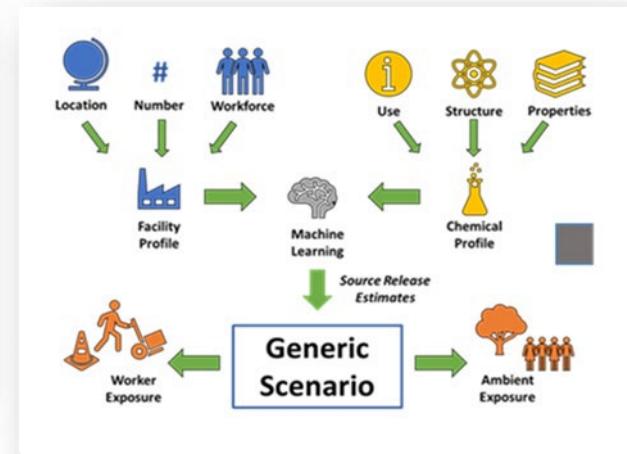


**Consumer** Isaacs et al., Env. Sci. Tech. (2014)



## Occupational

Minucci et al., Env. Int. (2023)



Ambient

Meyer et al., ACS Sustain Chem Eng. (2019)



Environment International  
Volume 108, November 2017, Pages 185-194



High-throughput dietary exposure predictions for chemical migrants from food contact substances for use in chemical prioritization

**Dietary**

Biryol et al., (2017)



Science of The Total Environment  
Volumes 605–606, 15 December 2017, Pages 471-481

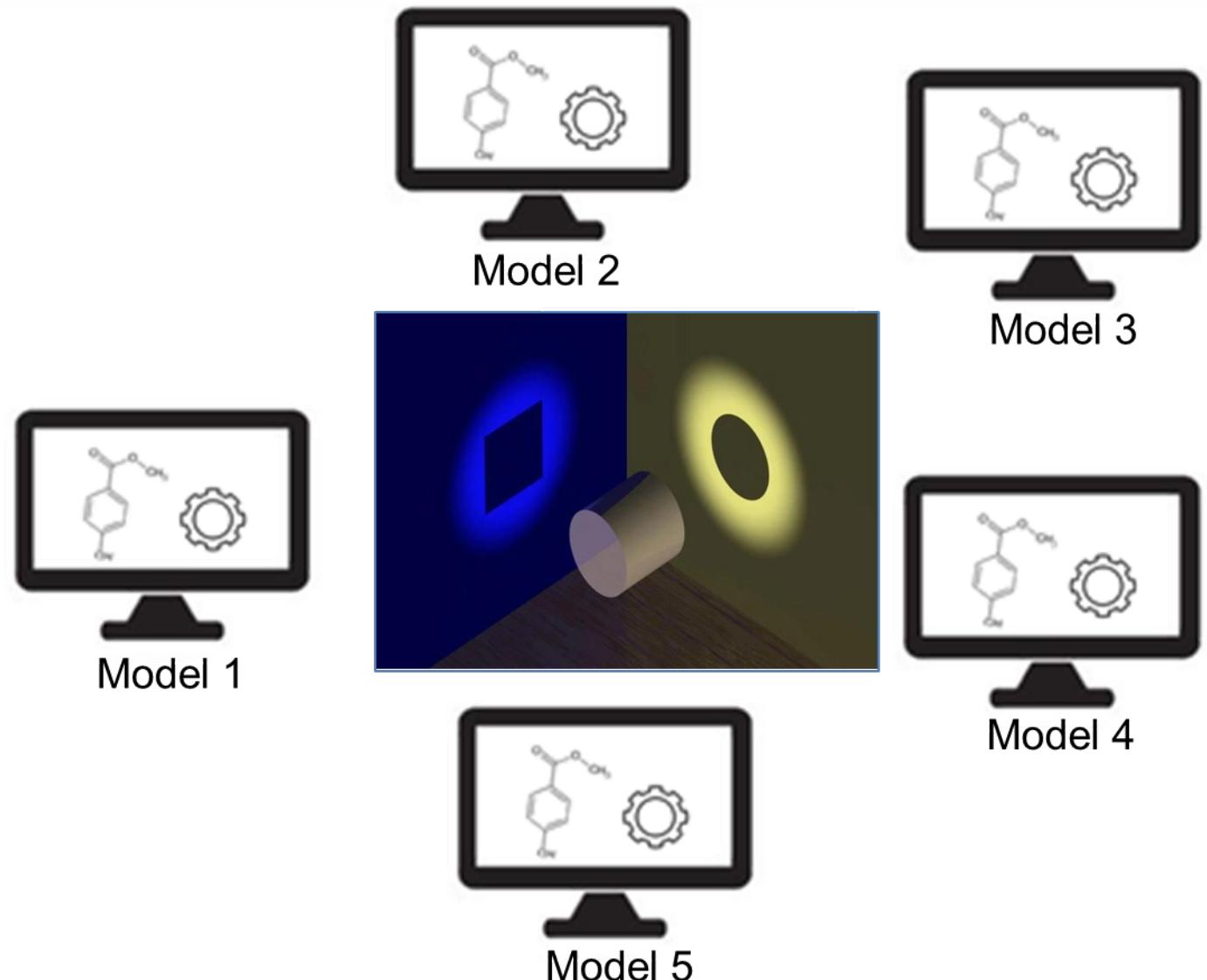
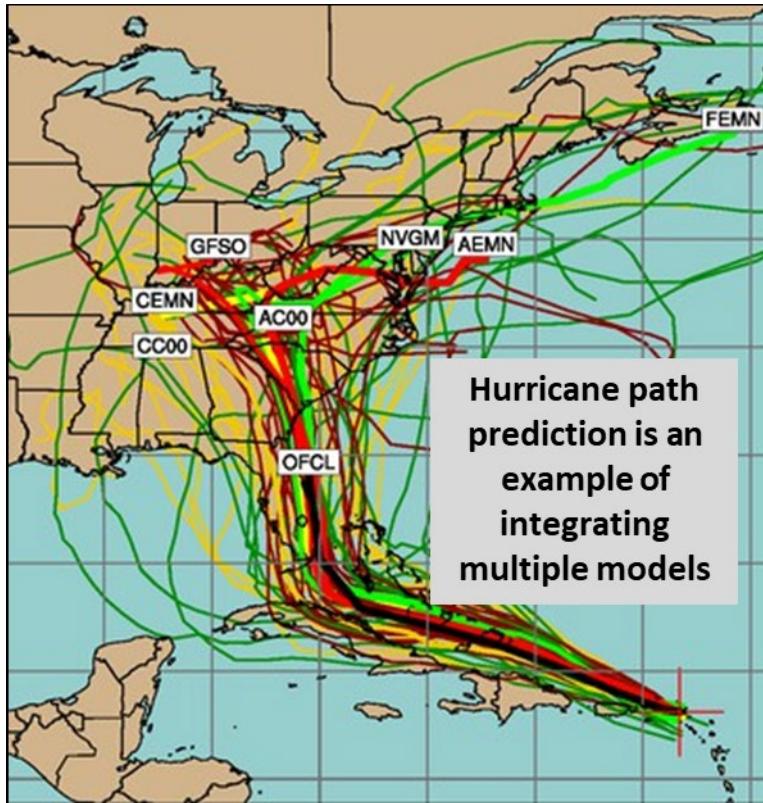


Developing and applying metamodels of high resolution process-based simulations for high throughput exposure assessment of organic chemicals in riverine ecosystems

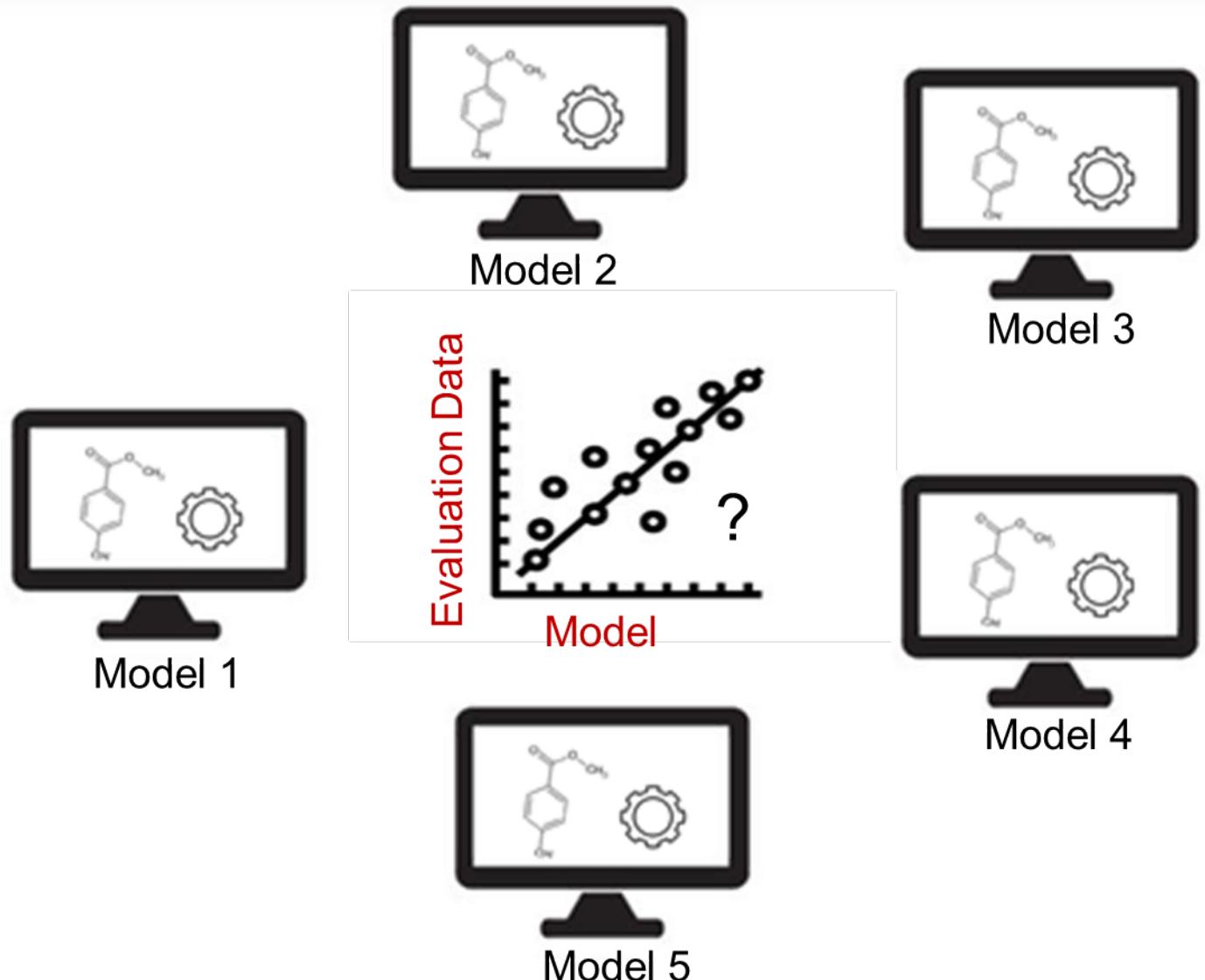
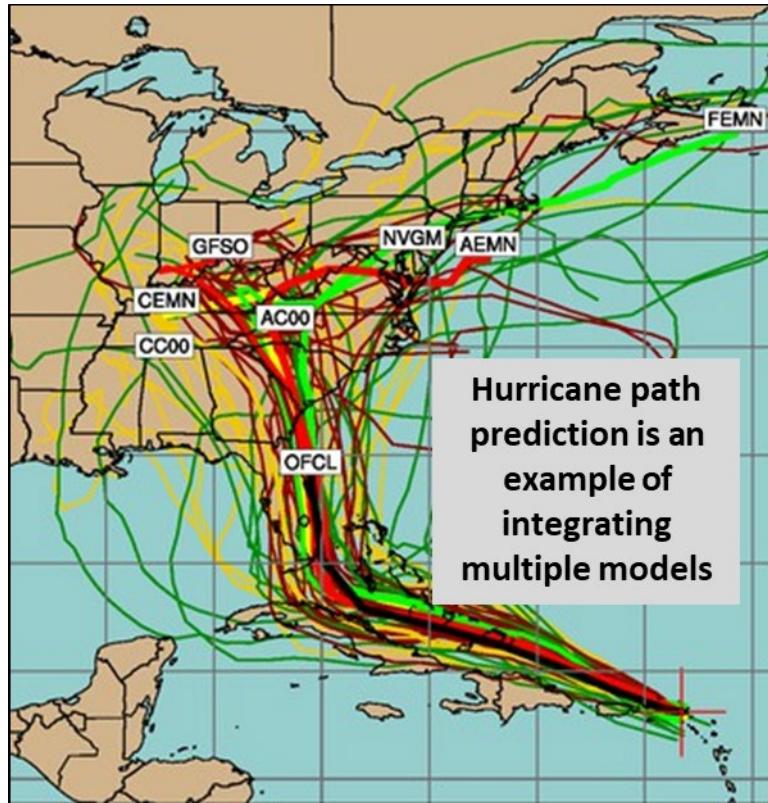
**Ecological**

Barber et al., (2017)

# Evaluation Framework NAMs and Consensus Models

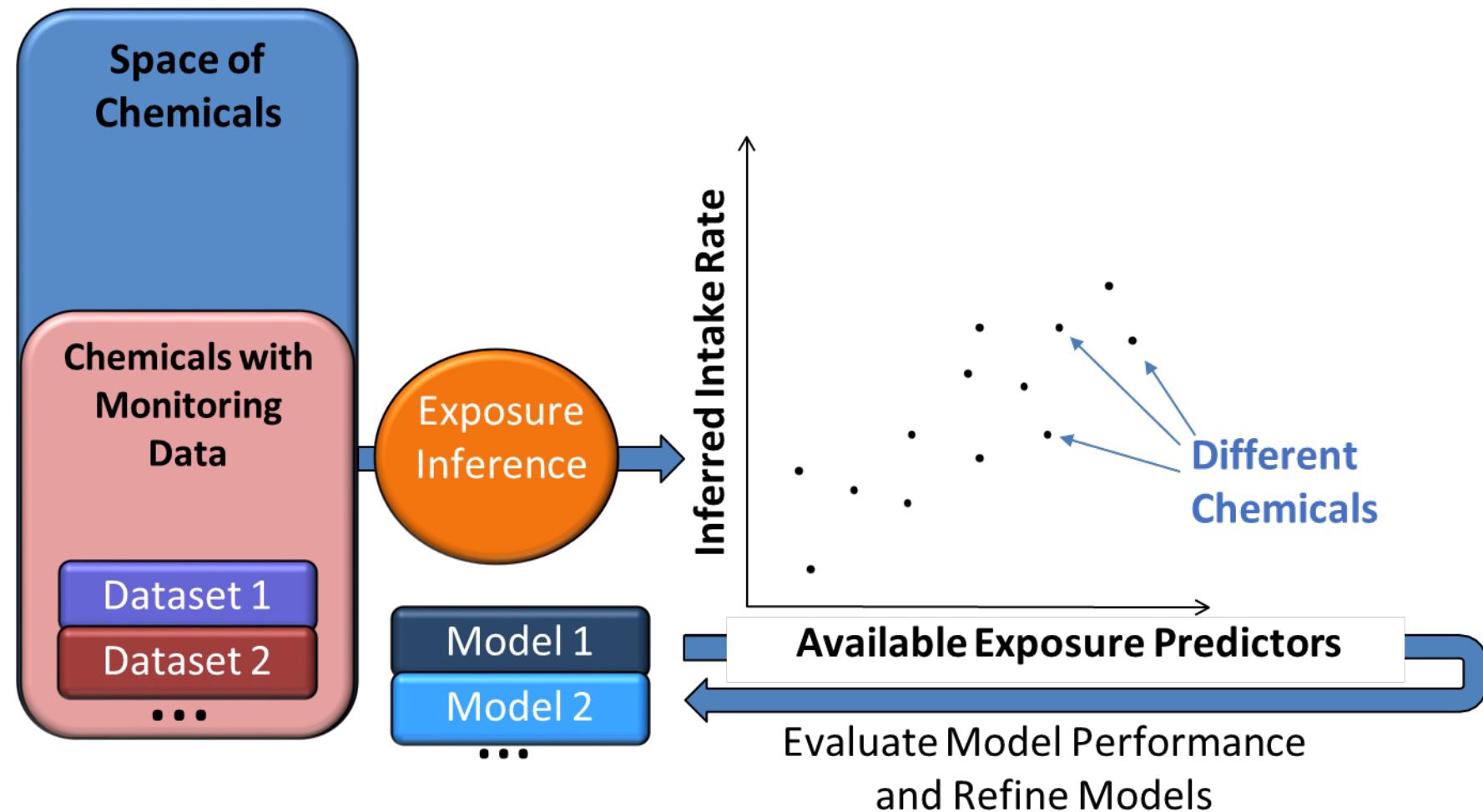


# Evaluation Framework NAMs and Consensus Models



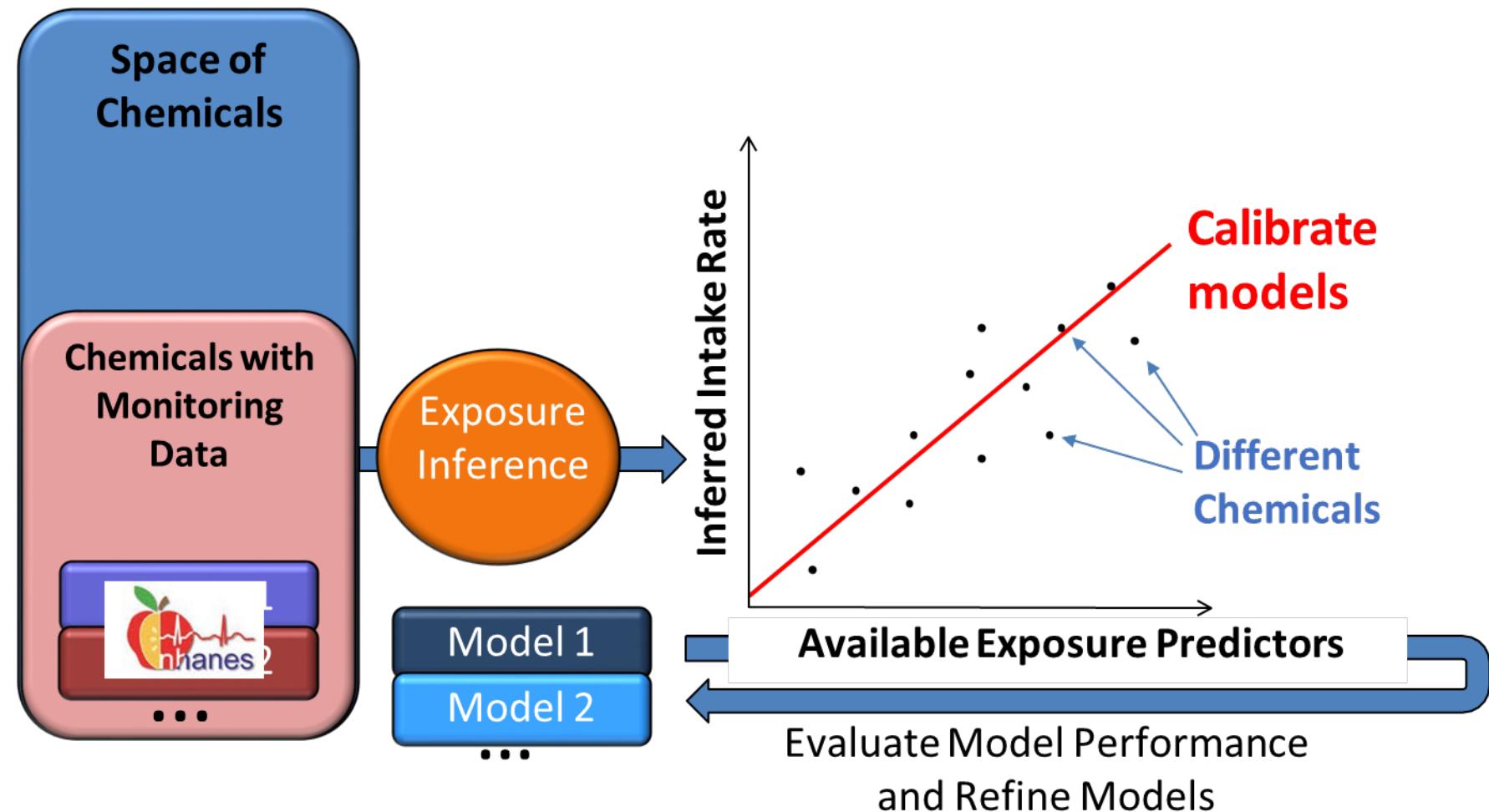
# Evaluating Exposure Models with the SEEM Framework

- We use Bayesian methods to incorporate multiple models into consensus forecasts for 1000s of chemicals within the **Systematic Empirical Evaluation of Models (SEEM)** (Wambaugh et al., 2013,2014)



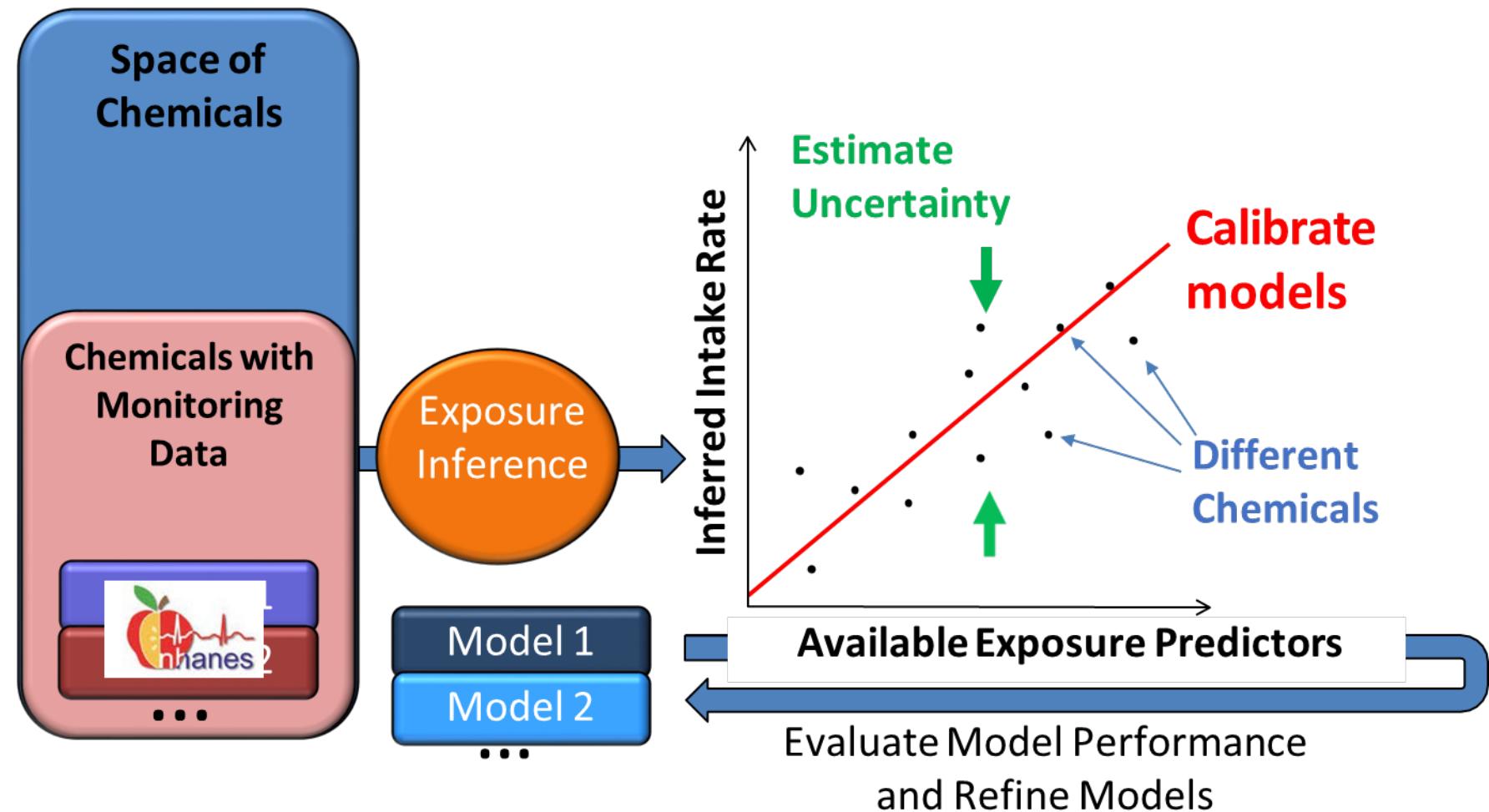
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  - We infer parent chemical exposures from NHANES urine and serum metabolite measurements



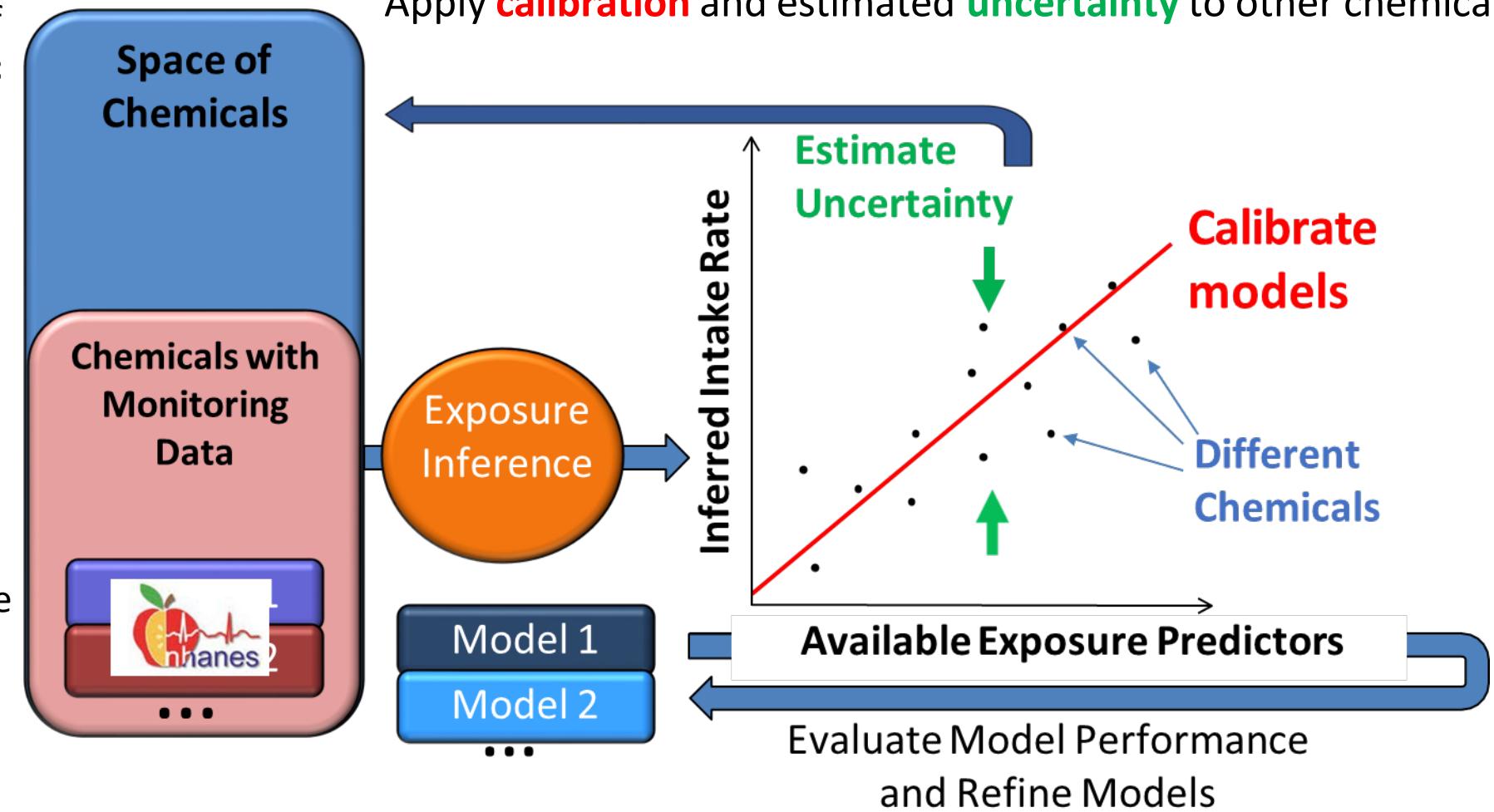
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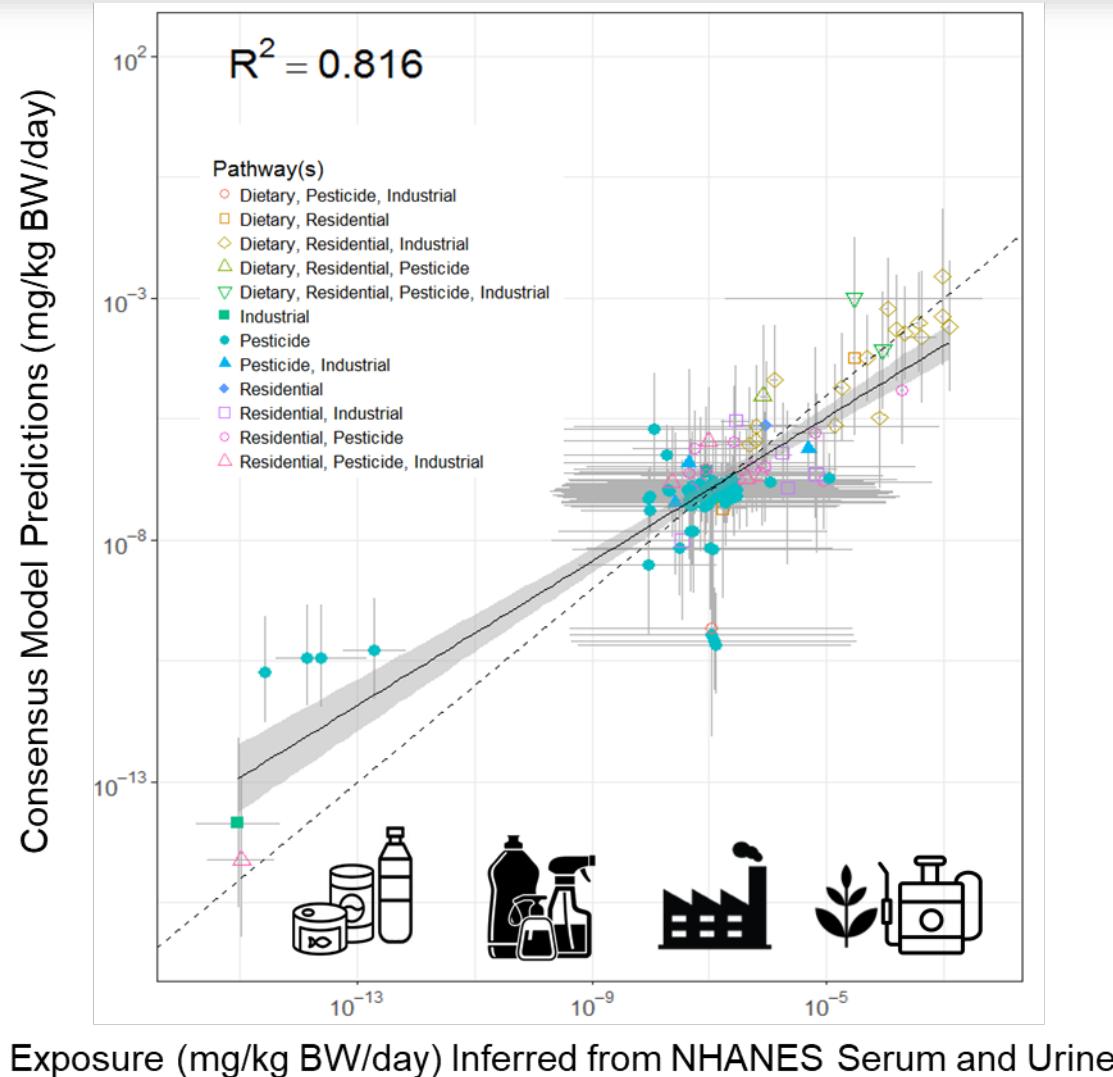
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- We apply the SEEM regression to thousands of other chemicals with predictors

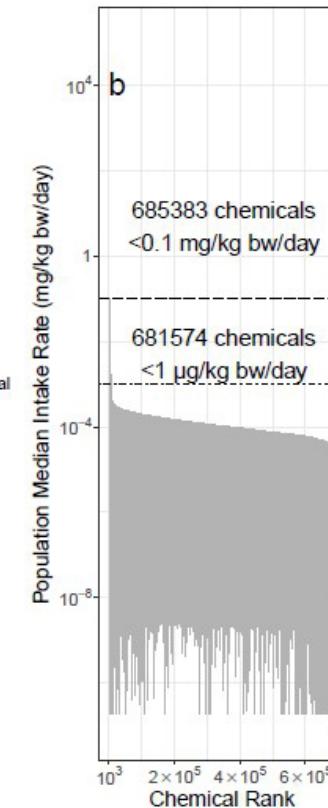
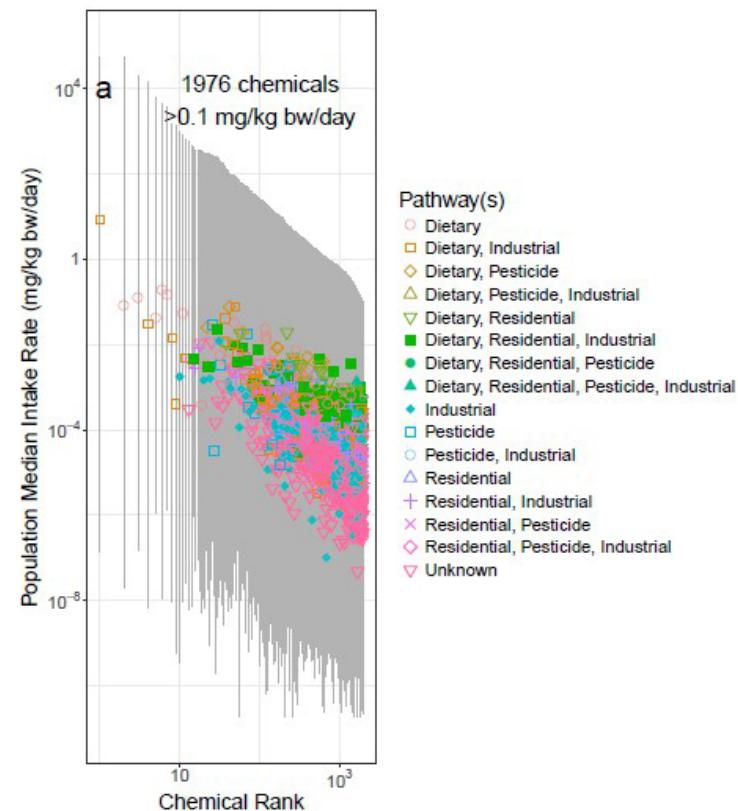


# SEEM3 Collaboration

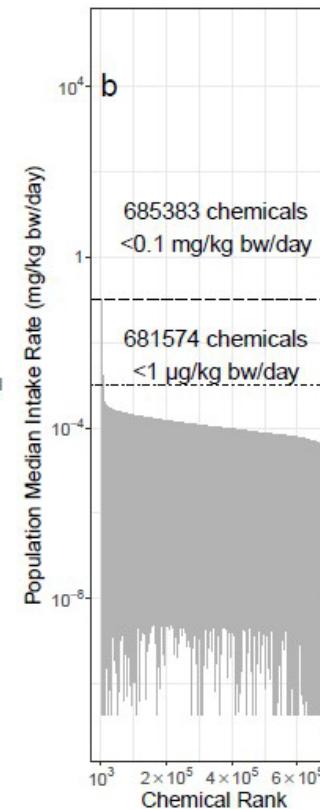
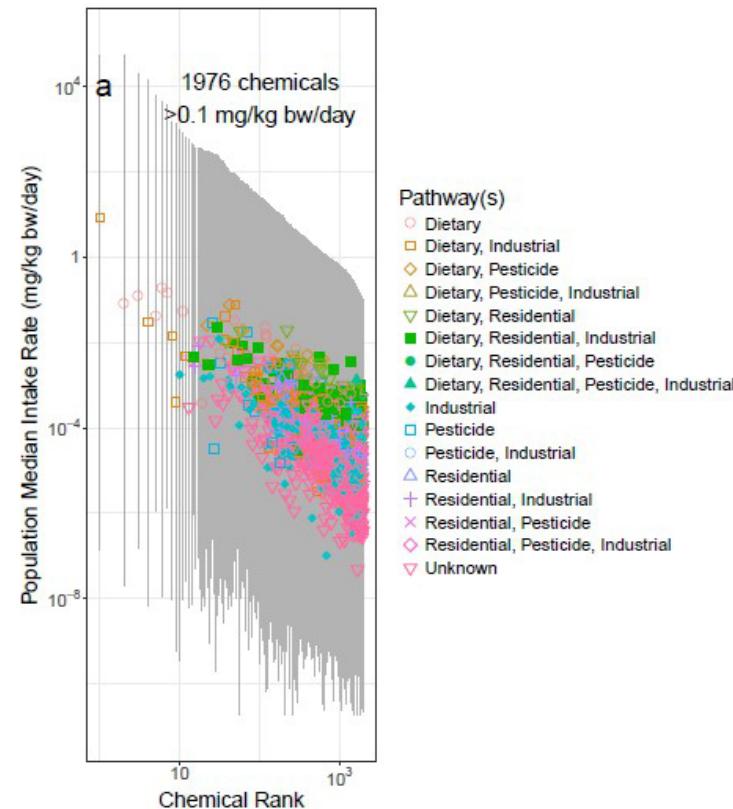
- Third generation SEEM model incorporates 12 exposure predictors, including high-throughput exposure models from ExpoCast and its collaborators
- SEEM3 first predicts relevance of **four exposure pathways** from chemical structure using machine learning
- Predictors are weighted according to their ability to explain NHANES data



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- SEEM3 consensus model provides estimates of human median intake rate (mg/kg/day) for nearly 500,000 chemicals via the CompTox Chemicals Dashboard (<http://comptox.epa.gov/dashboard>)

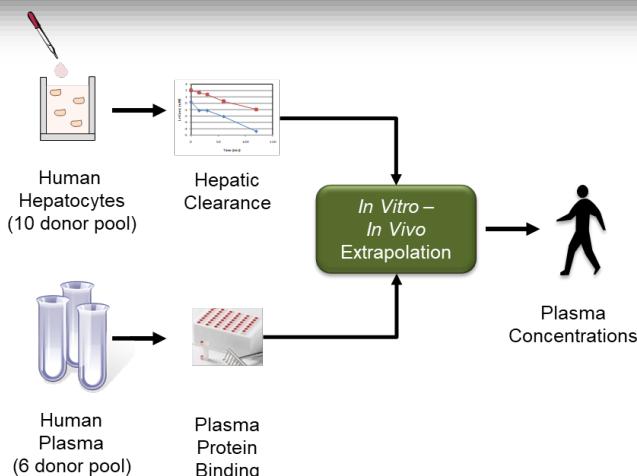


- Third generation SEEM model incorporates 12 exposure predictors, including high-throughput exposure models from ExpoCast and its collaborators
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- **SEEM4: Considering cohort in evaluation data (including new NHANES data for children) and HT model predictions**



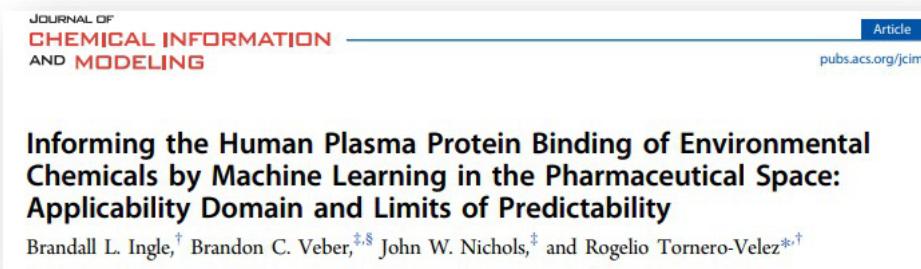
# Toxicokinetics NAMs

- Chemical-specific data for toxicokinetics (TK) are as sparse as they are for exposure
- High-throughput TK measurements have provided data for nearly 1000 chemicals over the past decade
- However, thousands of chemicals still have no data – therefore we employ machine learning and QSAR approaches
- Data and models incorporated into open source R package, `httk`:



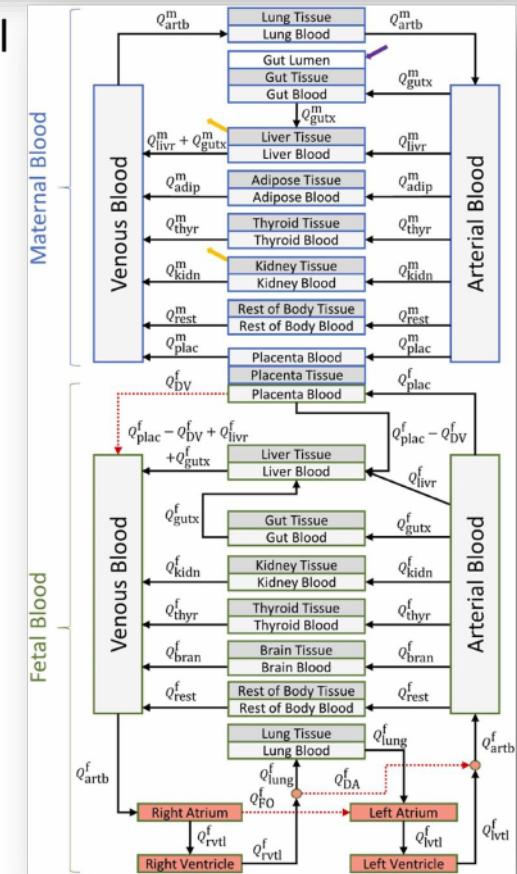
- Rotroff et al. (2010) 35 chemicals
- Wetmore et al. (2012) +204 chemicals
- Wetmore et al. (2015) +163 chemicals
- Wambaugh et al. (2019) + ~300 chemicals

## ***In vitro Measurements***



## Machine Learning Models

## Fetal-Maternal PBTK Model

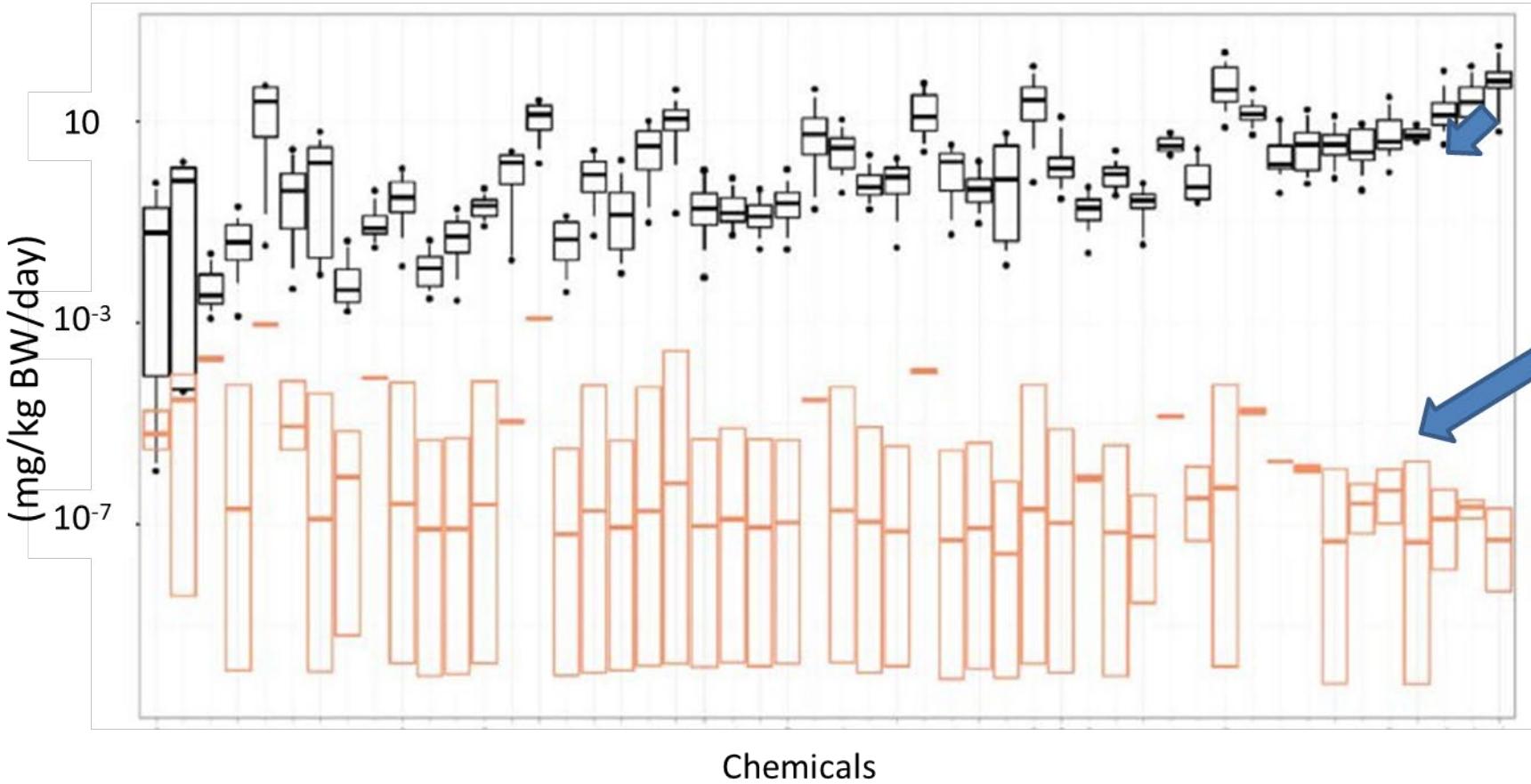


Kapraun et al., *Reprotox* 2022

## Generic Physiologically-Based Toxicokinetic Models

## Material from John Wambaugh

Estimated Equivalent Dose or Predicted Exposure



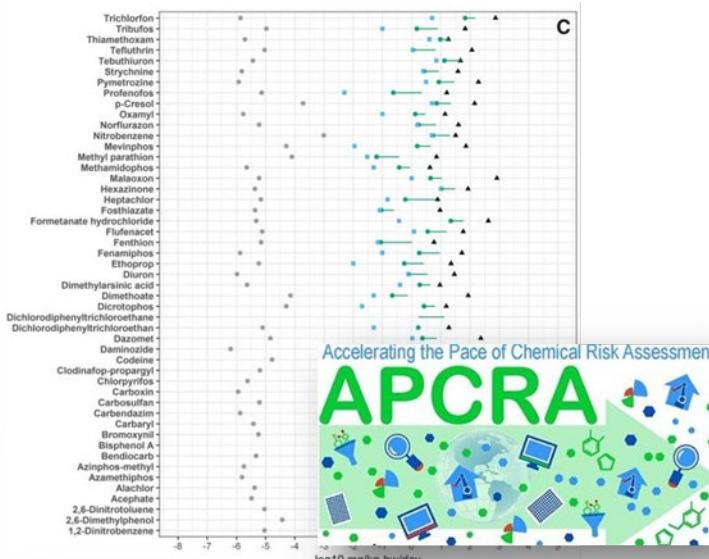
High throughput *in vitro* screening + toxicokinetics NAMs can estimate intake exposures needed to cause bioactivity

Consensus exposure rates with uncertainty (e.g., SEEM3)

Estimates of bioactivity, TK, and exposure are available for thousands of chemicals from:  
<https://comptox.epa.gov/dashboard/>

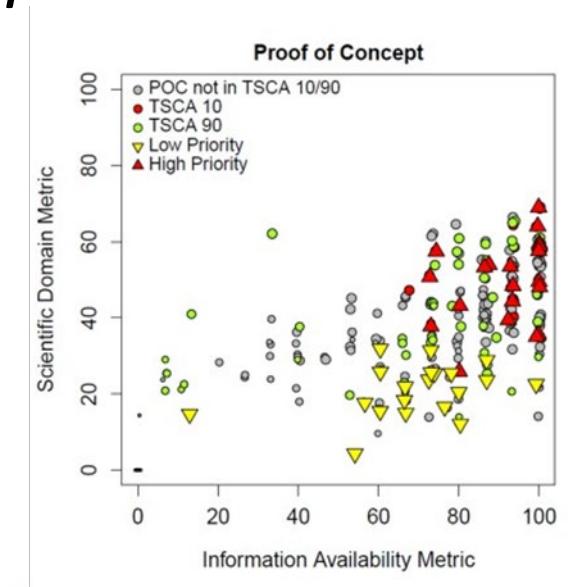
# Prioritization NAMs: Risk-Based Evaluation in Practice

- *Informing an international government-to-government initiative advancing risk evaluation*

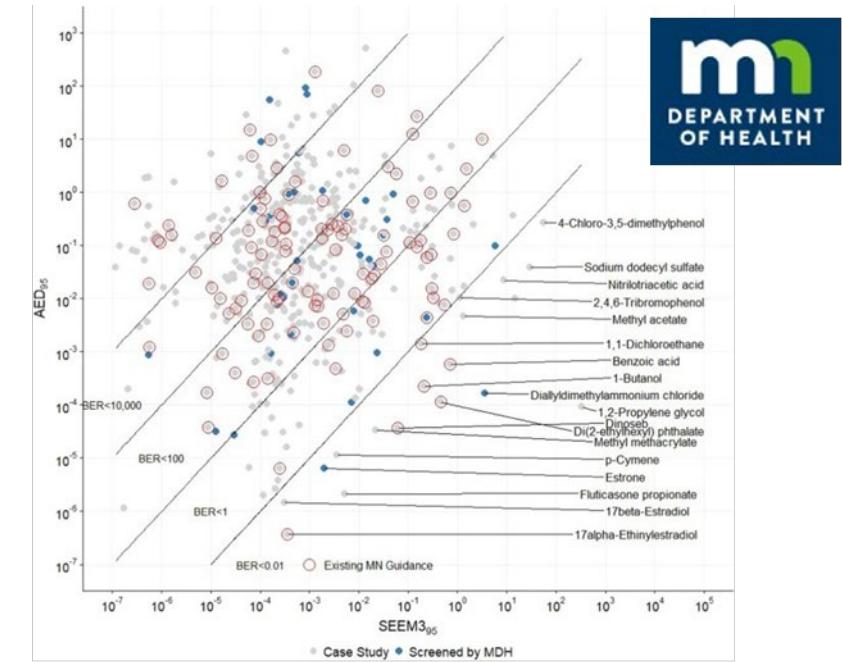


Paul-Friedman et al. (2020)

- *Screening candidates for chemical prioritization under TSCA*



- *Evaluating chemicals in state regulatory programs*



Journal of Exposure Science & Environmental Epidemiology

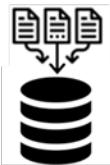
www.nature.com/jes

## ARTICLE

Screening for drinking water contaminants of concern using an automated exposure-focused workflow

Isaacs et al., (2024)

**Chemical descriptors** that provide information on chemicals in an exposure context (e.g., how chemicals are used or released)



**Machine-learning approaches** that use these descriptors to fill gaps in existing data

**Extrapolate**  
to fill data gaps

**High-throughput exposure models** for various exposure pathways

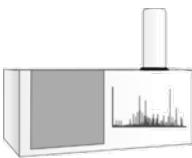
**Predict**  
exposures for specific pathways



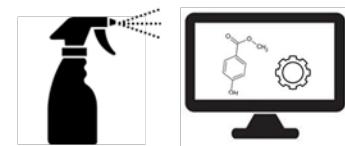
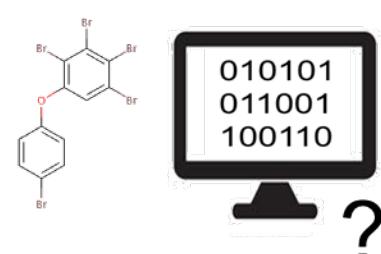
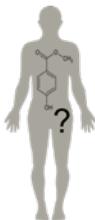
Integration of hazard and exposure NAMs for high-throughput **chemical prioritization**

**Collect**  
relevant data

**High-throughput measurements** to fill gaps in monitoring data



**High-throughput toxicokinetics** approaches for measuring and predicting chemical fate *in vivo*



**Forecast**  
exposures for 1000s of chemicals using consensus approaches

**Statistical frameworks** for integrating models for multiple pathways with exposure data to provide calibrated exposure predictions



**Prioritize**  
chemicals for further evaluation

- Exposure and toxicokinetic data are required as critical input to risk-based prioritization and screening of chemicals.
- The ExpoCast project seeks to develop the data, tools, and evaluation approaches required to generate rapid and scientifically-defensible:
  - Exposure predictions for the full universe of existing and proposed commercial chemicals.
  - The toxicokinetic data required to relate bioactive concentrations identified in high-throughput screening to predicted real-world doses (i.e. *in vitro-in vivo extrapolation*).
- We are developing and applying computational and analytical new approach methods for exposure science and toxicokinetics that are appropriate for application to 1000s of chemicals.
- Rapid prediction of chemical exposure and bioactive doses allows prioritization based upon risk.
- We aim to expand our current approaches to individual cohorts and populations.



# ExpoCast Project (Exposure Forecasting)

Center for Computational Toxicology and Exposure

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Nikki DeLuca\*

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Rachel Hencher\*

Paul Kruse \*

Seth Newton

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## Collaborators

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Jon Arnot

Johnny Westgate

Integrated Laboratory Systems

Xiaoqing Chang

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National Toxicology Program

Steve Ferguson

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Silent Spring Institute

Robin Dodson

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Michael Lawless

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Alice Yau

Kristin Favela

Summit Toxicology

Lesa Aylward

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Julia Rager

Marc Serre

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# **Integrating geospatial exposure models with NAMs to evaluate health risks from environmental chemicals**

Kyle P Messier, PhD

Tenure Track Investigator

Division of Translational Toxicology & Division of Intramural Research

# Overview

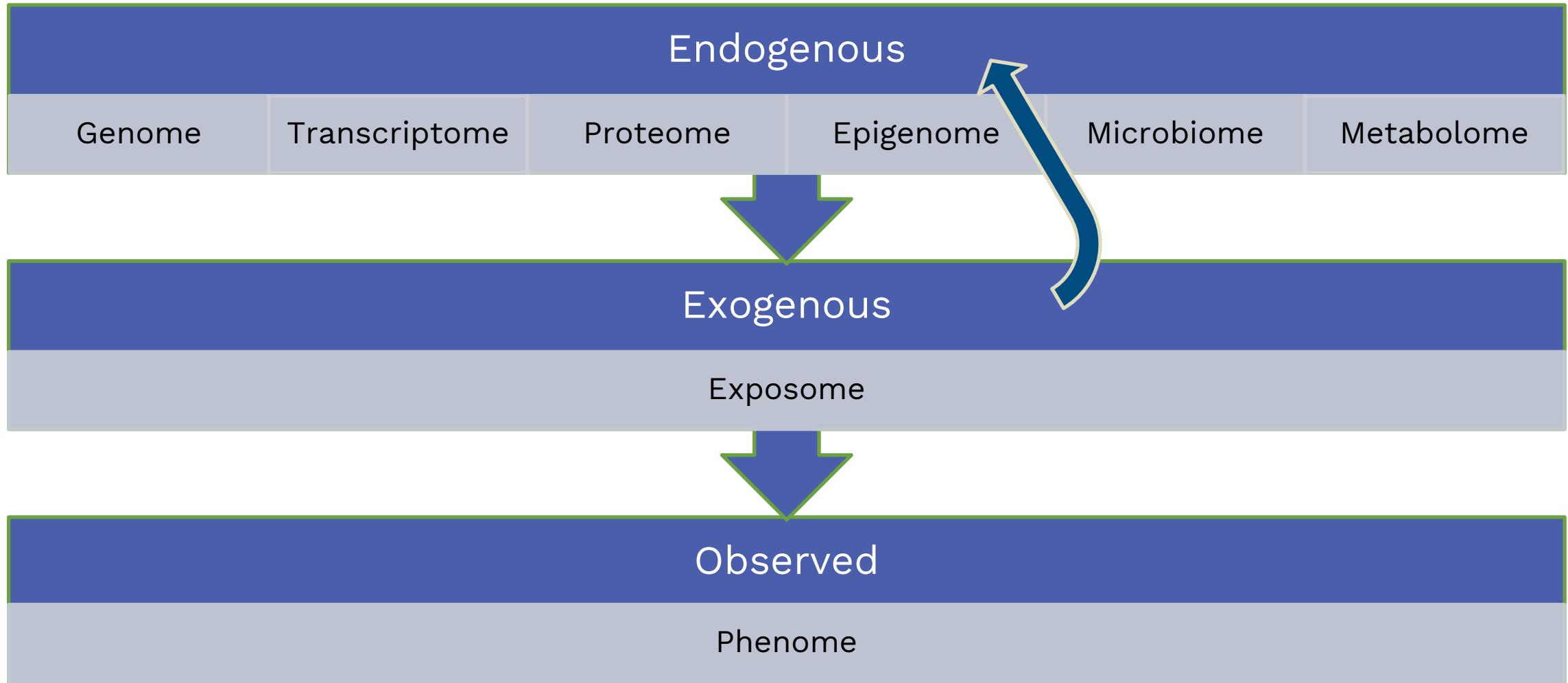
Exposome

Geospatial Exposomics via integration of spatial exposure and hazard NAMs

GeoTox Software

Best-Practices in Software to Address Complex Challenges

# Exposomics



# Internal Exposomics

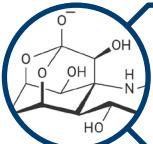
## Strengths



Non-Targeted Analysis



Novel Exposure Discovery



High Chemical Throughput



Individual Biological Samples



-Omic measurements

## Limitations

Low Sample Size Throughput

Expensive

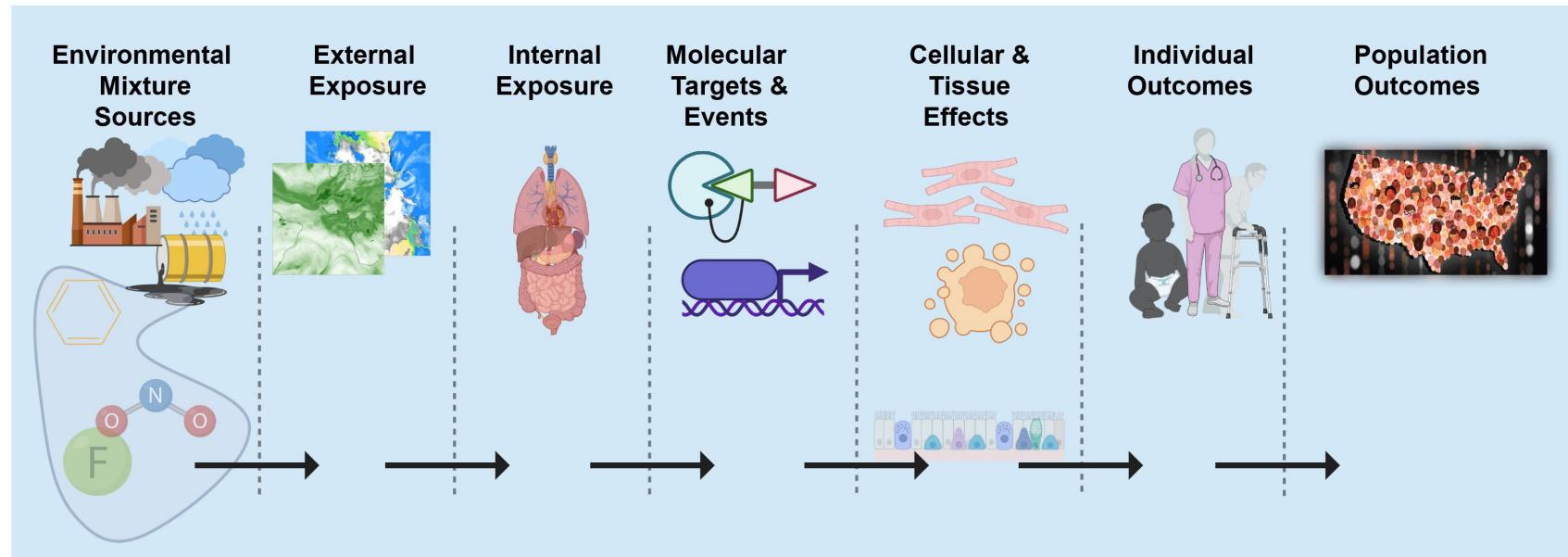
Batch/Lab Effects

[Geo] Space/Time Resolution

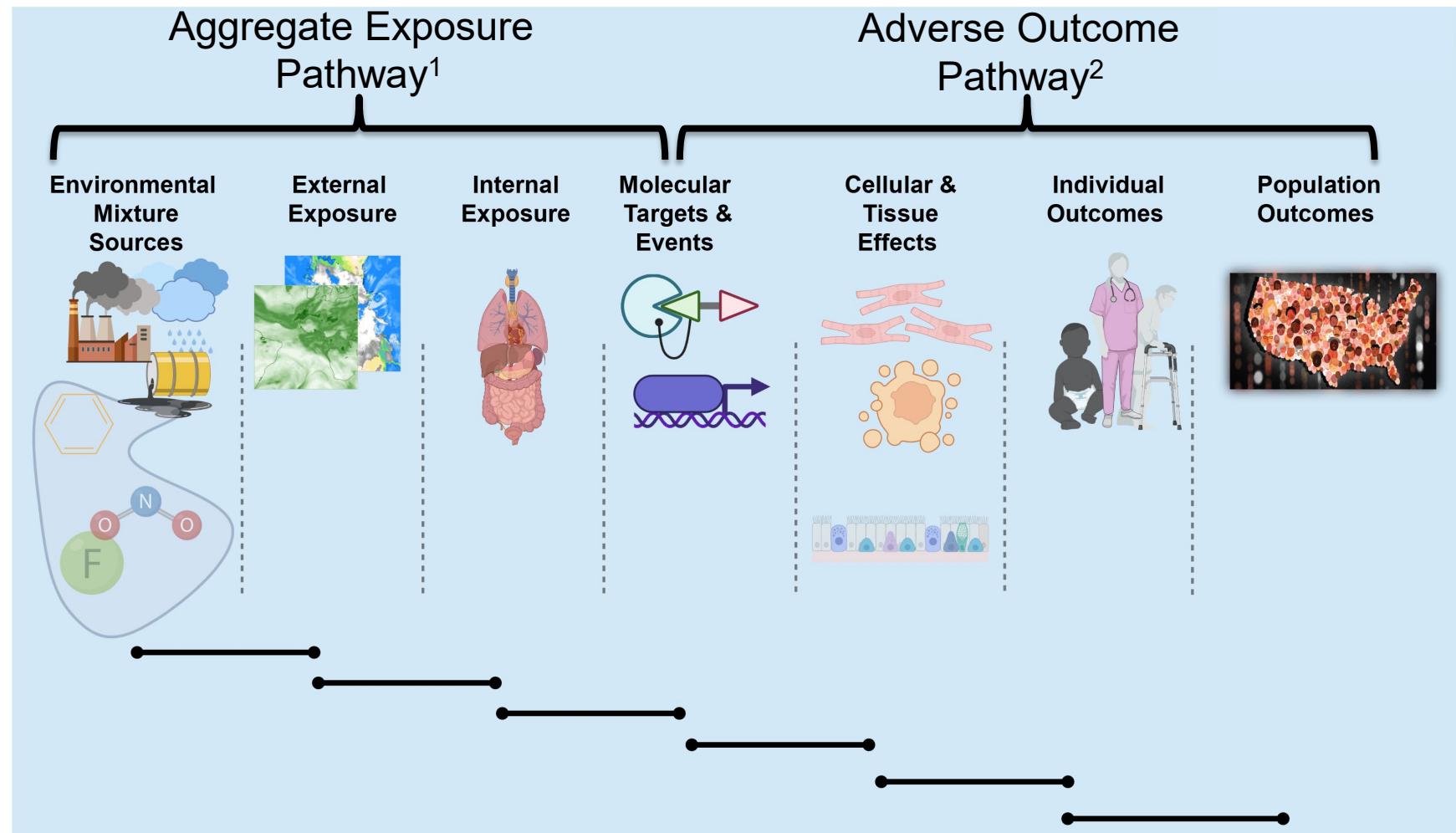
[Geo] Space/Time Variability

Endogenous/Exogenous Specificity

# Geospatial Exposomics



**Source-to-Outcome Cascade:**  
Sequential and necessary steps to  
result in an individual or population  
health outcome



1. Teeguarden JG, Tan YM, Edwards SW, Leonard JA, Anderson KA, Corley RA, Kile ML, Simonich SM, Stone D, Tanguay RL, Waters KM. Completing the link between exposure science and toxicology for improved environmental health decision making: the aggregate exposure pathway framework.

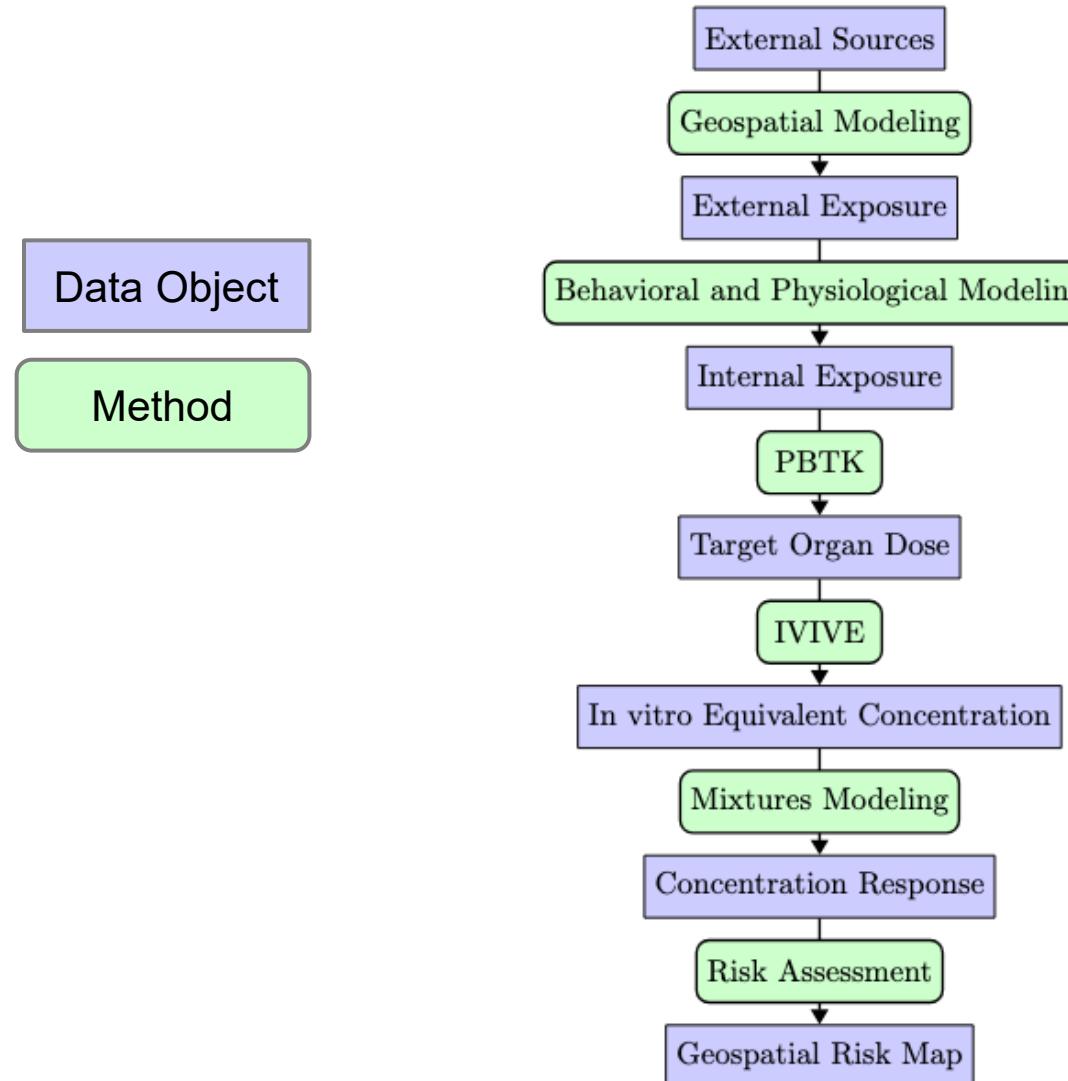
2. <http://aop.wiki.org>; Society for the Advancement of Adverse Outcome Pathways

## AEP + AOP = **GeoTox**

AEP is a comprehensive external analysis of source, media, and transformations

AOPs provide a linkage specific biological target, pathway or process by a stressor and an adverse outcome(s) considered relevant to risk assessment

# Source-to-Outcome Workflow



# Introducing the GeoTox R package



GeoTox 0.1.2

- Computational Best Practices**
  - Open Source
  - Test Driven
  - Documented
  - Extensible
- Methodology**
  - Object Oriented
  - Tidyverse, pipe-able, |>
  - Tracks exposure, population characteristics, dose-response, spatial boundaries, etc.

## GeoTox



GeoTox open-source R software package for characterizing the risk of perturbing molecular targets involved in adverse human health outcomes based on exposure to spatially-referenced stressor mixtures via the GeoTox framework - otherwise known as source-to-outcome-continuum modeling. The package, methods, and case-studies are described in [Messier, Reif, and Marvel, 2024, medRxiv-Preprint](#).

The GeoTox framework was first described in [Eccles et al. A geospatial modeling approach to quantifying the risk of exposure to environmental chemical mixtures via a common molecular target. Sci Total Environ. 2023 Jan 10;855:158905.](#)

## Installation

The package will be on CRAN in the near future - please stay tuned. You can install the development version of GeoTox from [GitHub](#) with:

```
if (!require("pak", quietly = TRUE)) {  
  install.packages("pak")  
}  
pak::pkg_install("NIEHS/GeoTox")
```

### Links

[Browse source code](#)

### License

[Full license](#)

[MIT + file LICENSE](#)

### Community

[Contributing guide](#)

[Code of conduct](#)

### Citation

[Citing GeoTox](#)

### Developers

Skylar Marvel

Author, contributor

David Reif

Author, contributor

Kyle Messier

Maintainer, author, contributor

### Dev status

# GeoTox Object

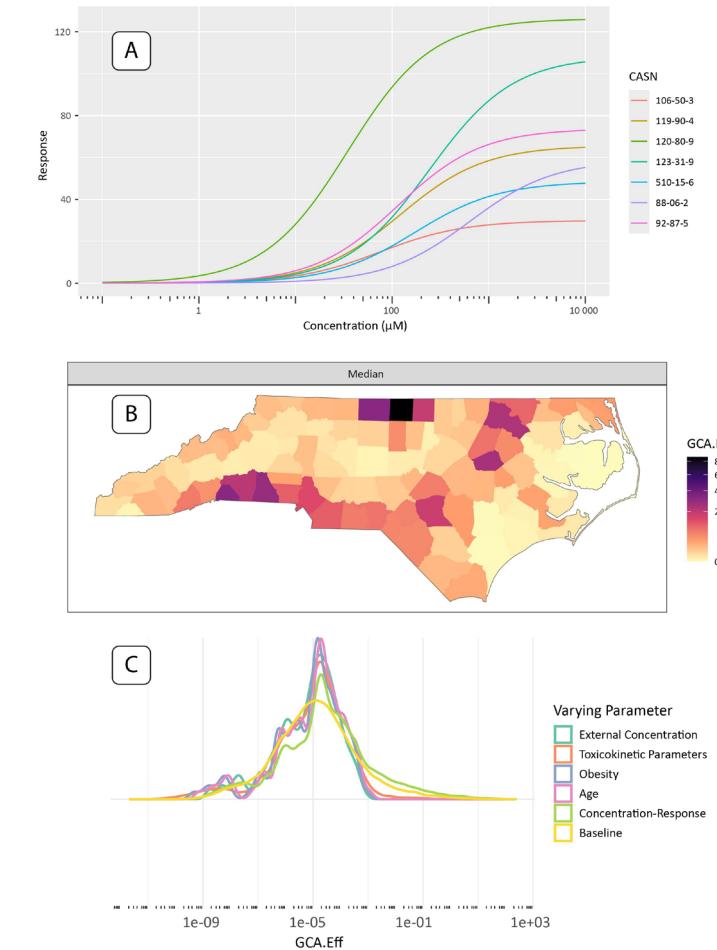
```
set.seed(2357)
geoTox <- GeoTox() |>
  # Set region and group boundaries (for plotting)
  set_boundaries(region = geo_tox_data$boundaries$county,
                 group = geo_tox_data$boundaries$state) |>
  # Simulate populations for each region
  simulate_population(age = split(geo_tox_data$age, ~FIPS),
                       obesity = geo_tox_data$obesity,
                       exposure = split(geo_tox_data$exposure, ~FIPS),
                       simulated_css = geo_tox_data$simulated_css,
                       n = n) |>
  # Estimated Hill parameters
  set_hill_params(geo_tox_data$dose_response |>
    fit_hill(assay = "endp", chem = "casn") |>
    filter(!tp.sd.imputed, !logAC50.sd.imputed)) |>
  # Calculate response
  calculate_response() |>
  # Perform sensitivity analysis
  sensitivity_analysis()

geoTox
#> GeoTox object
#> Assays: 13
#> Chemicals: 20
#> Regions: 100
#> Population: 250
#> Data Fields:
#>   Name          Class          Dim
#>   age    list(integer)  100 x (250)
#>   IR     list(numeric)  100 x (250)
#>   obesity list(character) 100 x (250)
#>   C_ext   list(matrix)  100 x (250 x 21)
#>   C_ss    list(matrix)  100 x (250 x 21)
#> Computed Fields:
#>   Name          Class          Dim
#>   D_int    list(matrix)  100 x (250 x 21)
#>   C_invitro list(matrix)  100 x (250 x 21)
#>   resp     list(data.frame) 100 x (3250 x 6)
#>   sensitivity list(list)  5 x (100)
#> Other Fields: par, boundaries, exposure, css_sensitivity, hill_params
```

# Geospatial Risk Mapping of Chemical Mixtures

## Example 1

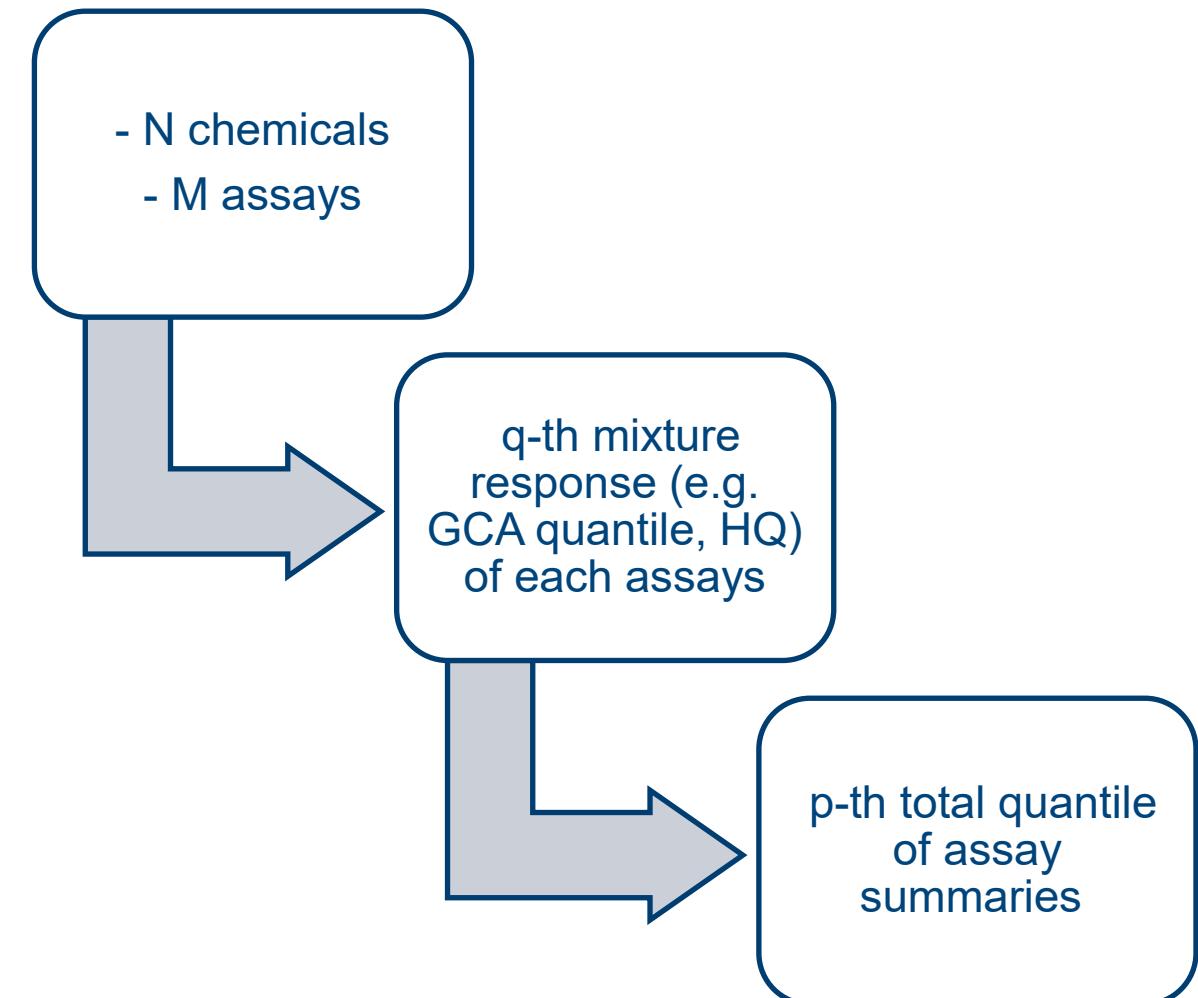
- 7 chemicals in air
- 1 Assay: H2AX Histone Modification
- Generalized Concentration Addition
- Mapped risk as quantified by assay response



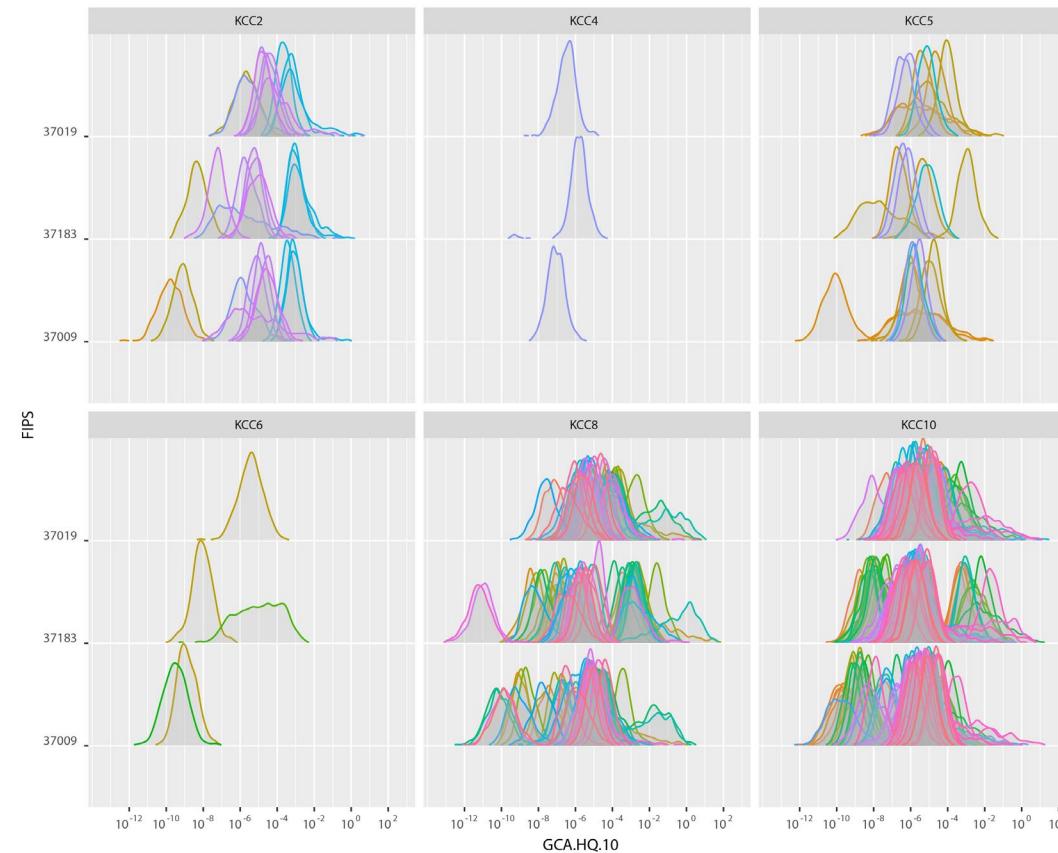
# Multi-Assay Risk Mapping of Chemical Mixtures

## Example 2

- 40+ chemicals in air
- 200+ Assays Based on Key Characteristics of Carcinogens (KCC)
- GCA, IA, Hazard Quotient
- “ $p$ -th total quantile of the  $q$  assay-level quantiles”

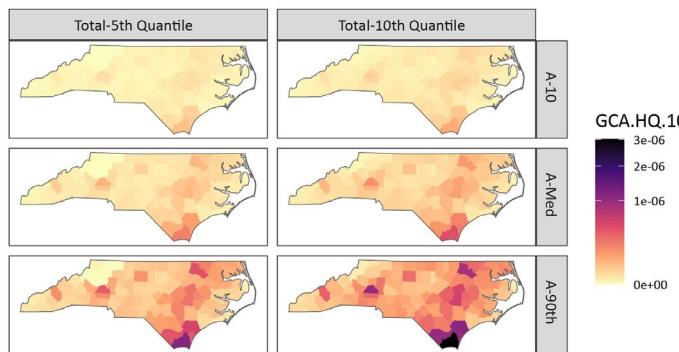


# Multi-Assay Risk Mapping of Chemical Mixtures

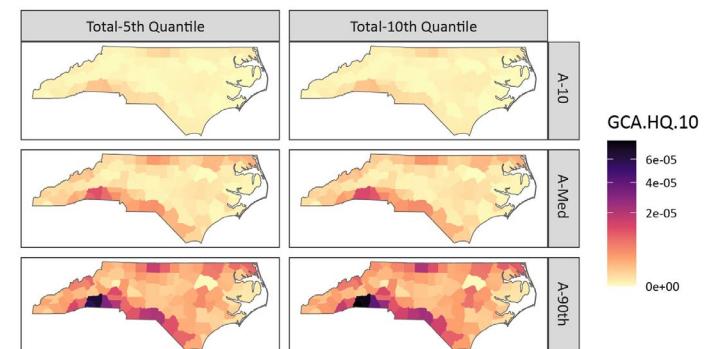


# Multi-Assay Risk Mapping of Chemical Mixtures

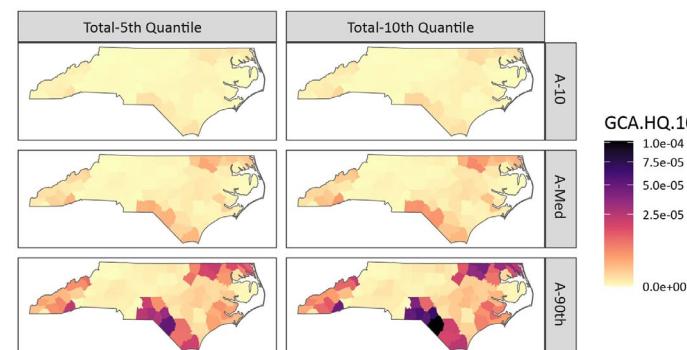
Multi-Assay Summaries: All KCC Assays



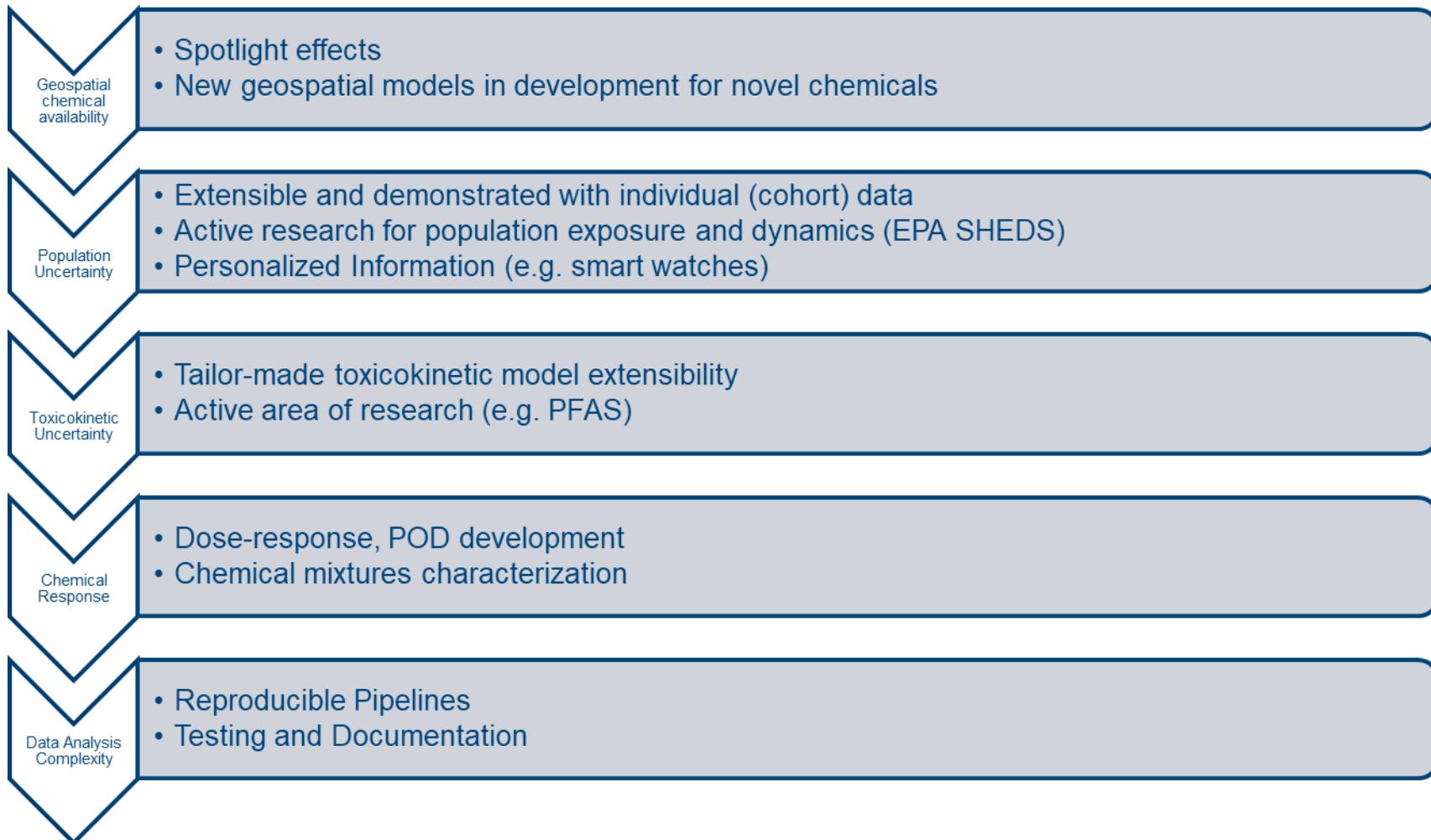
KCC5: Oxidative Stress



KCC2: Genotoxic Effects



## Limitations + Future Directions



## Software and Computational Best-Practices

Test Driven Development

Continuous Integration

Build Checks

Style / Linting

Workflows / Pipelines

# Best Practices are Needed for Complex Environmental Health Pipelines



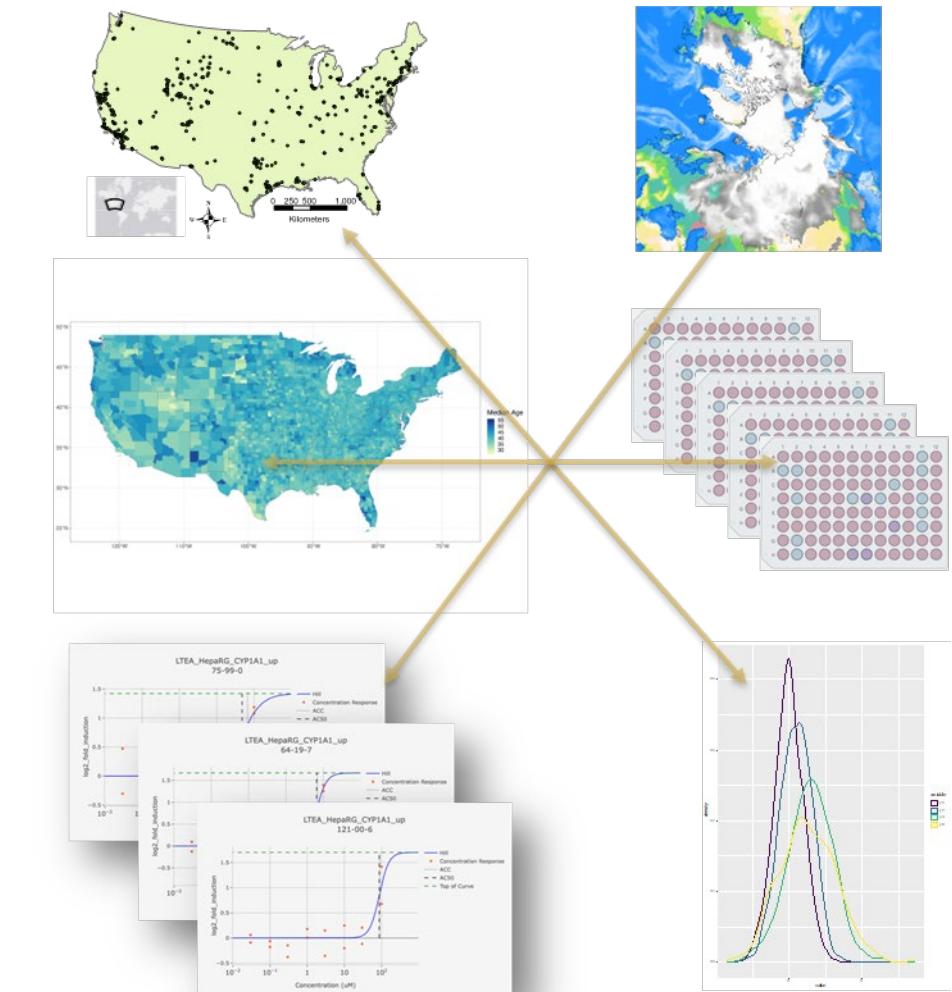
Geospatial modeling and GeoTox offer a tractable approach for quantifying the exposome health impacts



Need best practices to build towards a very complex analysis and understanding



(1) In-Situ Monitoring Data (2) Atmospheric / Geophysical Data (3) Census Data (4) In-Vitro Screening Data (5) Concentration-Response Modeling (6) Probabilistic Models (7) ...



## Acknowledgements

- Co-authors on GeoTox package and manuscript: David Reif & Skylar Marvel
- Lead author on GeoTox concept paper and former postdoc: Kristin Eccles
- SET group member: Eva Marques, Mariana Alifa, Mitchell Manware
- NIEHS Computational Support: Office of Scientific Computing





JOHNS HOPKINS  
WHITING SCHOOL  
*of* ENGINEERING



JOHNS HOPKINS  
BLOOMBERG SCHOOL  
*of* PUBLIC HEALTH

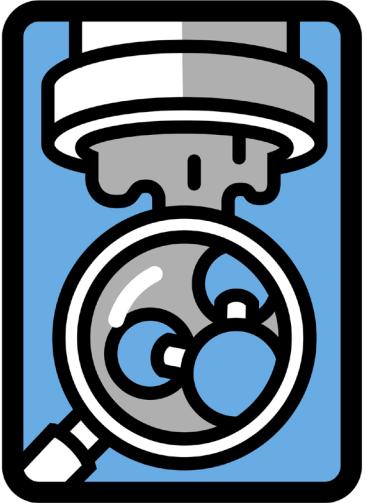
# Human and Environmental Exposure Framework for Biosolids

Carsten Prasse, Ph.D.

Department of Environmental Health & Engineering,  
Johns Hopkins University

- **Disclaimer:** The views expressed in this presentation are those of the authors and do not necessarily reflect the views or policies of the U.S. Environmental Protection Agency. Any use of trade, firm, or product names is for descriptive purposes only and does not imply endorsement by the U.S. Government.

# Funding



## **Fate and Transport of Unregulated Organic Contaminants in Biosolids: Development of a Human and Environmental Exposure Risk Framework**

EPA Grant: R840247



# Project Team



**Principal Investigator (PI):** Carsten Prasse

**Co-PI:** Keeve Nachman

Thomas Burke

Sara Lupolt

Matthew Newmeyer

Riley Demo

Noor Hamdan

Kate Burgener

Dominic Sanchez

**EPA Collaborators:** Antony Williams

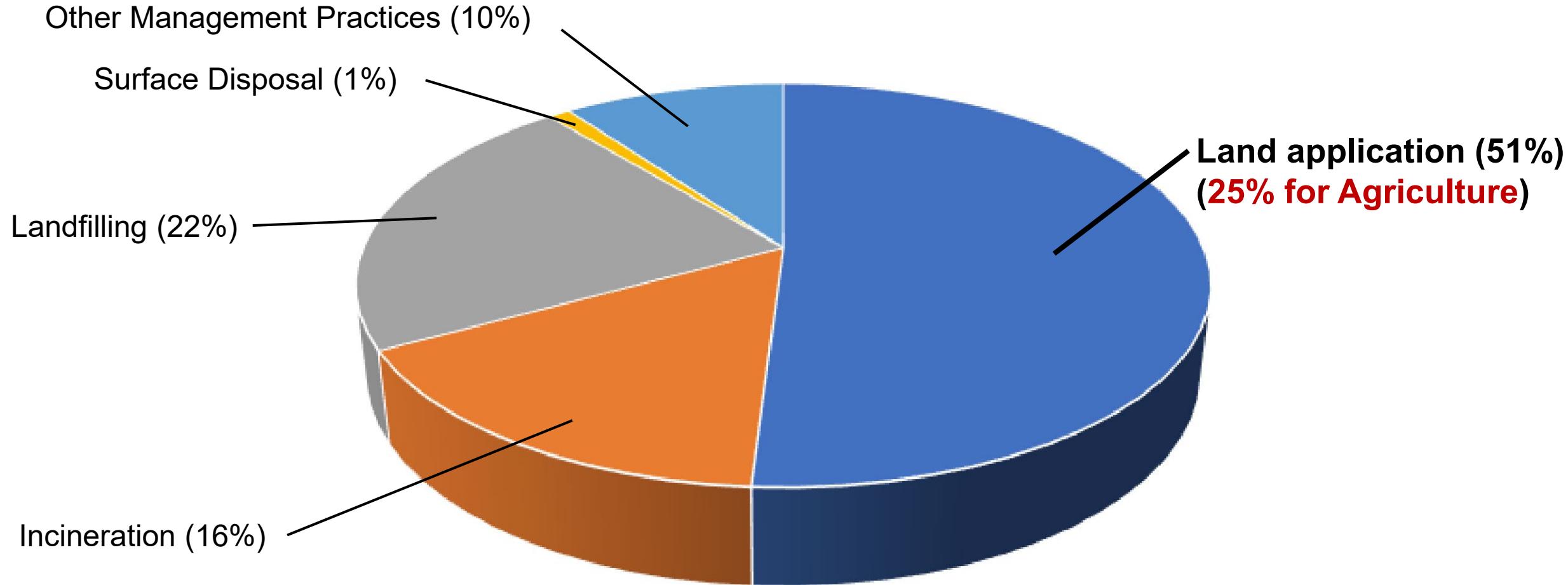
Jon Sobus

# What are biosolids?

→ solid byproduct of sewage treatment processes



# 4.75 million dry metric tons of biosolids are generated each year



# Land application of biosolids



Biosolid land application can be beneficial:

- ✓ Improves soil qualities
- ✓ Supplies nutrients
- ✓ Diverts from landfilling & incineration

Current federal regulation (40 CFR Part 503) of biosolid quality includes:

- Limits on 10 heavy metals
- Requirements for pathogen & vector attraction reduction

**→ Currently, no organic contaminants are regulated in biosolids**

# Example of organic contaminants in biosolids: PFAS

 The New York Times

## Something's Poisoning America's Land. Farmers Fear 'Forever Chemicals.'

Fertilizer made from city sewage has been spread on millions of acres of farmland for decades. Scientists say it can contain high levels of...

Aug 31, 2024



 Nebraska Public Media

## This farmer's livelihood was ruined by PFAS-contaminated fertilizer that few Midwest states test for

Biosolids — a type of treated sewage byproduct from wastewater treatment plants — are used as a nutrient-rich fertilizer on farms across the...



Mar 11, 2024



 The Guardian

## Texas farmers claim company sold them PFAS-contaminated sludge that killed livestock

Two ranches also allege biosolids with 'forever chemicals' ruined crops, polluted drinking water and left their properties worthless.

Mar 1, 2024



 Chemical & Engineering News

## PFAS in biosolids prompt lawsuits

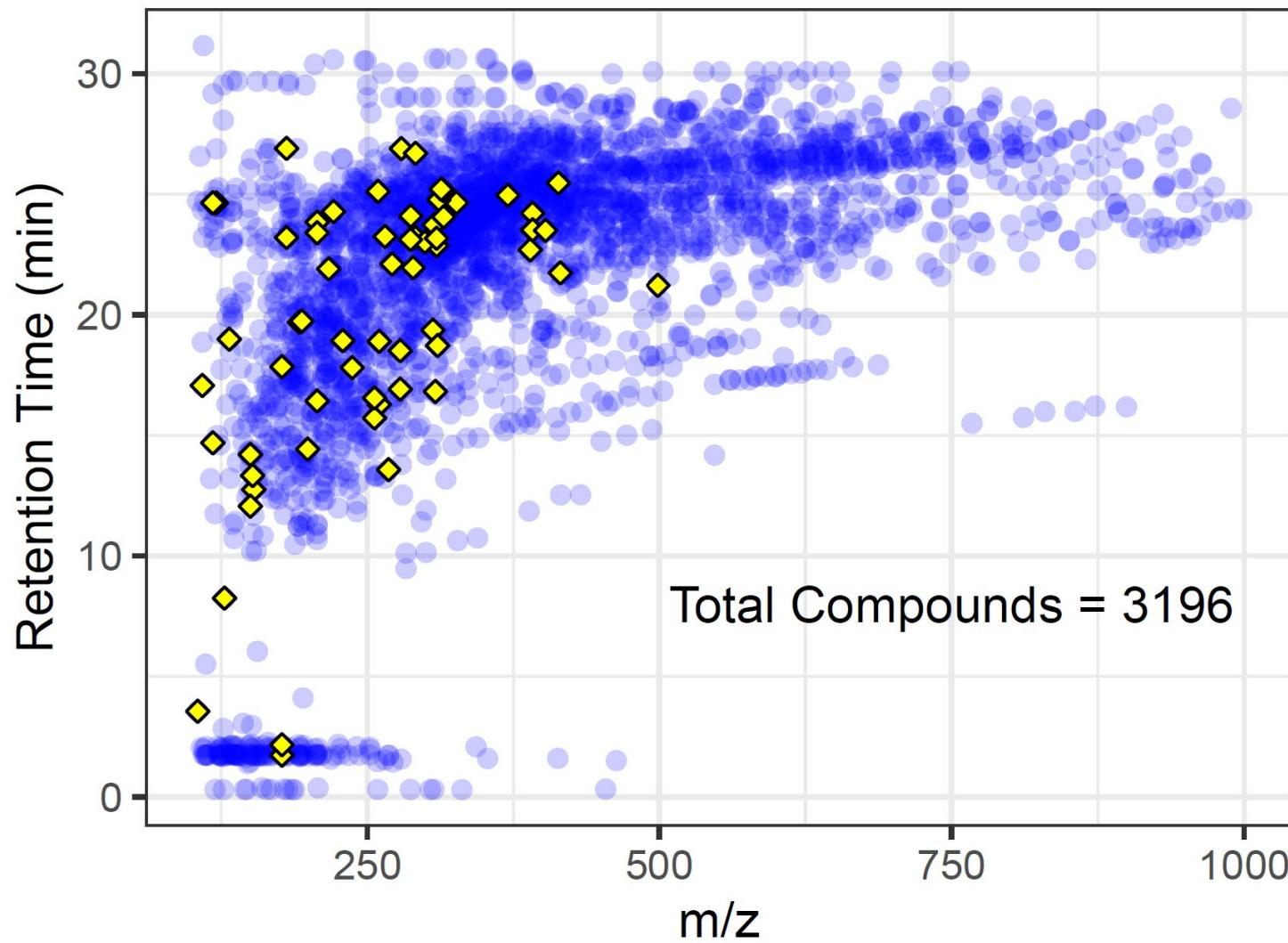
The lawsuit alleges that Synagro failed to warn product users about the adverse health effects associated with exposure to PFAS. Some of the...



Feb 28, 2024

# Biosolids contain a large number of unknown organic compounds

(e.g. pharmaceuticals, industrial chemicals, pesticides, naturally occurring compounds, ...)



< 5% of compounds are known  
(highlighted in yellow; based on  
EPA biosolids list)



High-resolution mass spectrometry  
system available in Prasse lab.

**Biosolids contain a large number of unknown organic compounds**  
(e.g. pharmaceuticals, industrial chemicals, pesticides, naturally occurring compounds, ...)



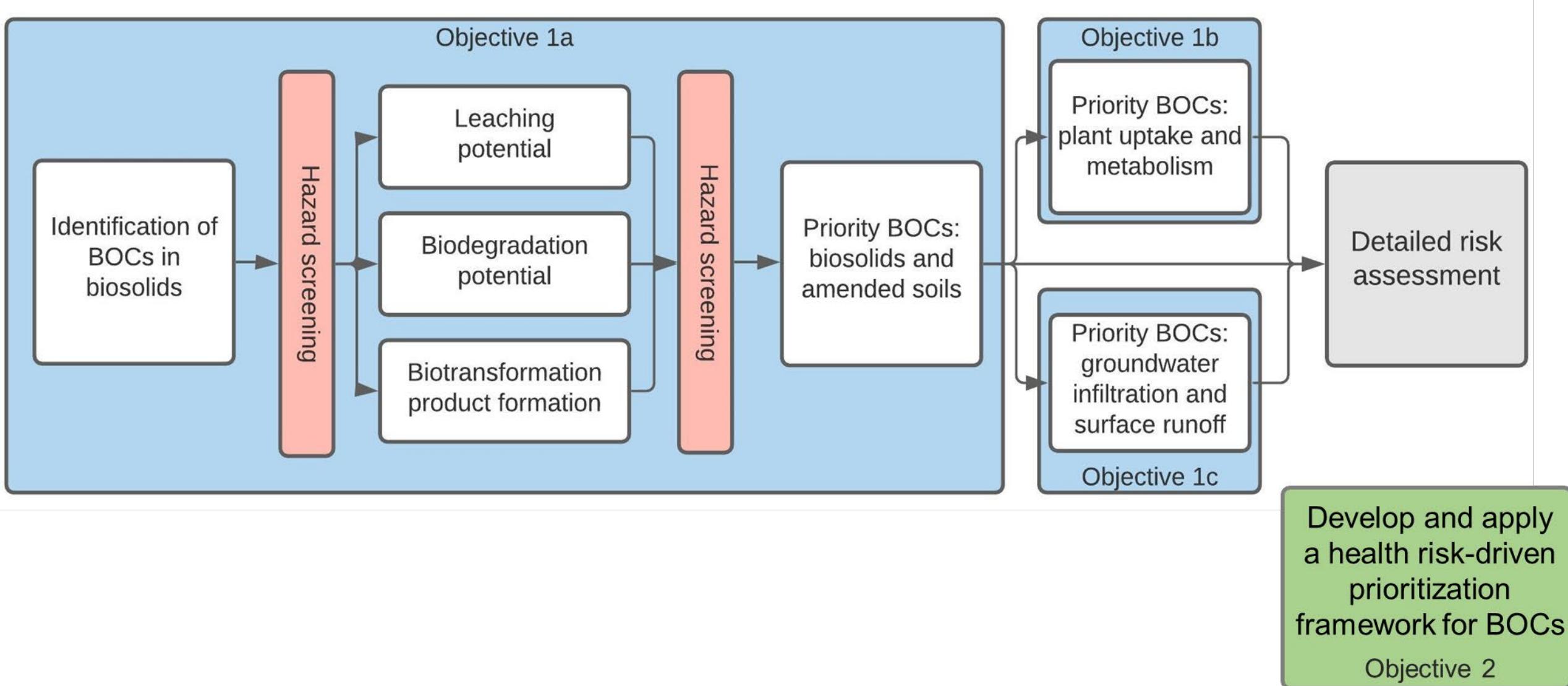
< 5% of compounds are known  
(highlighted in yellow; based on  
EPA biosolids list)

**We need to develop approaches that aid in the identification of **toxic** compounds in complex mixtures**

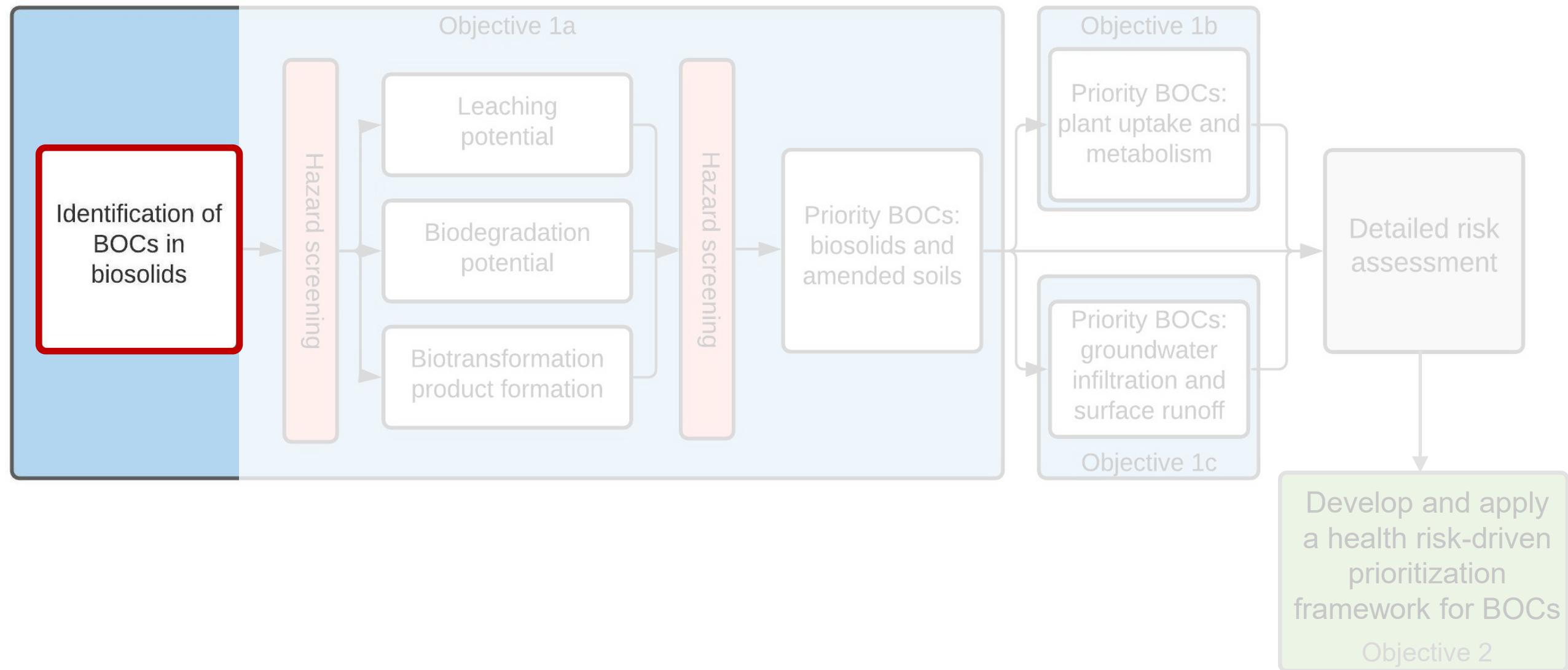


High-resolution mass spectrometry system available in Prasse lab.

# Characterize the occurrence, fate, transport and risks of novel biosolid-associated contaminants (BOCs)

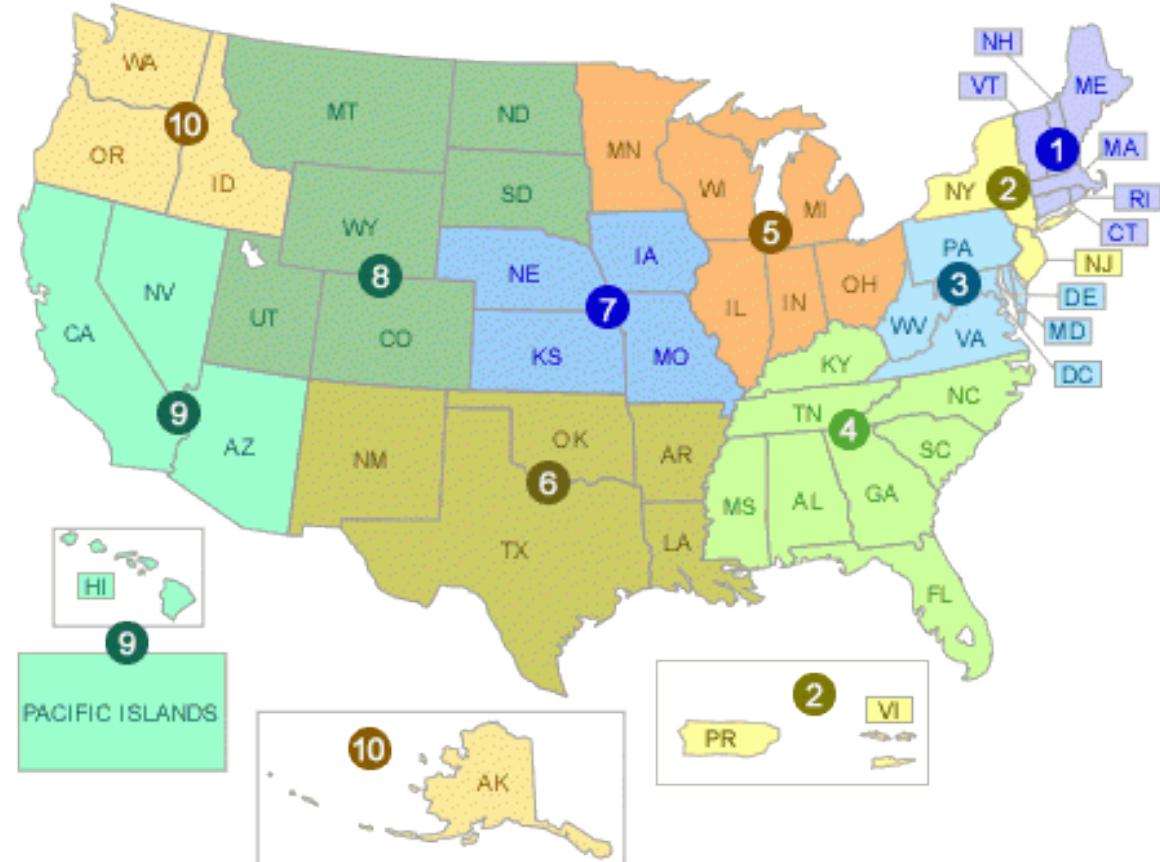


# Characterize the occurrence, fate, transport and risks of novel biosolid-associated contaminants (BOCs)



# Biosolid sampling

- 16 samples across U.S. and Canada:
  - U.S.: 13 samples
    - Region 2: 2 samples
    - Region 3: 3 samples
    - Region 5: 2 samples
    - Region 7: 2 samples
    - Region 9: 4 samples
  - Canada: 3 samples

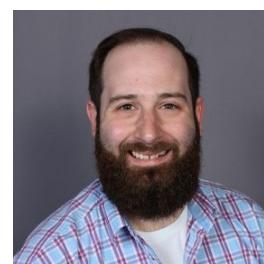


## Regional and Geographic Offices.

<https://www.epa.gov/aboutepa/regional-and-geographic-offices>. Accessed 2023 Aug 08

Interested in compounds present in > 80% of samples

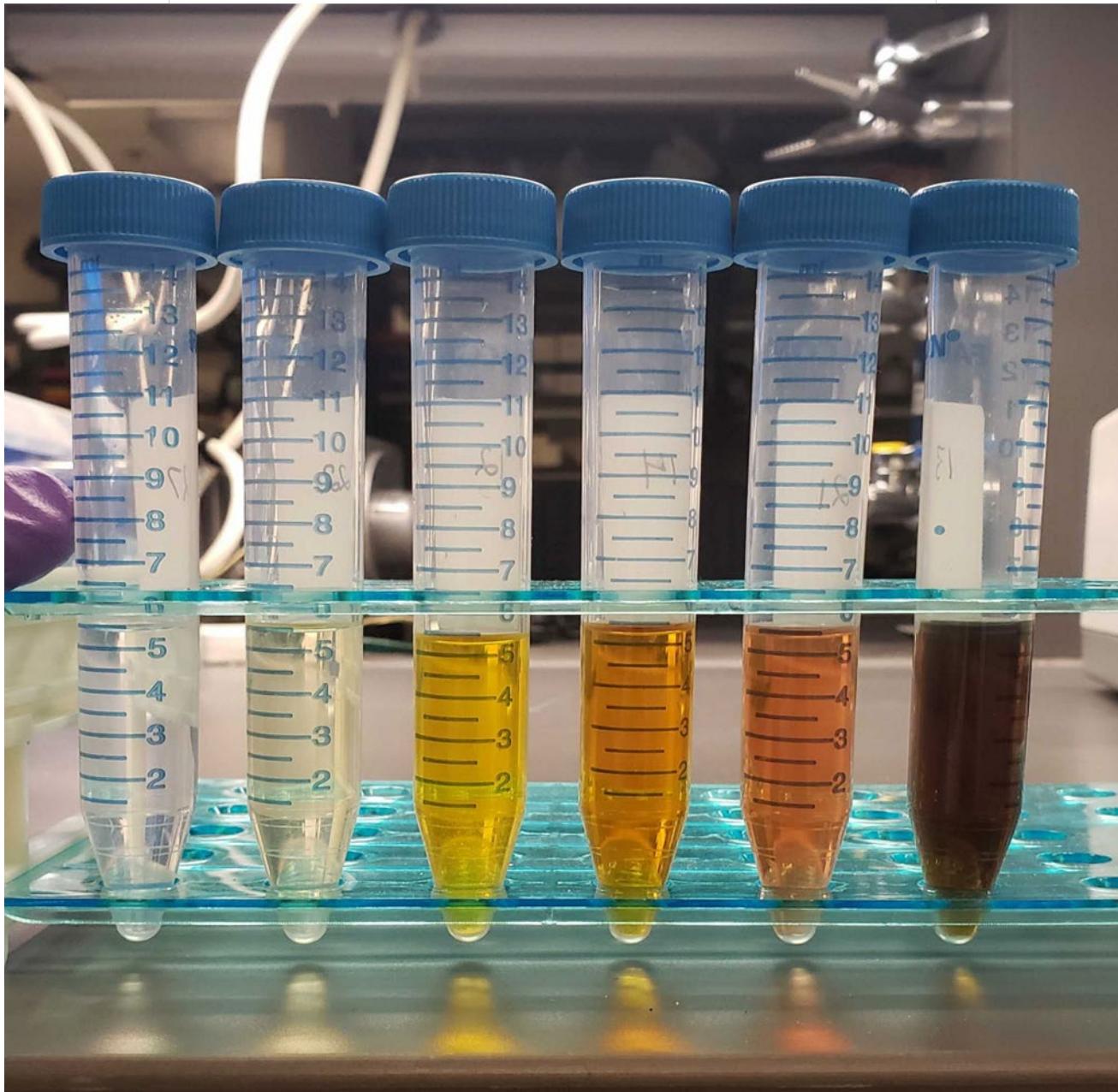
# Biosolid preparation and analysis



- Specimens were extracted via QuEChERS with dSPE
- Extracts analyzed via LC-HRMS
  - Positive: A) 1 mM ammonium fluoride, B) 0.1% formic acid in methanol
  - Negative: A) 1 mM ammonium fluoride, B) acetonitrile
  - Full Scan/data-dependent MS<sup>2</sup> (top 10)



## After dSPE and centrifuging

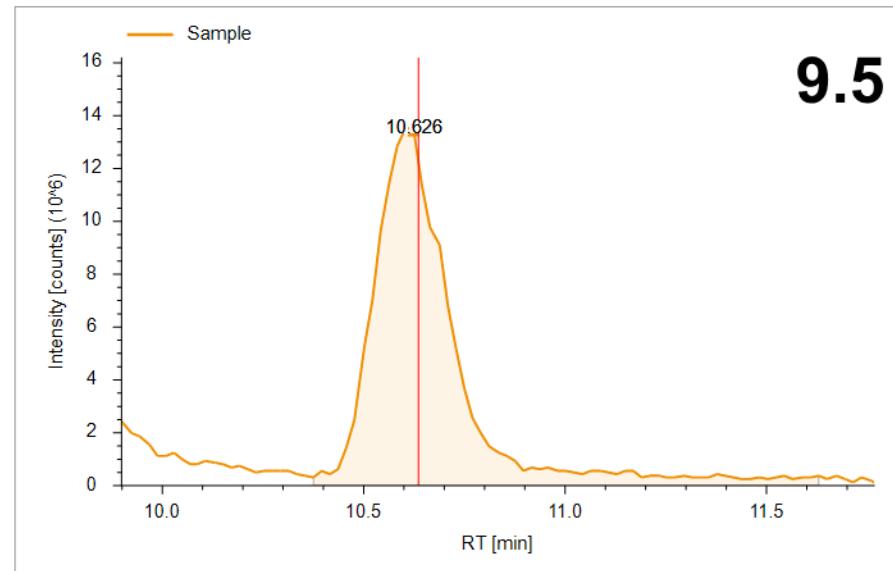
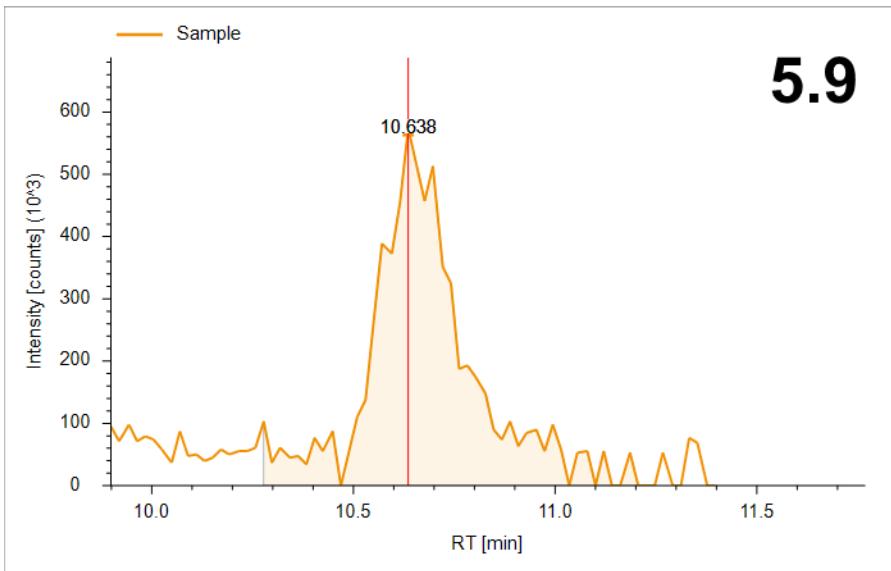
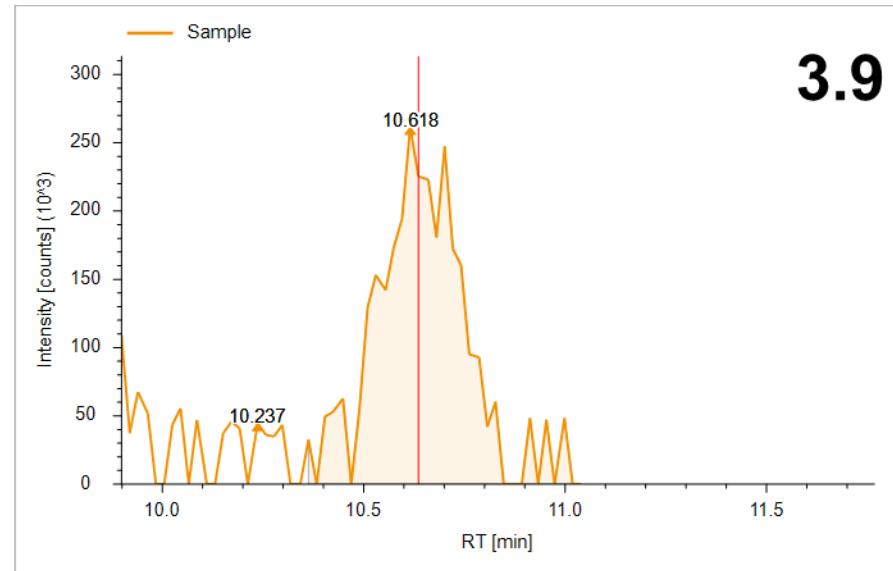
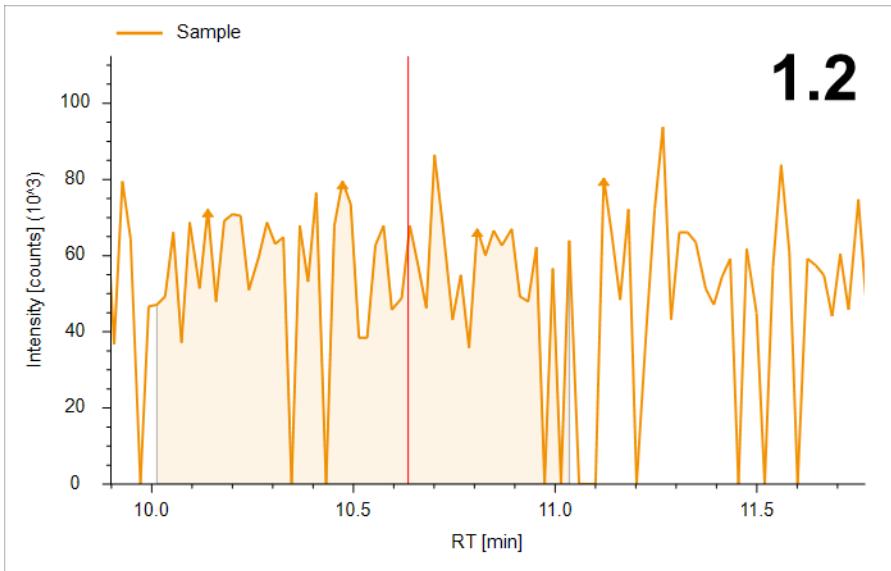


# LC-HRMS data analysis



- Perform Quality Control (QC)-based peak area normalization
  - Detection in  $\geq 50\%$  pooled QCs (pooled biosolid samples)
  - %RSD before normalization  $\leq 30\%$
  - %RSD after normalization  $\leq 25\%$
- Detected in all pooled QC replicates
- Peak rating  $\geq 6$

# Peak rating examples for the same feature in different samples (scored on a scale from 0-10)



# LC-HRMS data analysis



- Perform Quality Control (QC)-based peak area normalization
  - Detection in  $\geq 50\%$  pooled QCs (pooled biosolid samples)
  - %RSD before normalization  $\leq 30\%$
  - %RSD after normalization  $\leq 25\%$
- Detected in all pooled QC replicates
- Peak rating  $\geq 6$
- **Detected in at least 80% of biosolid samples**

# Feature counts after different processing steps

# Features originally detected

71,651

# Features that fulfilled all QC-based filter criteria

792

# Features remaining after review and removing background and low-quality peaks, in-source fragments, and isotopes

451

# Confidence Level assignment (Schymanski criteria)

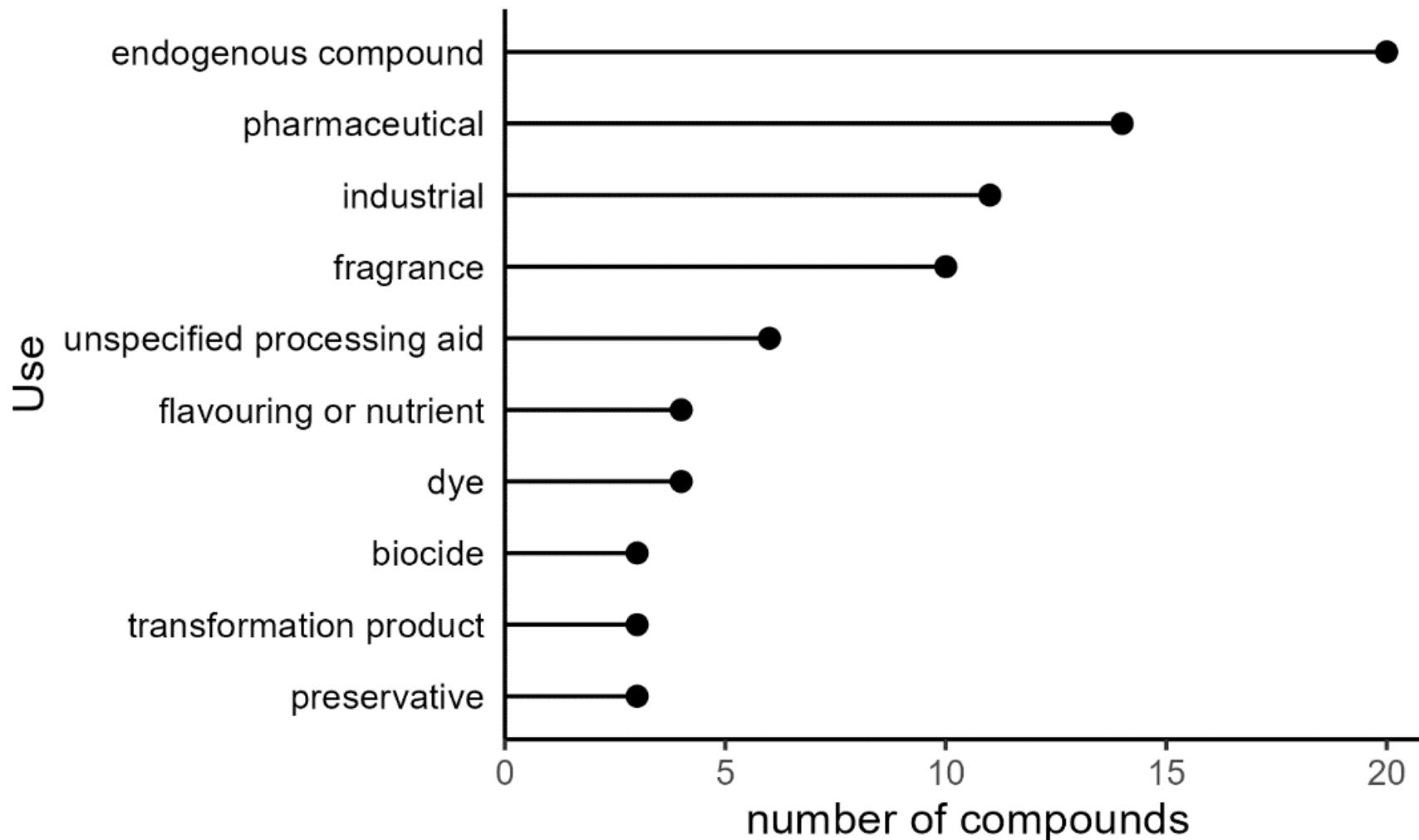
Confidence Level	Notes
Level 1	RT and MS <sup>2</sup> match between sample and reference standard
Level 2 – mzCloud	Spectral library match with score $\geq 70$
Level 2 – MoNA	Spectral library match with score $\geq 70$ and no mzCloud match
Level 3	Multiple plausible spectral library matches with score $\geq 70$
Level 4	<ul style="list-style-type: none"><li>-No spectral library matches</li><li>-Single predicted formula from CD also found in ChemSpider</li></ul>
Level 5	<ul style="list-style-type: none"><li>-No spectral library matches</li><li>-Either:<ul style="list-style-type: none"><li>-None of the CD predicted formulas in ChemSpider</li><li>-Multiple predicted formulas from CD also found in ChemSpider</li></ul></li></ul>

# Confidence Level assignment (Schymanski criteria)

Confidence Level	Notes
Level 1	RT and MS <sup>2</sup> match between sample and reference standard
Level 2 – <u>mzCloud</u>	Spectral library match with score $\geq 70$
Level 2 – <u>MoNA</u>	Spectral library match with score $\geq 70$ and no <u>mzCloud</u> match

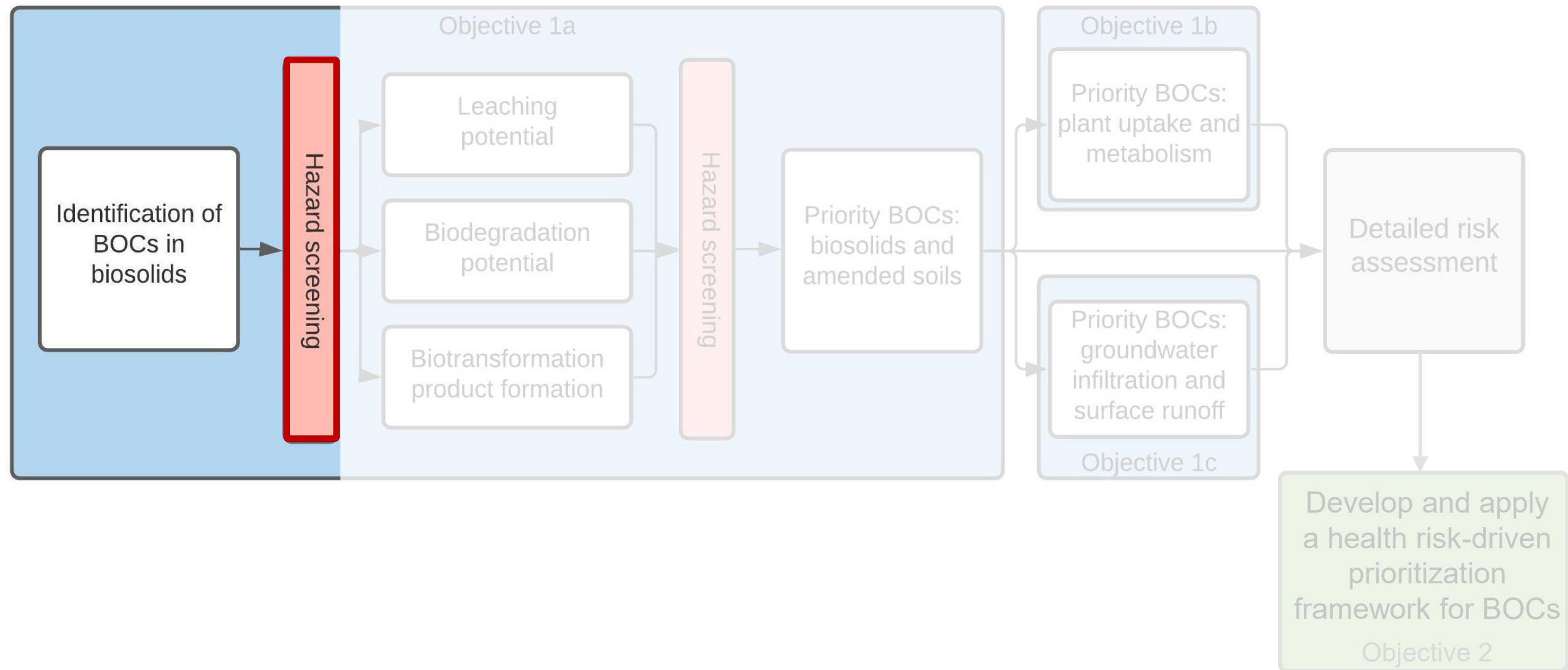
- **92 compounds** assigned confidence Level 1 & 2
  - **58/92 compounds** reported for the first time in biosolids
  - **77.4% of compounds** have an unknown structure (Level 3-5)

# Top 10 use categories of detected compounds



Source: Harmonized Functional Use data from the Chemicals and Products Database (CPDat)  
via EPA's ChemExpo site ([https://comptox.epa.gov/chemexpo/get\\_data/](https://comptox.epa.gov/chemexpo/get_data/))

# Characterize the occurrence, fate, transport and risks of novel biosolid-associated contaminants (BOCs)



# EPA Cheminformatics Hazard Comparison Module (HCM)

Cheminformatics Modules  
version: DEV, build: 2023-03-09 06:08:29 UTC

HAZARD ALERTS PREDICT 2.0 SEARCH STANDARDIZE TOXPRINTS

Full ▼ ▼ ▼ ▼

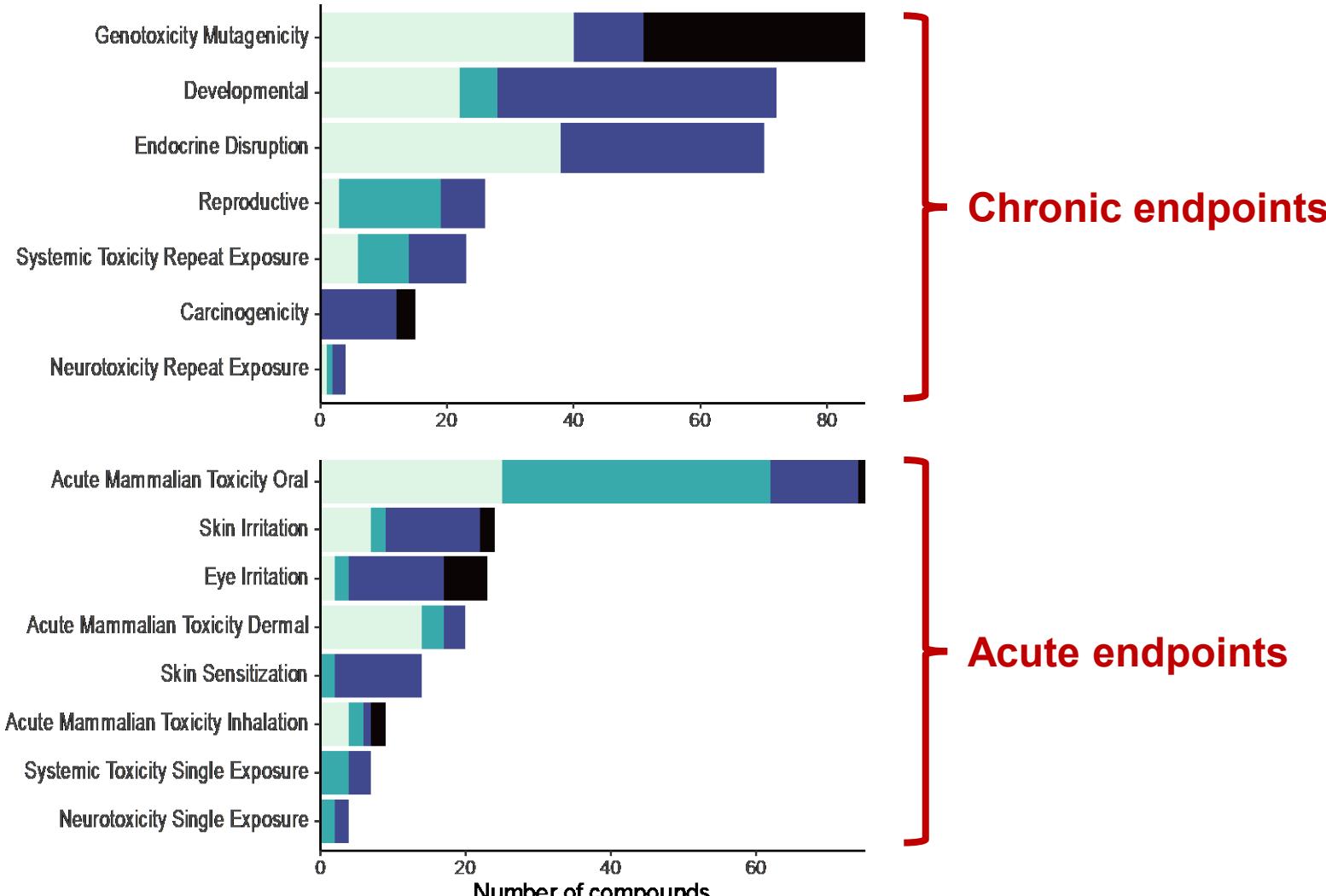
Chemicals: 32

Toxicity: VH - Very High H - High M - Medium L - Low I - Inconclusive N/A - Not Applicable Authority: Authoritative Screening QSAR Model

CAS Name	Human Health Effects												Ecotoxicity	Fate				
	Acute Mammalian Toxicity			Carcinogenicity	Genotoxicity Mutagenicity	Endocrine Disruption	Reproductive	Developmental	Neurotoxicity		Systemic Toxicity							
	Oral	Inhalation	Dermal						Repeat Exposure	Single Exposure	Repeat Exposure	Single Exposure				Skin Sensitization	Eye Irritation	Acute Aquatic Toxicity
59-31-4 2(1H)-Quinolinone	M			L	L		H					H	H	L		L	I	
95-14-7 1,2,3-Benzotriazole	M	VH	M	H	VH	L	I	M	I	L	M	I	L	H	M	L	M	H
59729-33-8 Citalopram	M			L	H		H							H		M	M	I

# Hazard assessment – Biosolid samples

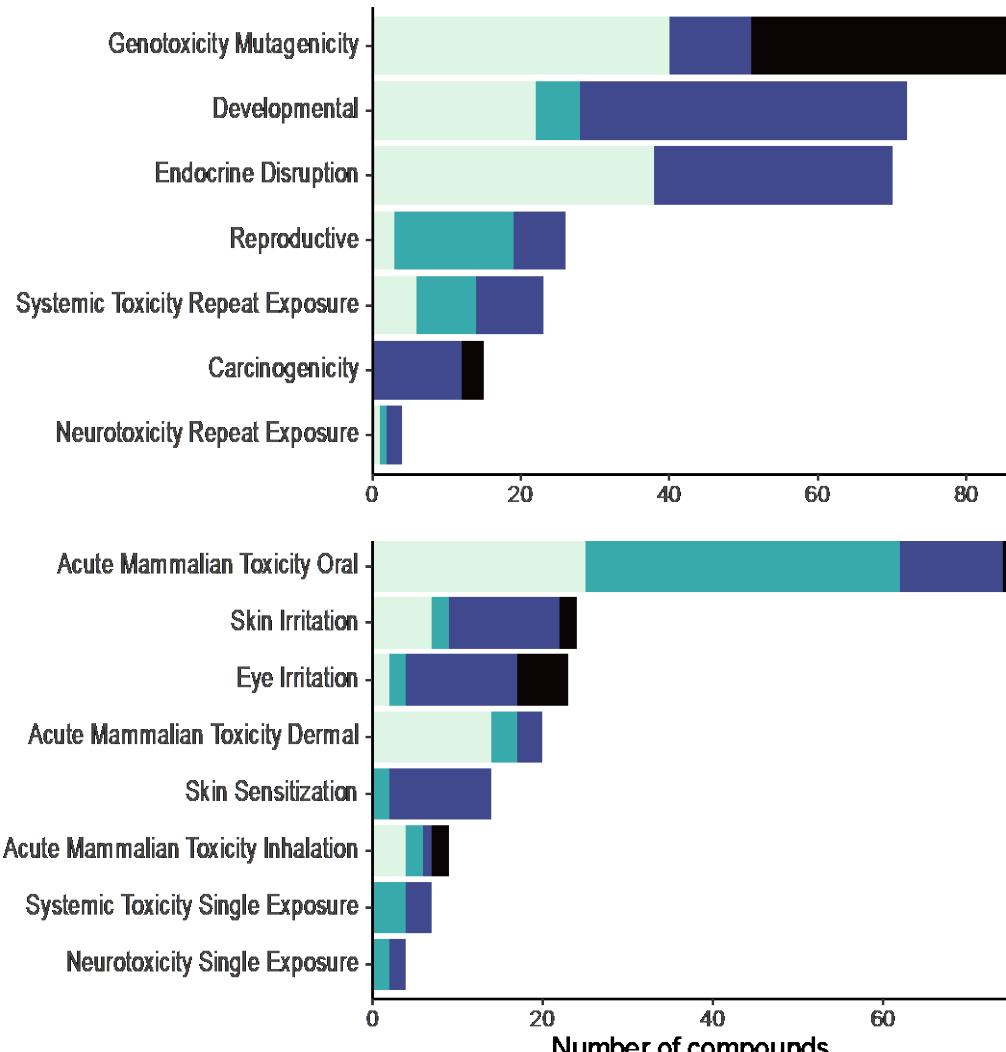
Categorization based on hazard score



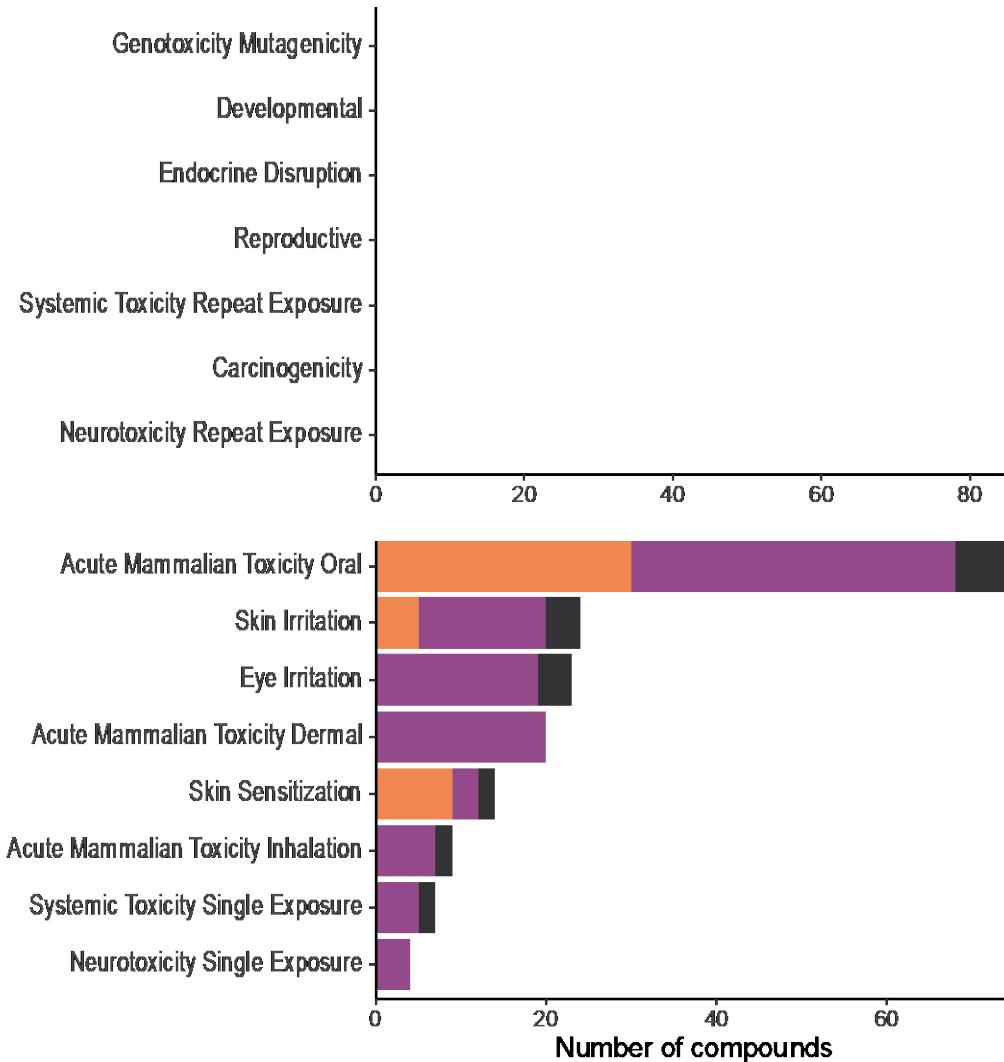
Hazard score    █ Very high    █ High    █ Medium    █ Low

# Hazard assessment – Biosolid samples

Categorization based on hazard score



Categorization based on data source authority

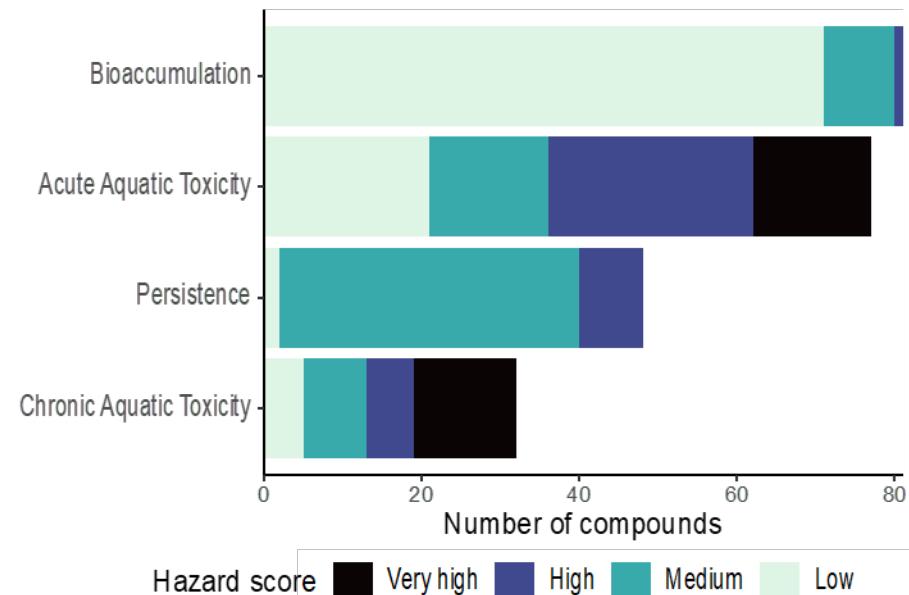


Hazard score    █ Very high    █ High    █ Medium    █ Low

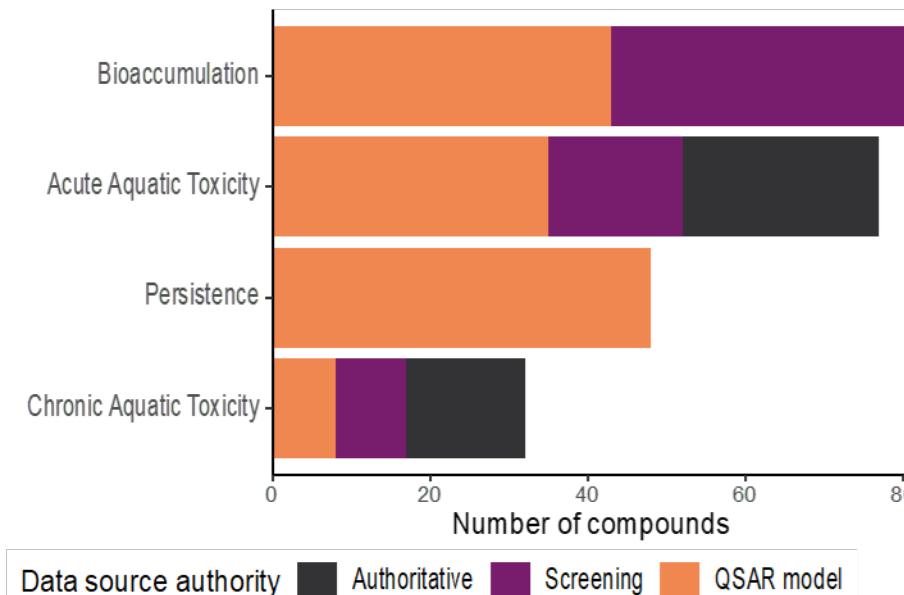
Data source authority    █ Authoritative    █ Screening    █ QSAR model

# Hazard assessment – Biosolid samples

Categorization based on hazard score



Categorization based on data source authority



# Compound prioritization approach

$$Avg. Hazard score = \frac{\sum \text{individual hazard scores}}{\text{number of endpoints with data available}}$$

Hazard score categories: low (1), medium (2), high (3), very high (4)

# Compound prioritization approach

$$\begin{aligned} \text{Avg. Hazard score} &= \frac{\sum \text{individual hazard scores}}{\text{number of endpoints with data available}} \\ \text{Avg. Quality score} &= \frac{\sum \text{individual quality scores}}{\text{number of endpoints with data available}} \end{aligned}$$

Data quality score categories: QSAR (1), screening (2), authoritative (3)

# Compound prioritization approach

$$Avg. Hazard score = \frac{\sum \text{individual hazard scores}}{\text{number of endpoints with data available}}$$

$$Avg. Quality score = \frac{\sum \text{individual quality scores}}{\text{number of endpoints with data available}}$$

$$Quality adjusted hazard score = Avg. Hazard score * Avg. Quality Score$$

# Compound prioritization approach

$$\text{Avg. Hazard score} = \frac{\sum \text{individual hazard scores}}{\text{number of endpoints with data available}}$$

$$\text{Avg. Quality score} = \frac{\sum \text{individual quality scores}}{\text{number of endpoints with data available}}$$

$$\text{Quality adjusted hazard score} = \text{Avg. Hazard score} * \text{Avg. Quality Score}$$

$$\text{Completeness score} = \frac{\text{number of endpoints with data available}}{\text{number of endpoints searched}}$$

# Compound prioritization approach - Example

Compound	Endpoint A	Endpoint B	Endpoint C	Hazard Score	Quality Score	Completeness Score	Quality-adjusted Hazard Score
1	VH Authoritative	H Screening	L QSAR	2.67	2.00	1.00	5.34
2	H QSAR	L Authoritative	M Screening	2.00	2.00	1.00	4.00
3	L QSAR		L Screening	1.00	1.00	0.67	1.00

# Hazard prioritization results

## Top 5 Prioritized Compounds for Each End Point Group Based on Quality-Adjusted Hazard Scores

compound (confidence level)	quality-adjusted hazard score	completeness group <sup>a</sup>	use <sup>b</sup>
Human Health Effects—chronic Exposure (7 End Points)			
ketoconazole (2b)	8.01	medium low	pharmaceutical
<i>p</i> -cresol (2a)	7.19	high	fragrance
4-androstene-3,17-dione (2a)	6.99	medium low	endogenous
clorophene (2a)	6.72	medium high	biocide
phenolphthalein (2a)	6.68	high	not specified
Human Health Effects—short-term Exposure (8 End Points)			
clorophene (2a)	7.56	high	biocide
<i>p</i> -cresol (2a)	7.14	high	fragrance
indole (2a)	6.99	medium low	fragrance
thymol (1)	6.88	medium high	fragrance
ketoconazole (2b)	6.25	medium low	pharmaceutical
Ecological End Points (4 End Points)			
fludioxonil (2a)	12.00	medium high	biocide
ketoconazole (2b)	8.01	high	pharmaceutical
4-(1,1,3,3-tetramethylbutyl)phenol (2a)	6.75	high	adhesion promoter
triclocarban (2a)	6.75	high	preservative
carbamazepine (1)	6.22	high	pharmaceutical

# Hazard prioritization results

**Table 2. Top 5 Prioritized Compounds for Each End Point Group Based on Quality-Adjusted Hazard Scores**

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# Target list for further in-depth investigations

- Triethyl phosphate

- Tris(2-chloroethyl) phosphate

- Tris(2-chloroisopropyl)phosphate

- Triphenyl phosphate

Flame Retardants



- 4-Methylphenol

- C12 BAC

- C12 DADMAC

- Clorophene

- Methylparaben

- Triclocarban

- Triclosan

- Thymol

Antimicrobials



- 2-Naphthol

- 4-Nonylphenol

- N,N'-Diphenylguanidine

- Isoquinoline

Industrial Chemicals



- Androstenedione

- Caffeine

- Indole

- Norharman

- Piperine

Natural Products



- Avobenzone

- Benzotriazole

- Octinoxate

- Octocrylene

- Oxybenzone

UV Filters



- Amitriptyline

- Carbamazepine

- Citalopram

- Diphenhydramine

- Fluoxetine

- Lamotrigine

- Metoprolol

- Spectinomycin

- Trazodone

Pharmaceuticals



- Atrazine

- Fludioxonil

- Ketoconazole

- Thiabendazole

Other Biocides



- Bisphenol A

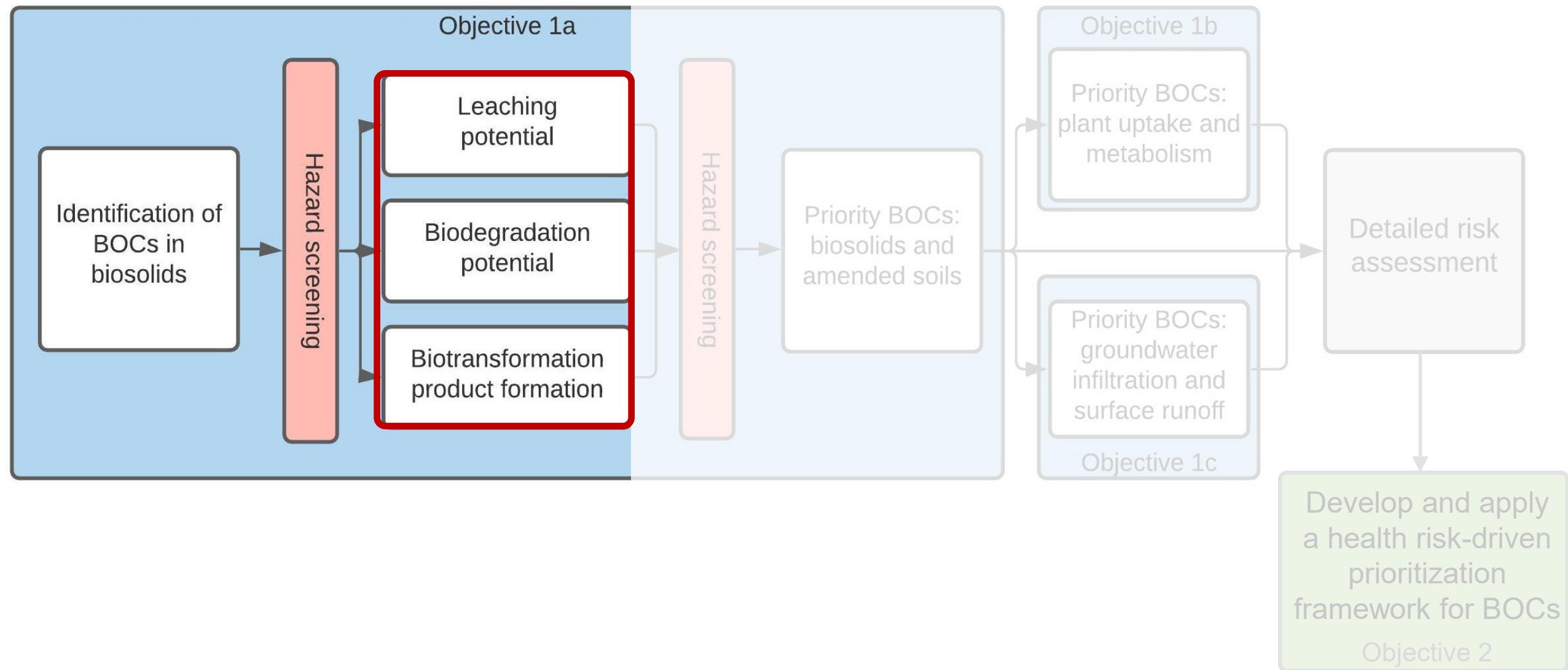
- Bisphenol F

- Bisphenol S

Plasticizers



# Characterize the occurrence, fate, transport and risks of novel biosolid-associated contaminants (BOCs)



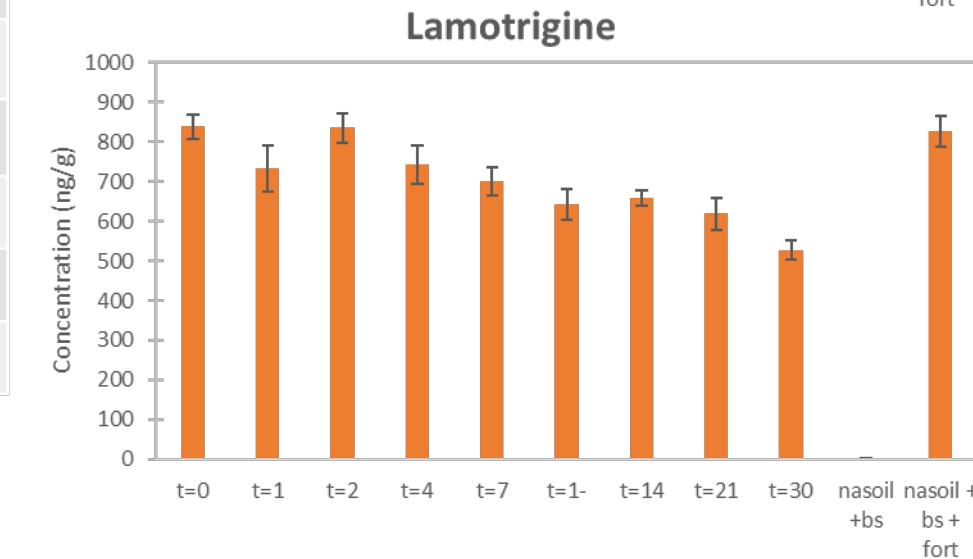
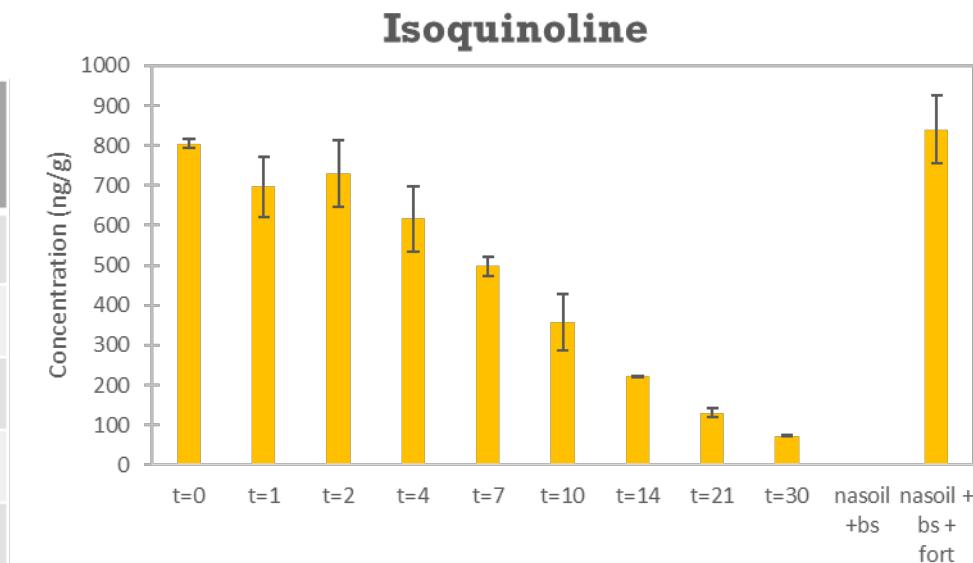
# Laboratory experiments - Biodegradation



## 30 day soil + biosolids batch experiments:

Compound	% Degradation	Compound	% Degradation
4-Methylphenol	100%	Triclosan	41.7%
2-Naphthol	98.5%	Bisphenol S	39.9%
Androstanedione	92.6%	Lamotrigine	37.2%
Isoquinoline	90.9%	Piperine	33.5%
Bisphenol F	79.9%	Caffeine	27.5%
Oxybenzone	78.4%	Tris(2-chloroethyl)phosphate	17.1%
Bisphenol A	74.5%	1,3-diphenylguanidine	15.9%
Ketoconazole	69.9%	Amitriptyline	15.8%
Clorophene	56.8%		
Trazodone	51.7%		

→ among 31 compounds tested,  
18 degraded by >15 %



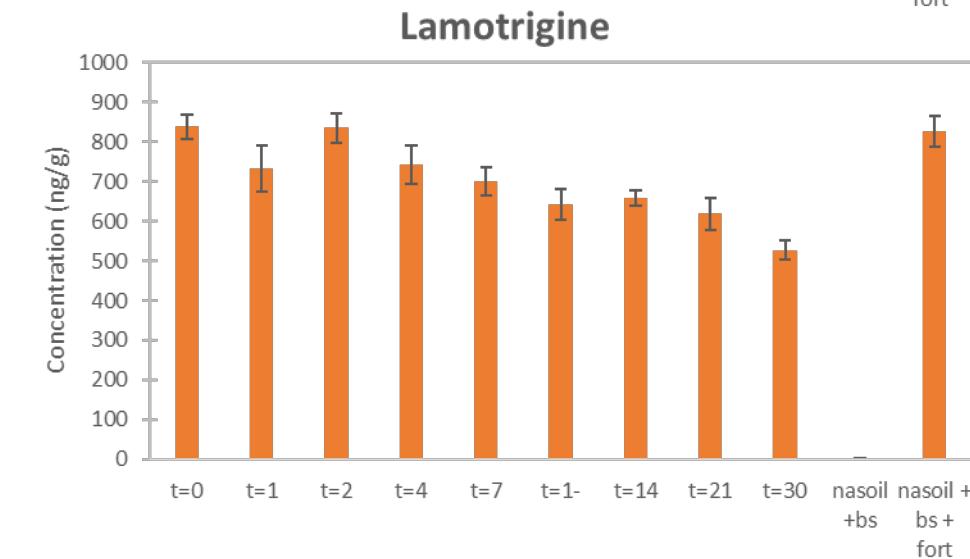
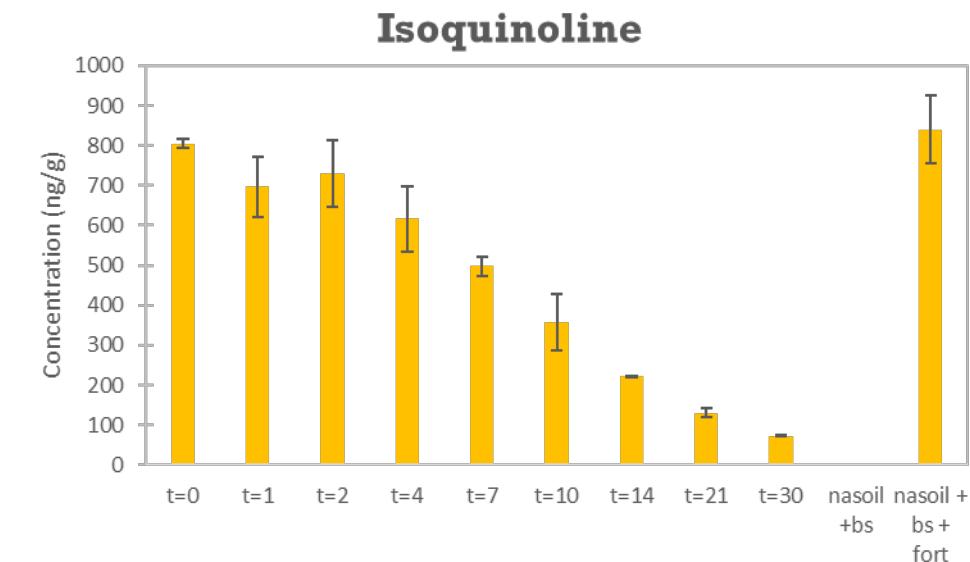
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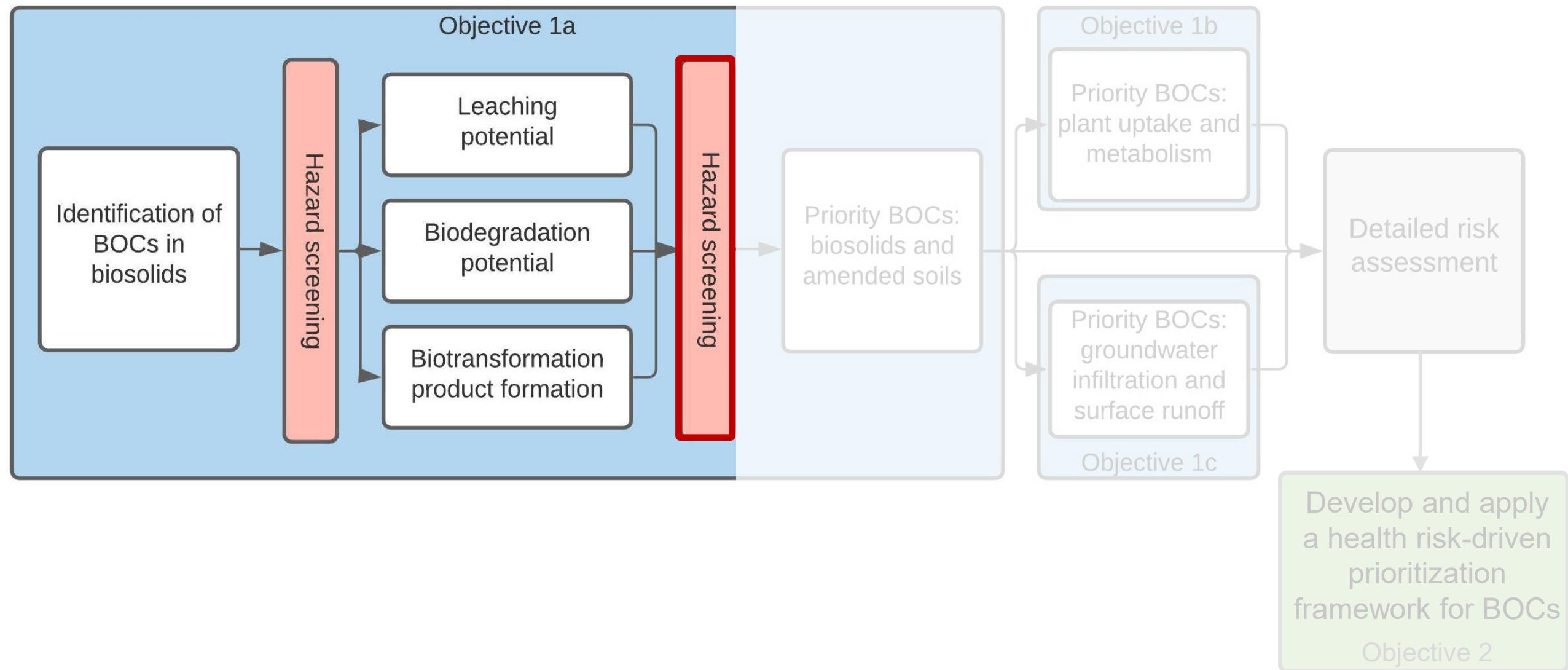
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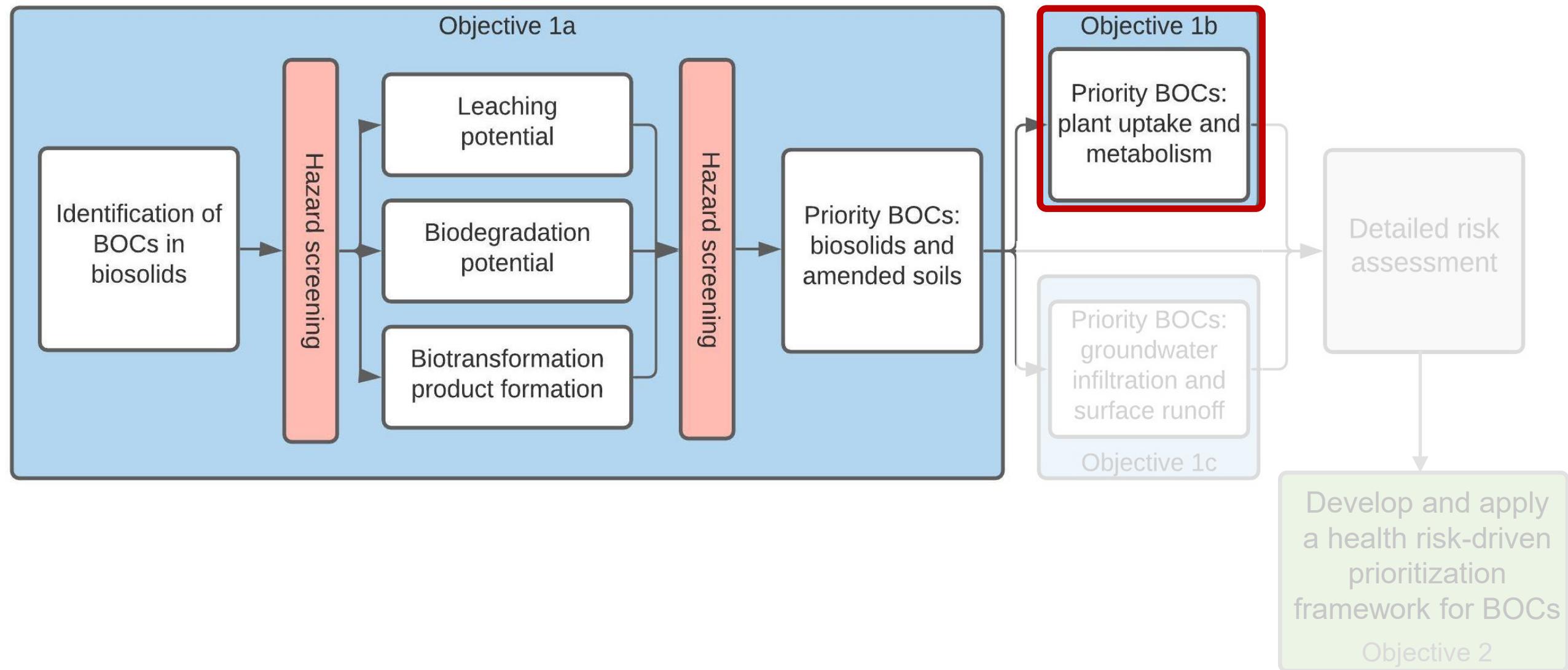
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# Characterize the occurrence, fate, transport and risks of novel biosolid-associated contaminants (BOCs)



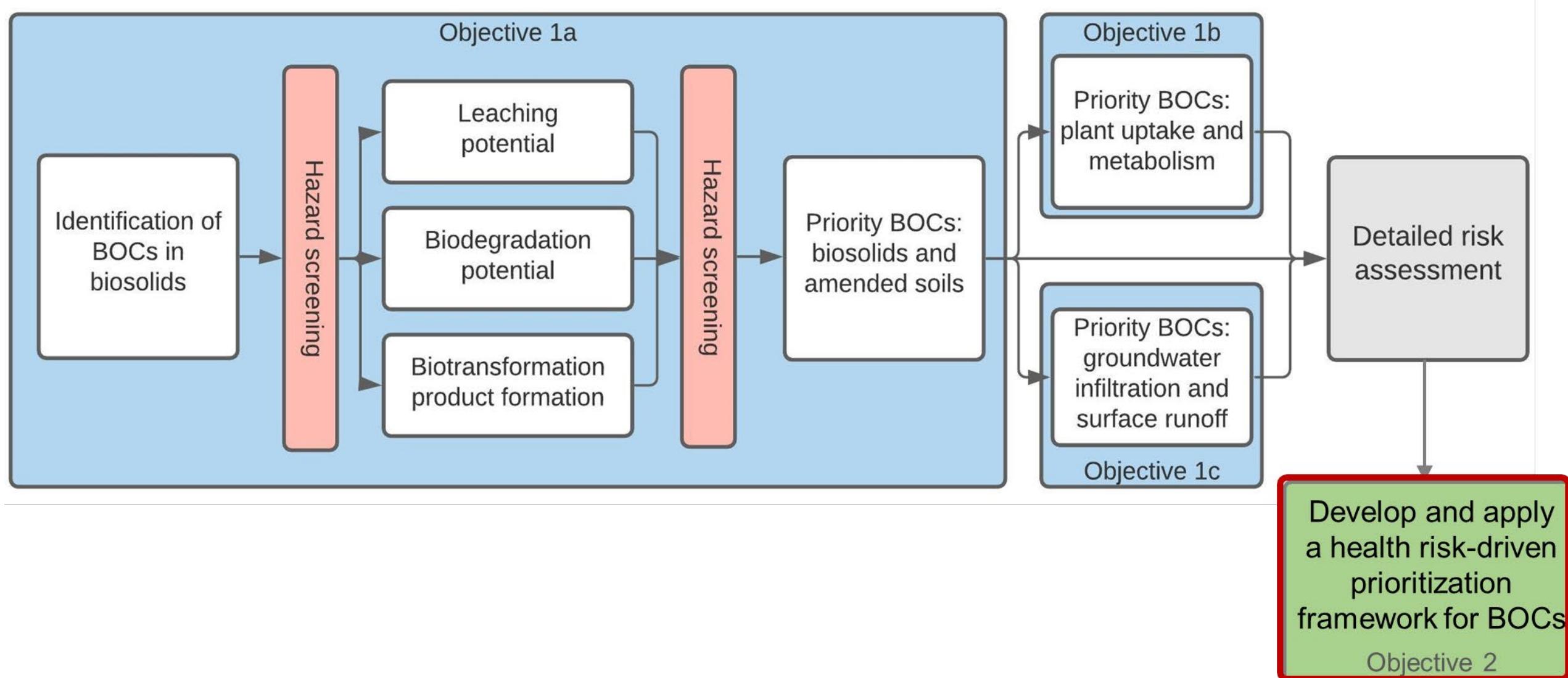
# Characterize the occurrence, fate, transport and risks of novel biosolid-associated contaminants (BOCs)



# Lab & field experiments – Plant uptake



# Characterize the occurrence, fate, transport and risks of novel biosolid-associated contaminants (BOCs)



# Pilot Study: Characterization of exposure scenarios and quantification of exposure factors unique to biosolids workers



- Semi-structured in-depth interviews (IDI's) with workers from seven states across the US.
- Land application involves six processes:
  - hauling, loading, spreading, post-application field work, cleaning, and maintenance
  - differing levels of exposure to biosolids based on the used processes
- Findings were used to develop a biosolids exposure questionnaire (BEQ) to quantify exposure factors unique to biosolids workers.
- We are currently administering the questionnaire to biosolids workers.

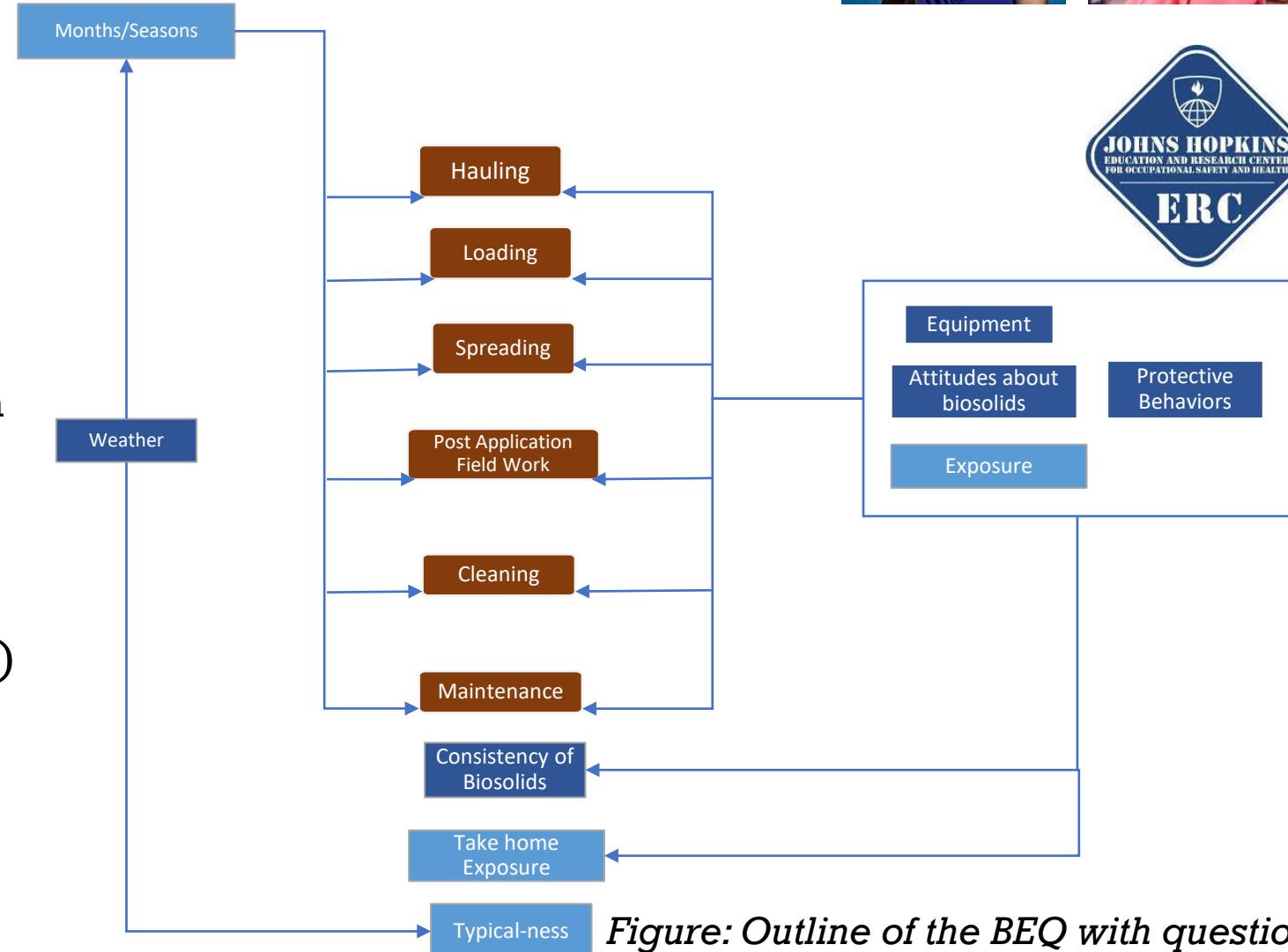


Figure: Outline of the BEQ with questions that leverage findings from the IDIs

# Conclusions

- Framework based on high-resolution mass spectrometry and hazard assessment tools is a promising approach for the prioritization of compounds in complex mixtures

# Conclusions

- Framework based on high-resolution mass spectrometry and hazard assessment tools is a promising approach for the prioritization of compounds in complex mixtures
- Prioritization can support:
  - Selection of analytes for identification using reference standards
  - Selection of compounds for in-depth toxicity assessment
  - Selection of compounds for risk assessment

# Conclusions

- Framework based on high-resolution mass spectrometry and hazard assessment tools is a promising approach for the prioritization of compounds in complex mixtures
- Prioritization can support:
  - Selection of analytes for identification using reference standards
  - Selection of compounds for in-depth toxicity assessment
  - Selection of compounds for risk assessment
- Major challenge is the limited number of compounds in MS databases

# Thank you!

Email: cprasse1@jhu.edu

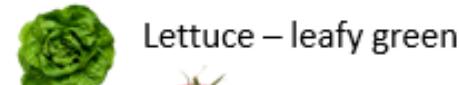
Lab homepage: [www.prasselab.com](http://www.prasselab.com)

# Laboratory experiments – Plant uptake/metabolism

## Pot-spike Experiments

- Target and non-target approach with LC-HRMS
- Determine uptake and accumulation of priority contaminants from biosolids-amended soil
- Identify *in-planta* transformation products of priority contaminants

Representative crop plants:



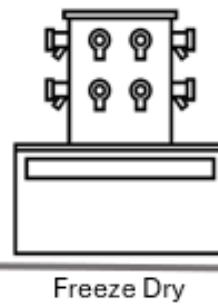
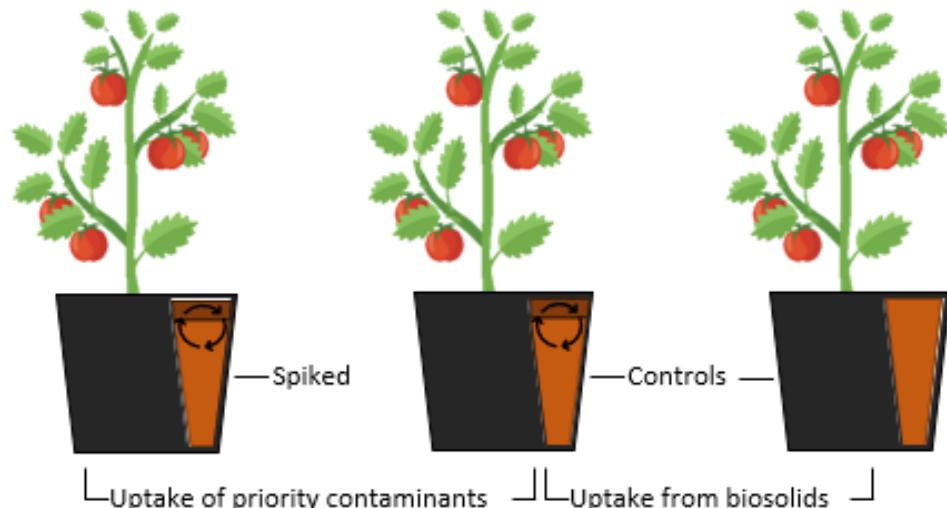
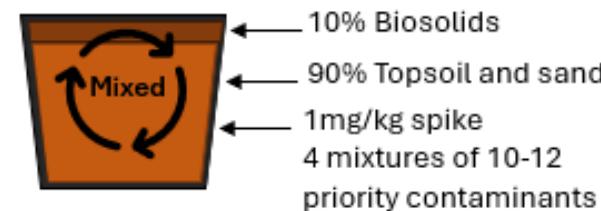
Lettuce – leafy green



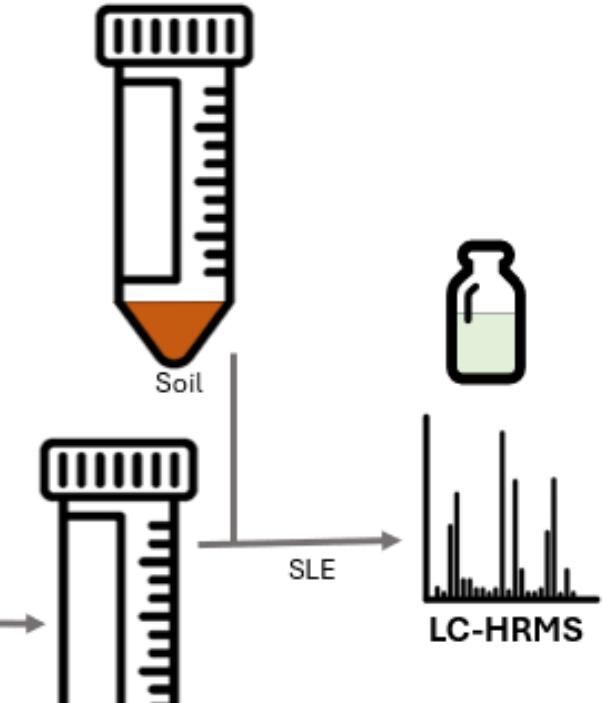
Tomato – fruiting



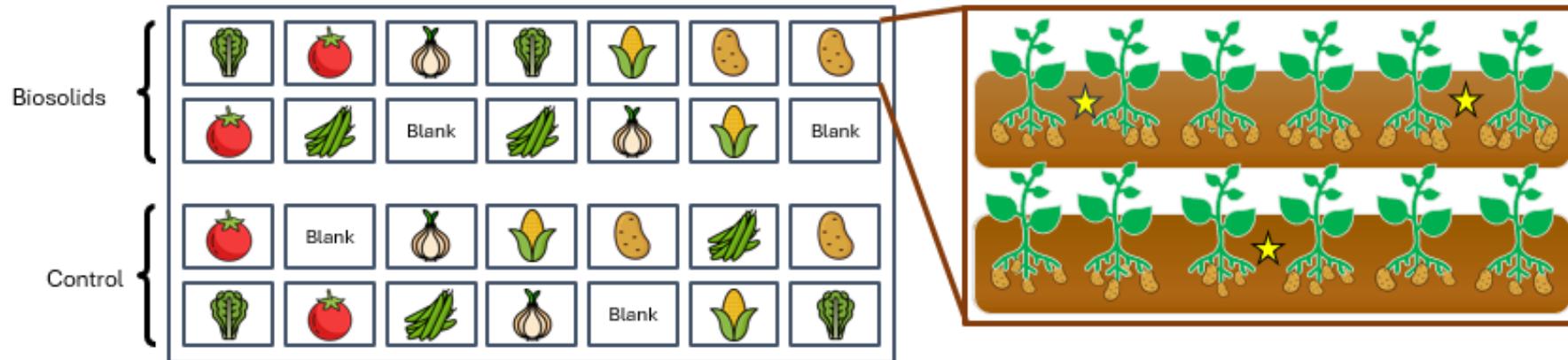
Onion – root



Freeze Dry



# Field experiments – Plant uptake

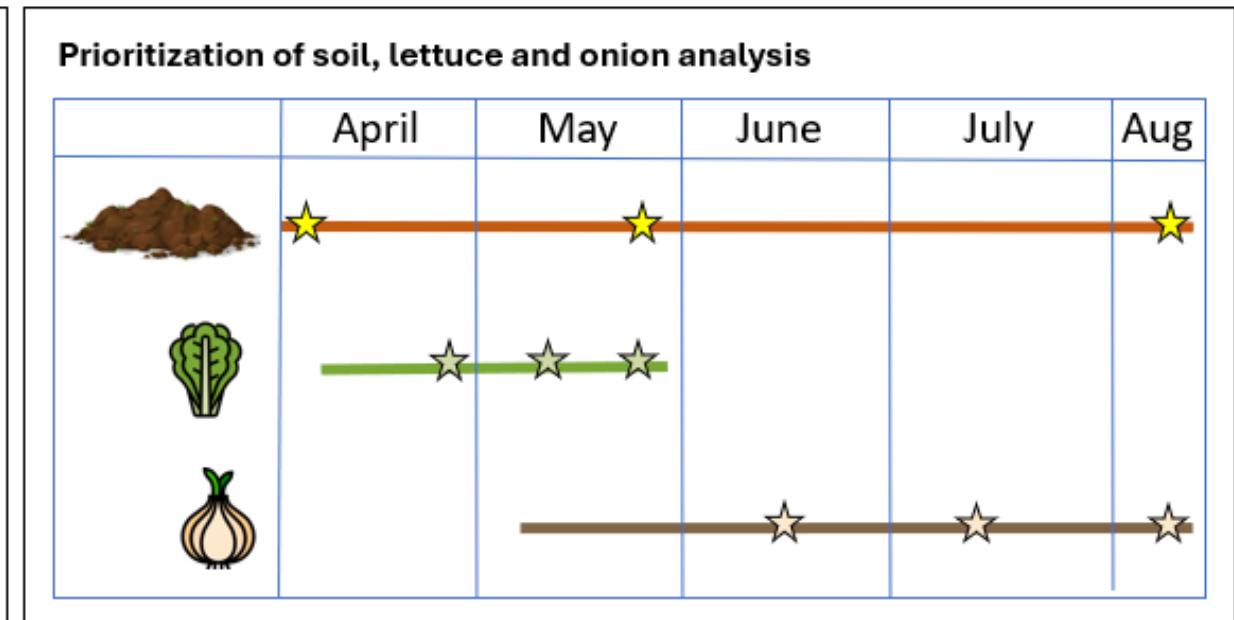
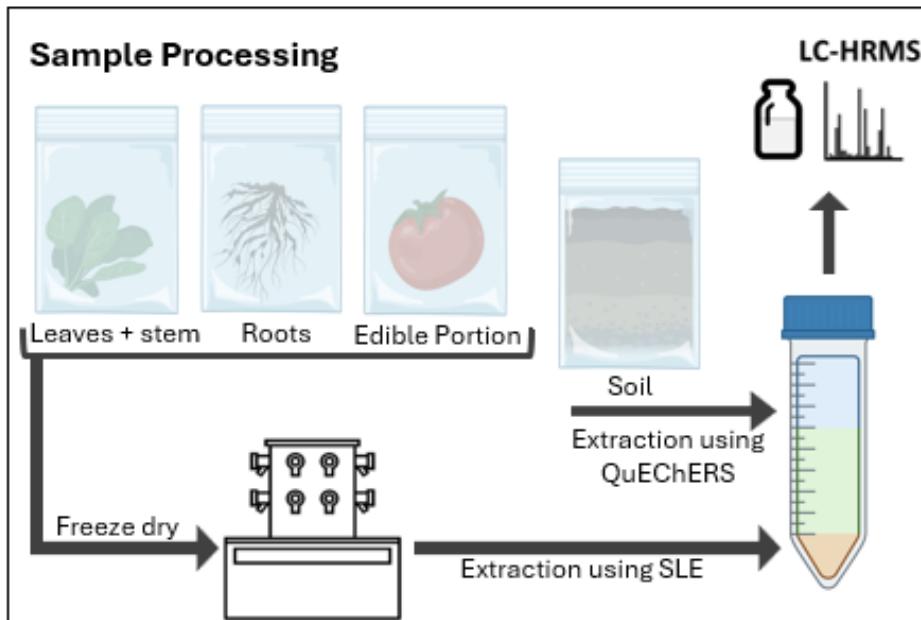


## Soil Sampling

- 3 locations per plot
- 2 time points (biosolid application and harvest)

## Plant Sampling

- 3 sampling events
- 3 plants / sampling event
- Up to 3 different tissue types



# Collaborative Vision for Omics-Based Chemical Testing

Connie Mitchell

Senior Scientific Program Manager

Health and Environmental Sciences Institute (HESI)

# The HESI Model: Bridging Research to Application

One of first and longest standing nonprofits to use multi-sector, int'l model - since 1989!

## IMPROVED SAFETY AND INNOVATION FOR HUMAN AND ENVIRONMENTAL HEALTH



Academic, Clinical,  
& Research Scientists  
& Organizations



NGOs, Patient  
Advocacy Groups,  
& Foundations



Industry  
Research &  
Development



Government  
Research &  
Regulation

# Emerging Systems Toxicology for the Assessment of Risk (eSTAR)

## Committee Mission

- Develop and deliver innovative systems toxicology approaches for risk assessment
- Catalyze adoption of new translational and predictive tools that guide decision-making based on mechanistic understanding of toxicological response



# eSTAR Participants

## Academia

- Broad Institute
- Cambridge University
- Clemson University
- Cornell University
- Georgetown University
- Kansas University Medical Center
- Indiana University
- McGill University
- Michigan State University
- MIT
- North Carolina State University
- Newcastle University
- Orebro University
- University of Ottawa
- University of Michigan
- University of North Carolina
- University of Pittsburgh

## Government

- BC Cancer Agency
- BfArM
- Dutch Medicines Evaluation Board
- Health Canada
- PMRA
- US Army
- US EPA
- US FDA
- US NIH NCI
- US NIH NIEHS

## Non-Profit

- PSCI
- Lhasa

## Industry

- AbbVie
- Amgen
- AstraZeneca
- Bayer AG
- Boehringer-Ingelheim
- Corteva
- FMC
- GSK
- Janssen
- Merck & Co., Inc
- Merck KGaA
- Newcells Biotech
- Ono
- Pfizer
- Recursion
- Roche/Genentech
- Sanofi-Aventis
- Servier
- Syngenta
- Taconic Biosciences
- Takeda
- TwinStrand Biosciences
- Vertex



eSTAR

A large, solid orange circle is positioned on the left side of the slide, covering approximately the first third of the vertical space.

Omics:  
what do I  
mean

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Transcriptomics

---

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Genomics

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Proteomics

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Metabolomics

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Epigenomics

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Phenomics (e.g., Cell Painting)

# Omics: what are they good for?

- Promise of omics at the turn of the century
  - Improved understanding of toxicology mechanisms, species, and susceptibility
- Difficulty to use omics in regulatory decision making
- “Hairball diagrams” – pathway diagrams, heat maps, long list of differentially expressed genes.
  - What do we do with these data?
- Move towards using omics for decision making
  - Biomarkers for pathways
  - Deriving points of departures

# Current eSTAR Working Groups



## Molecular POD

Using transcriptomic point of departure for chemical risk assessment.



## TGx-DDI

An *in vitro* transcriptomic biomarker to predict probability that an agent is DNA damage-inducing (DDI) or non-DDI.



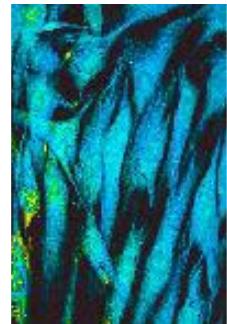
## Carcinogenomics

Identifying and evaluating transcriptomic biomarkers for rat-liver tumors to reduce the need for two-year carcinogenicity studies.



## Error Correct Sequencing

Exploring the use of error corrected sequencing to detect non-genotoxic carcinogens.



## OASIS

Exploring the use of Cell Painting, transcriptomics, and proteomics for safety assessment.



## miRNA Biomarkers

Investigating and evaluating the use of miRNAs as biomarkers for renal injury.

# **HESI Emerging Systems Toxicology for Assessment of Risk (eSTAR) Committee 2024 Annual Meeting**

Wednesday 13 November 9 am – 3 pm ET

Free, open meeting to learn about all the projects.



[Register Here](#)

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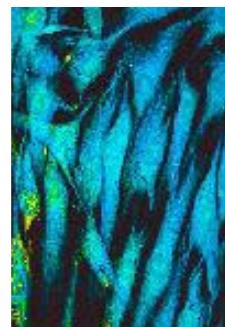
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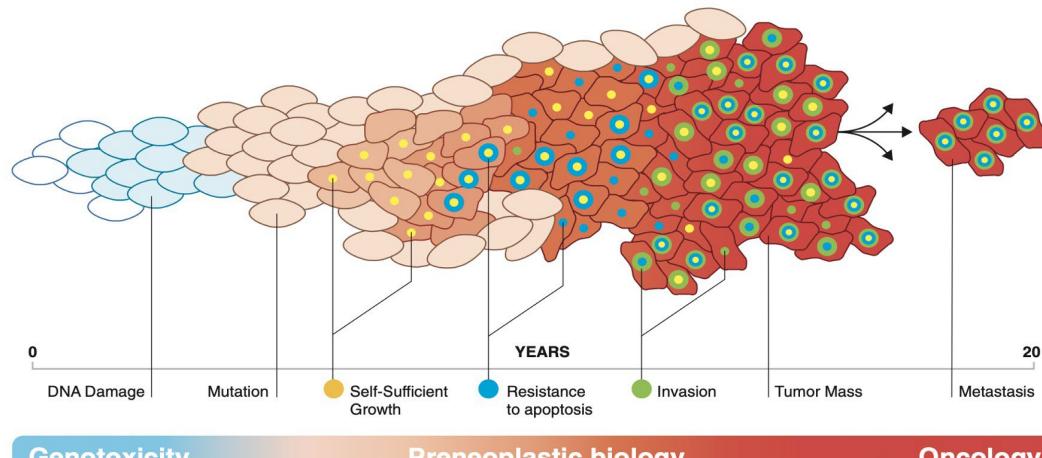


## miRNA Biomarkers

Investigating and evaluating the use of miRNAs as biomarkers for renal injury.

# Why a TGx-DDI Biomarker?

## Genotoxicity is a key driver of carcinogenesis



Salk and Kennedy, 2020

Need for high-throughput genotoxicity tests that provide mechanistic information in human-relevant cell culture models.

## Standard Tests for Chromosome Damage and Mutation:



Bacterial Reverse Mutation Test (Ames assay)

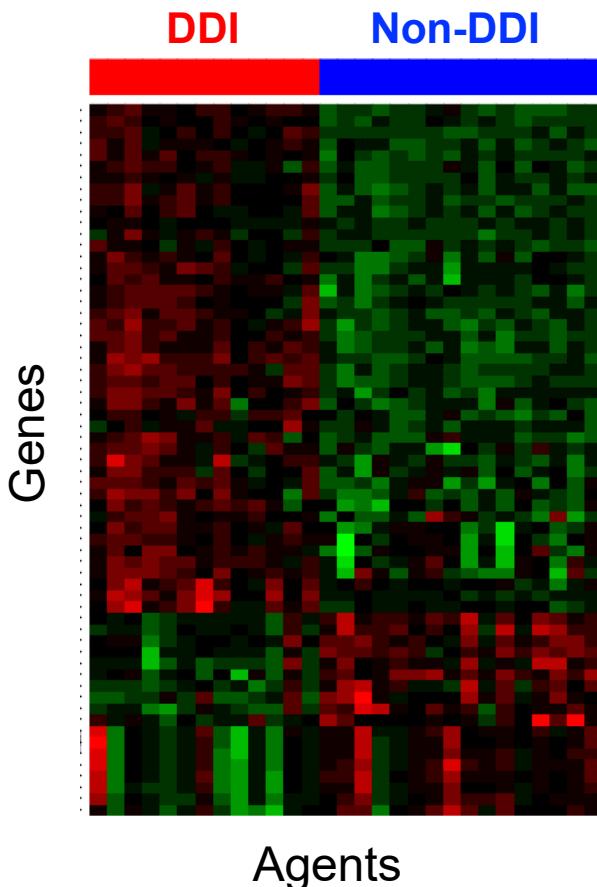
*In vitro* Mammalian Genetox Assay: chromosome aberrations, micronuclei, gene mutation

*In vivo* Genetox Assay: chromosome aberrations, micronuclei, transgene mutation

## Limitations of current tests:

- Test methods are sensitive but lack specificity leading to high rates of misleading positive results.
- Lack efficiency and do not provide information on the mechanisms causing genotoxicity.

# Fit for Purpose Solution: TGx-DDI



An *in vitro* transcriptomic biomarker that predicts the probability that an agent is DDI (DNA damage-inducing) or non-DDI using toxicogenomics (TGx).

- Developed in human TK6 cell cultures
- From exposure to 28 prototype DDI and non-DDI chemicals
- 64 genes identified as being predictive of DDI potential

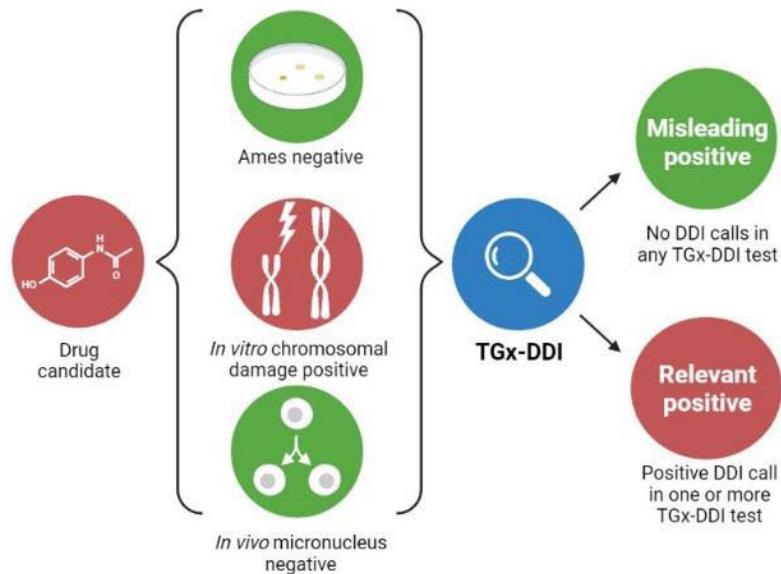
## Why do we like it?

- High specificity (correctly identifies 'irrelevant' chromosome damage results)
- Endogenous cellular responses to DNA damage in human cells
  - Biological relevance
- Transferable to other cell types
- Multiplex capacities
- Removes subjectivity of transcriptomic data analysis

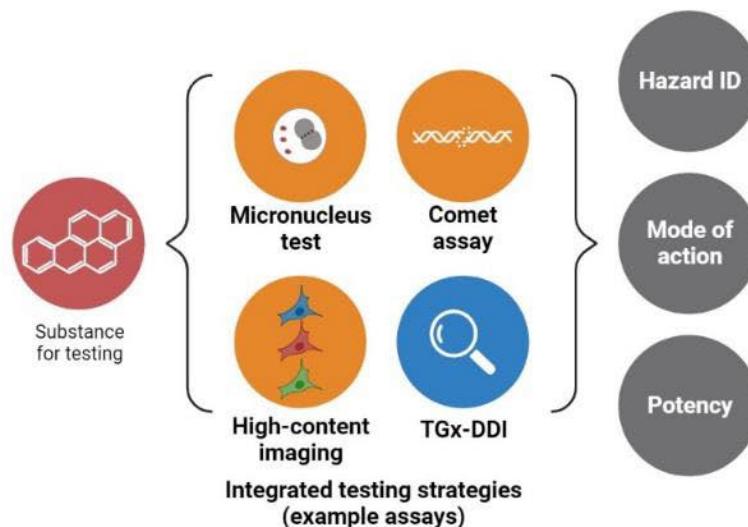
Li, HH et al. *Environ Mol Mutagen* (2015);  
*PNAS* (2017)  
Yauk, et al. *Environ Mol Mutagen* (2016)

# The context of use

## 1. Drug screening



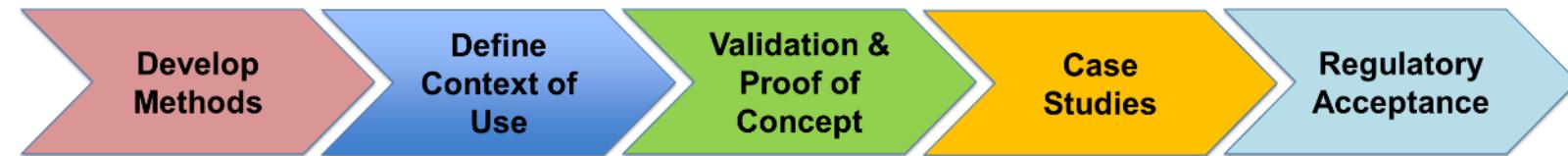
## 2. Chemical safety assessment



### Regulatory tests that TGx-DDI addresses

1. TGx-DDI can be used in weight of evidence analyses to inform irrelevant *in vitro* chromosome damage results (FDA).
2. TGx-DDI can be used in integrated testing to inform potential hazards and mode of action, and for potency assessment.

# A long ride to date...



- TGx-DDI analytical pipelines
- TGx-DDI biomarker software tool
- BMD modeling
- HepaRG
- Approximately 100 chemicals tested
- Metabolic activation
- Cross-platform – Affymetrix, qPCR, NanoString, RNA-seq, TempO-seq
- Three case studies completed
- **FDA Biomarker Qualification Plan**
- Part of Health Canada's GeneTox21 platform
- **Integrated Approaches to Testing and Assessment (IATA)**



2010

2024

18 Papers published  
NIEHS tool  
qPCR array  
CRO offering assay!

 ScitoVation  
INNOVATORS IN CHEMICAL SAFETY ASSESSMENT

[Services](#) [Publications](#) [Tools](#) [Resources](#)

## Genotoxicity

### Non-Animal Human-Relevant Genotoxicity Assessments for Lead Candidate Selection

Knowledge regarding the potential of pharmaceuticals, food additives or chemicals to damage the human genome and cause mutations remains critical to public health. Mutations are key events in the induction of cancer, birth defects, and neurological disease. The regulatory genetic toxicology test battery required by the US Food and Drug Administration (FDA) for Investigational New Drug applications to initiate clinical trials, and by the US Environmental Agency (EPA) for registration of pesticides, includes approved *in vitro* tests for assessing genotoxicity and mutagenicity. A positive response in this regulatory test battery can eliminate drug candidates from further development or require further *in vivo* testing to demonstrate safety. These *in vivo* tests add significant time to development and can be costly.

ScitoVation's genotoxicity program is aimed at eliminating potential genotoxicity hazards early. This saves time and resources by reducing positive outcomes in the regulatory genetic toxicology test battery that can stop further development. Current genetic toxicology testing is plagued with long backlogs for study starts, outdated technologies focused on rodent cell lines, and genotoxicity assessment using high dose animal testing (up to 2000 mg/kg) with significant risk assessment assumptions used for extrapolation of those data to humans.

**What we offer:**

- Computational tools based on compound structure for screening early candidates that are highly predictive of bacterial mutagenicity testing.
- For lead candidates, the approved *in vitro* micronucleus assay using human TK6 or HepaRG cells.
- Our depth of expertise in toxicogenomics, enables us to offer the TgX DDI biomarker, which is currently undergoing FDA validation as biomarker predictive of mammalian genotoxicity. The advantage of using TgX DDI is that it provides a point of departure that can be used to translate the *in vitro* results to human equivalent dose.

Each stage can be offered separately, depending on the needs of the client.

# Recently Completed: Validation Study

**Objectives:** To assess the cross-laboratory reproducibility of TGx-DDI classification calls involving one platform (NanoString)

**Resourcing:** *Funding from FDA (1U01FD007473-01) and in-kind effort and materials from HESI partner organizations.*

## Experimental Plan:

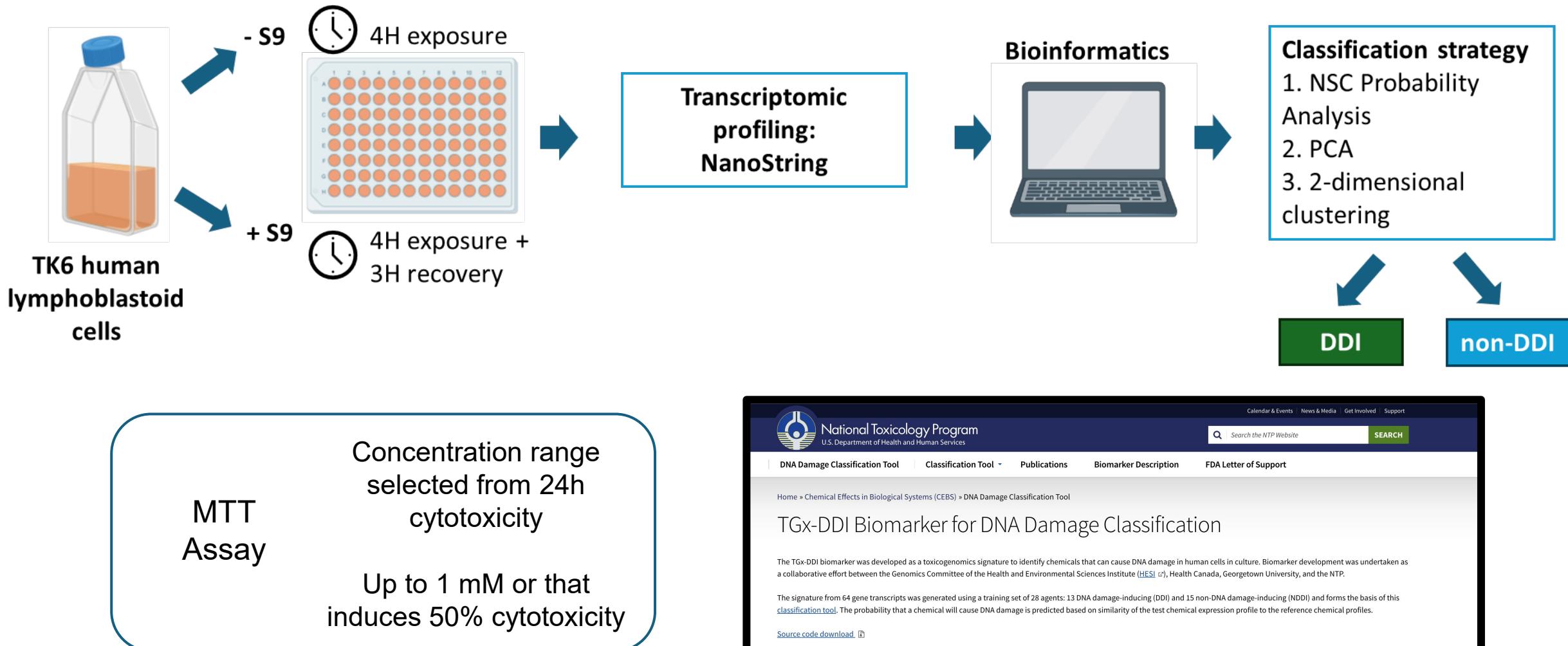
- 4 study sites
- Sites receive test compound in solution (three concentrations and solvent)
- 14 chemicals tested
- TK6 cells, samples in triplicate
- Culture and exposure per the SOP
- Analysis and classification using public software tool



Study design reviewed by FDA and Agency feedback incorporated on multiple occasions.

INSTITUTION	MULTI-SITE STUDY CONTRIBUTIONS				
	Study coordination (meetings, logistics, supply procurement, shipping)	TGx-DDI Assay (cell culture, exposure, RNA isolation)	NanoString (RNA QC & Transcriptomics)	Data Analysis, Interpretation & Reporting	Data Compilation, Presentation and Cross Site Data Analysis
 HESI	x				x
 Georgetown University	x	x	x	x	
 SANOFI		x		x	
 Procter & Gamble Laboratories		x			x
 Burleson Research Technologies		x	x	x	
 Children's National Genomics Core			x		
 THE WISTAR INSTITUTE			x		

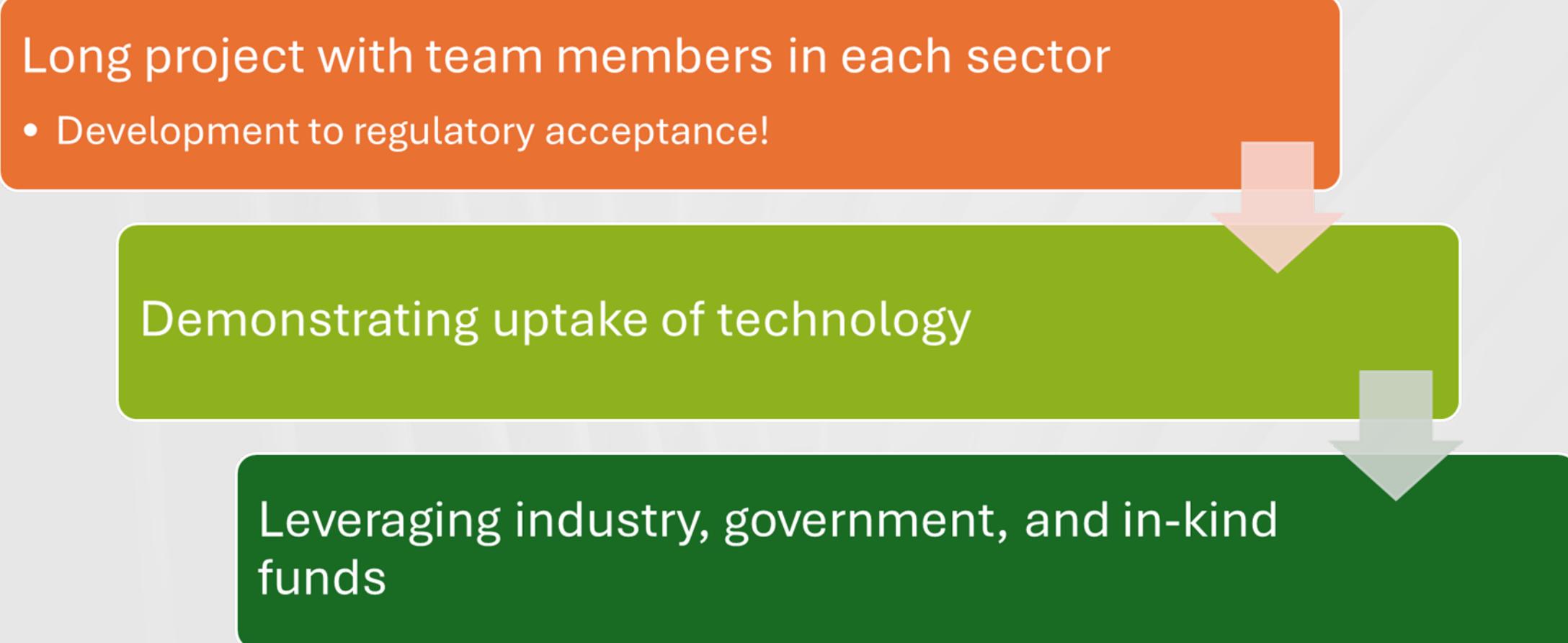
# TGx-DDI Ring Trial Experimental Workflow



# Collaboration is key

Long project with team members in each sector

- Development to regulatory acceptance!



Demonstrating uptake of technology

Leveraging industry, government, and in-kind funds

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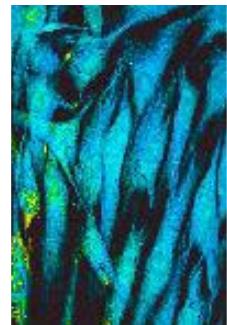
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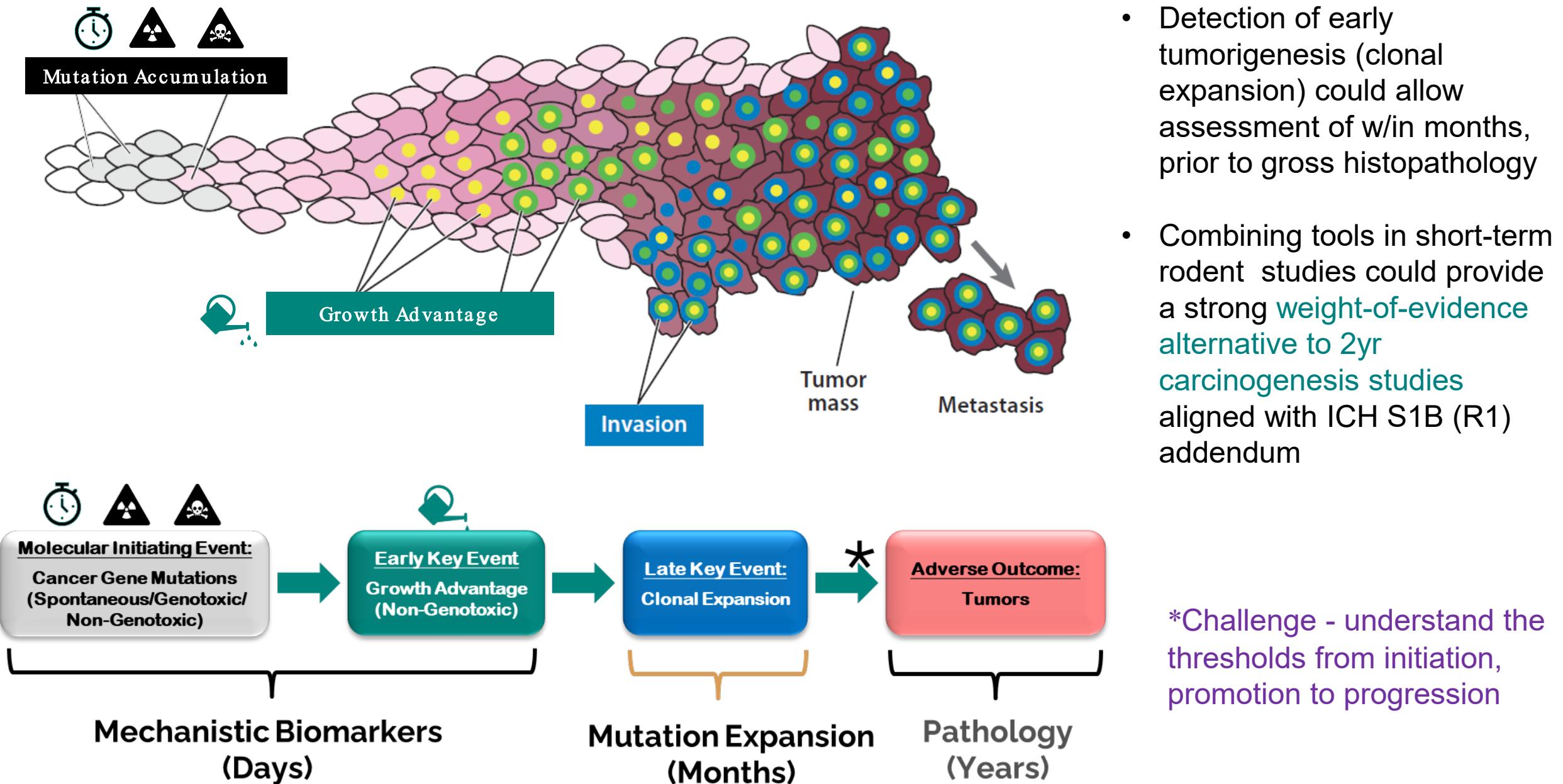
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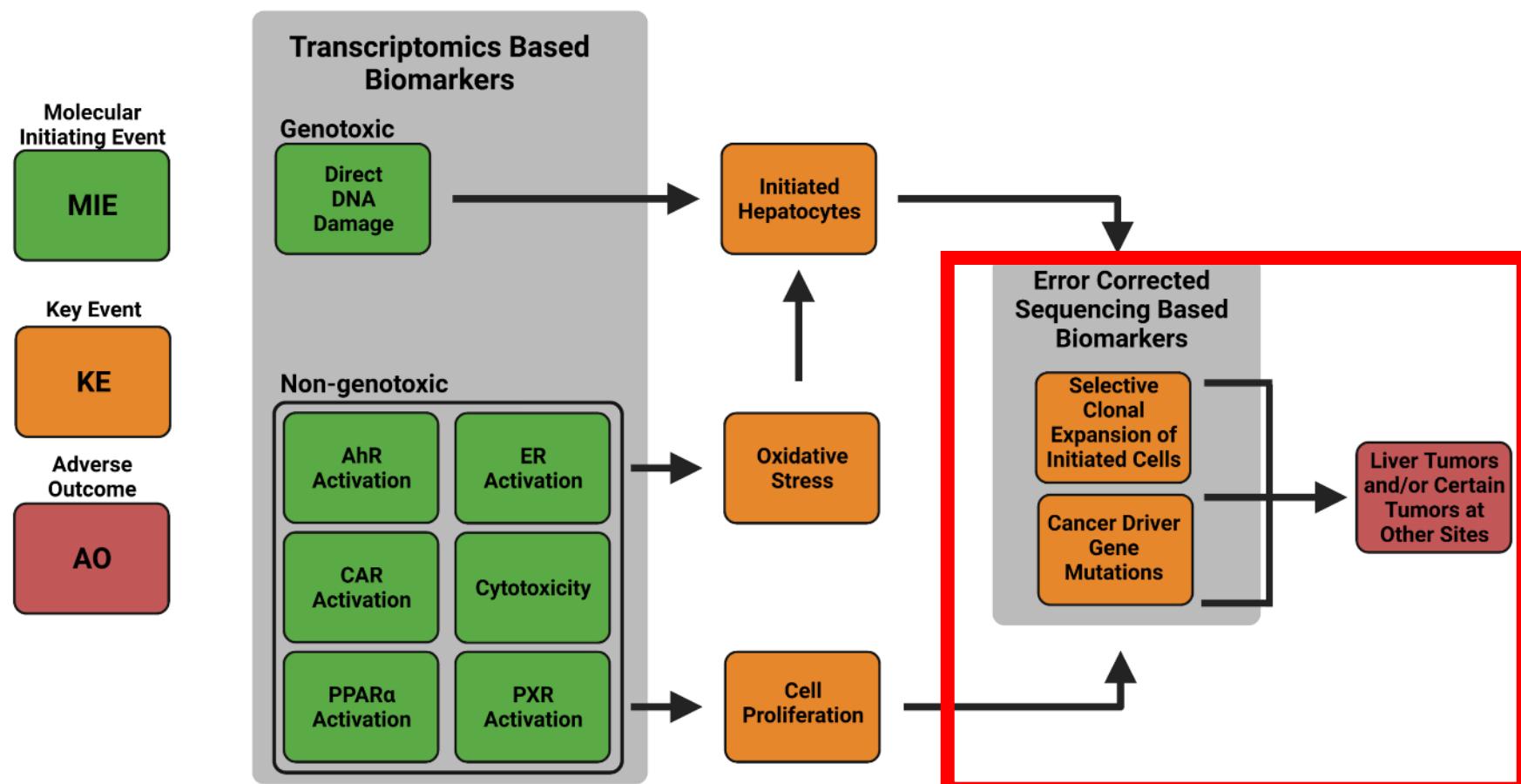


## miRNA Biomarkers

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# Progression of Cancer Biology: Opportunities for Earlier Detection

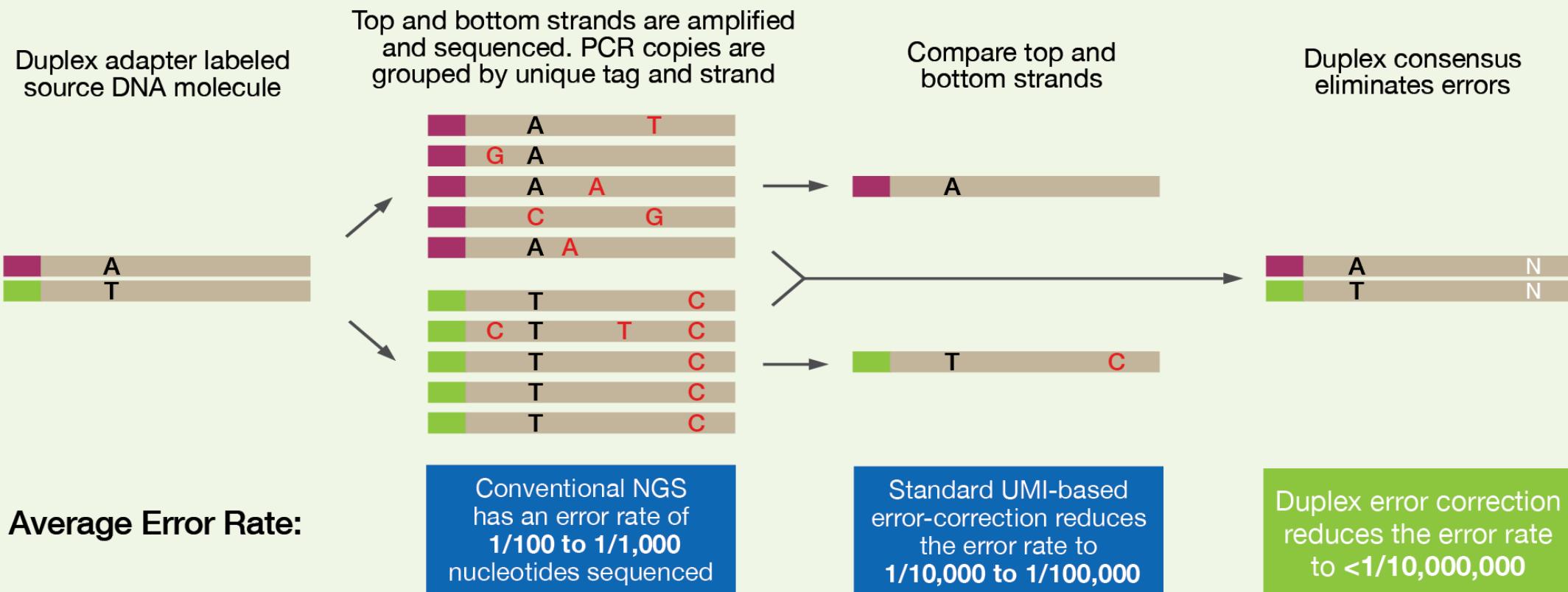




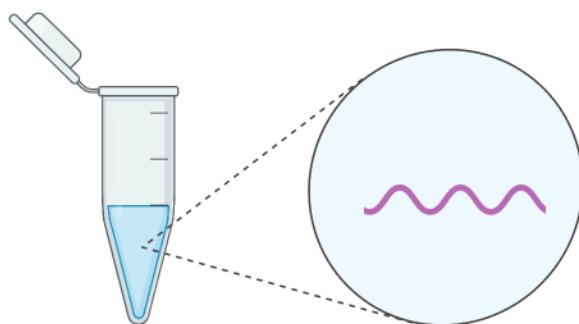
Use Transcriptomics Based Biomarkers to query 7 commonly observed molecular initiating events in sub-chronic/chronic rat studies with histologic risk factors of neoplasia to provide explanations for chemical carcinogenic mechanisms and inform human relevance.

Error Corrected Next Generation Sequencing technology to identify DNA Cancer Driver Gene Mutation based biomarkers for selective clonal expansion that could address earlier hypothetical concerns of carcinogenic risk.

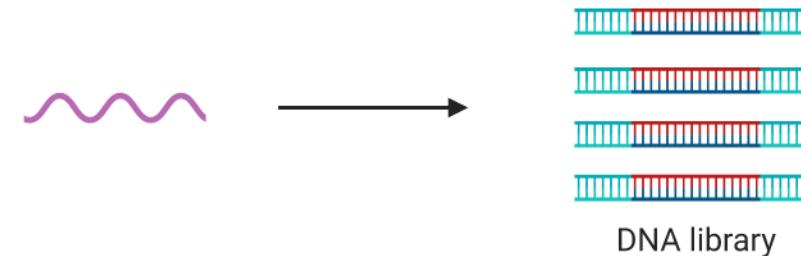
# Error Corrected Sequencing (eg. DuplexSeq™) to Measure Clonal Expansion of Cancer Driver Genes



**Step 1:**  
DNA extraction

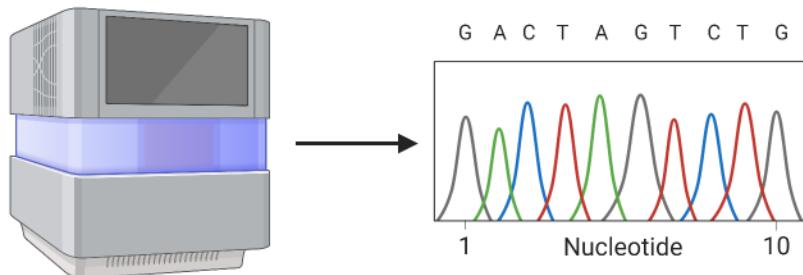


**Step 2:**  
Library preparation

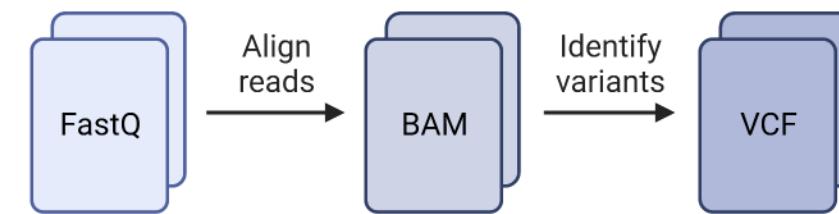


**Sequencing Workflow**

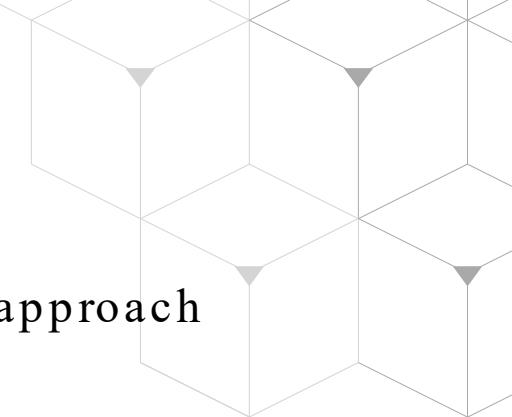
**Step 3:**  
Sequencing



**Step 4:**  
Analysis



# How do we build confidence?



## 1. Additional Compounds/ Studies:

- Studies with additional compounds will help establish sensitivity/ specificity of the approach as well as the ability to distinguish tumorigenic from non-tumorigenic dose levels

## 2. Additional Genes:

- A sensitive assay will require broad coverage of potential driver genes across different NGT mechanisms
- Studies with additional compounds will help identify gaps in existing panels and WES on these samples can be used to identify novel driver genes.

## 3. Establish Thresholds:

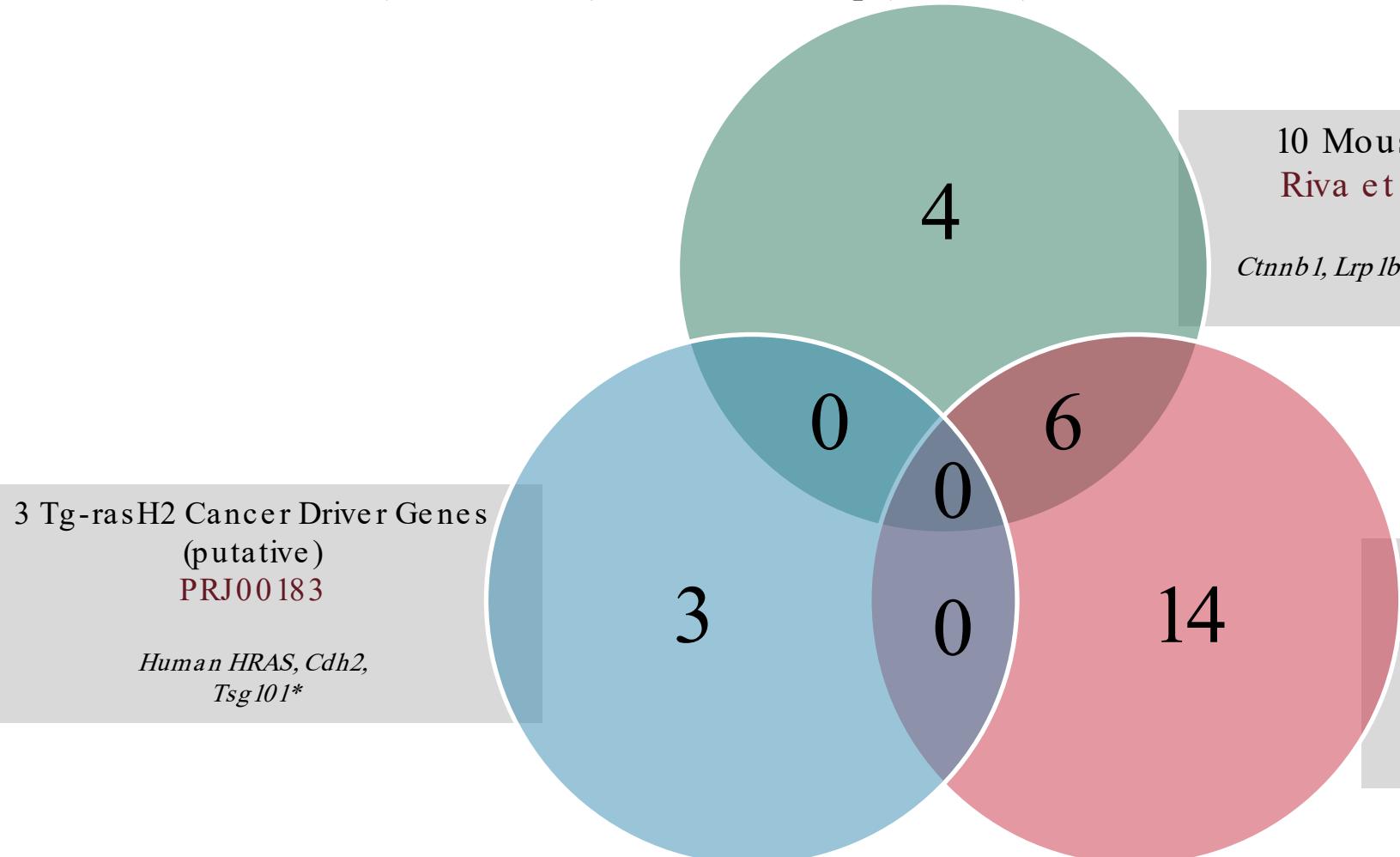
- Establish normal baseline of expanded clones across time points, and normalized thresholds of concern to enable early risk assessment of novel compounds

## 4. Regulatory Acceptance:

- Ultimate goal is to use as part of a weight-of-evidence approach for carcinogenic risk assessment and potential waivers of long-term carcinogenesis studies

# Mouse carcinogenesis panel v1

Panel design covers 27 cancer driver genes that were identified in Tg-rasH2 tumors, Riva et al. (Wt-mouse), and CarcSeq (human)



\*Not known cancer driver gene

10 Mouse Cancer Driver Genes  
Riva et al., Nat Genetics (2020)

*Ctnnb1, Lrp1b, Cnot3, Egfr, Fgfr2, Kras, Hras, Braf, Trp53, EphA3*

3 Tg-rasH2 Cancer Driver Genes  
(putative)  
PRJ00183

*Human HRAS, Cdh2,  
Tsg101\**

20 Human Cancer Driver Genes  
Harris et al., Tox Sci (2021), CarcSeq

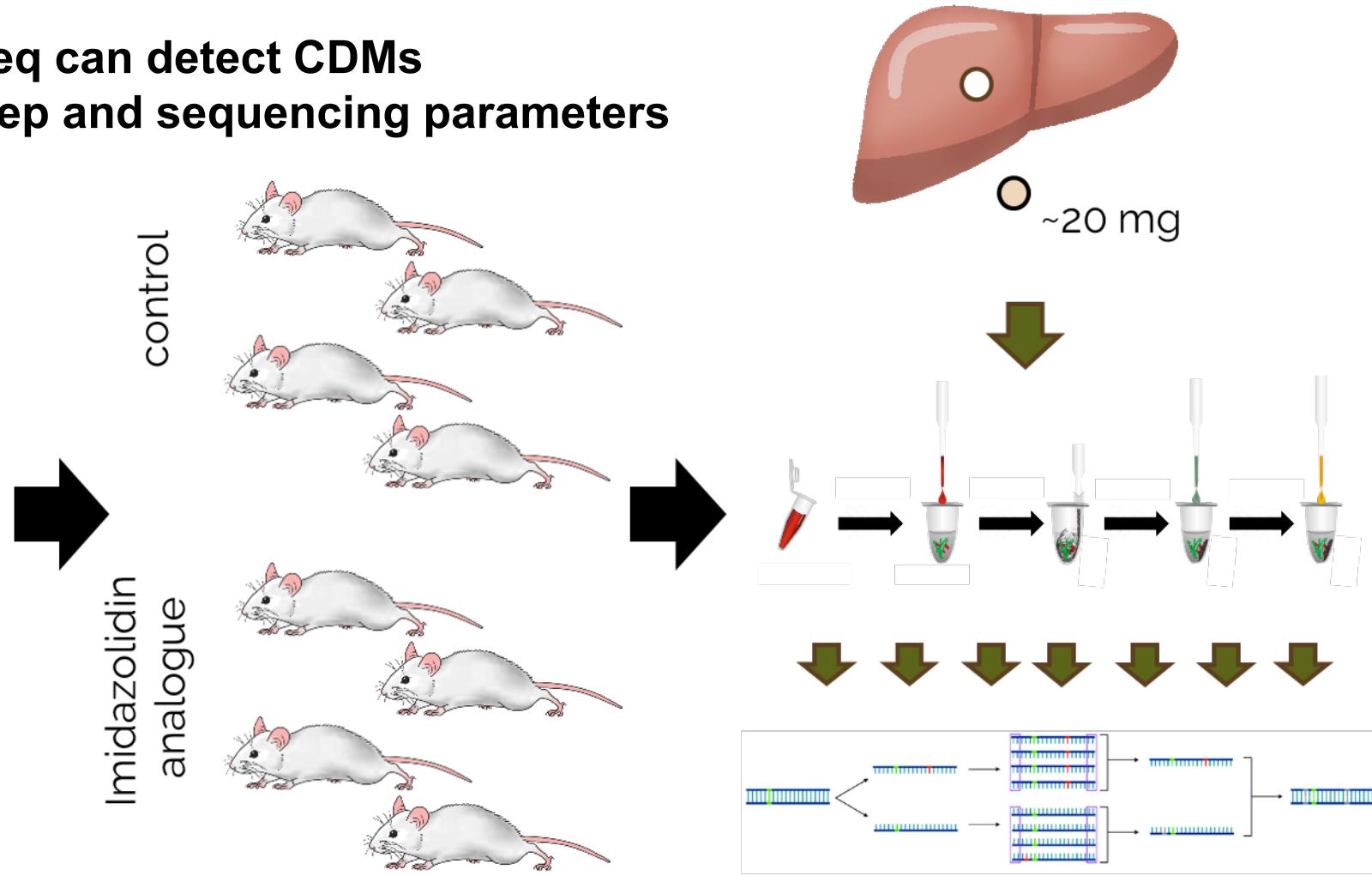
*Pik3ca, Trp53, Stk11, Kras, Hras, Braf, Egfr, Lrp1b, Nfe2l2, Apc, Setbp1, Tert, Rb1, Axin1, Cdkn21, Pten, Acvr2a, Foxl2, Kmt2c, Nras*

# Pilot Study Design

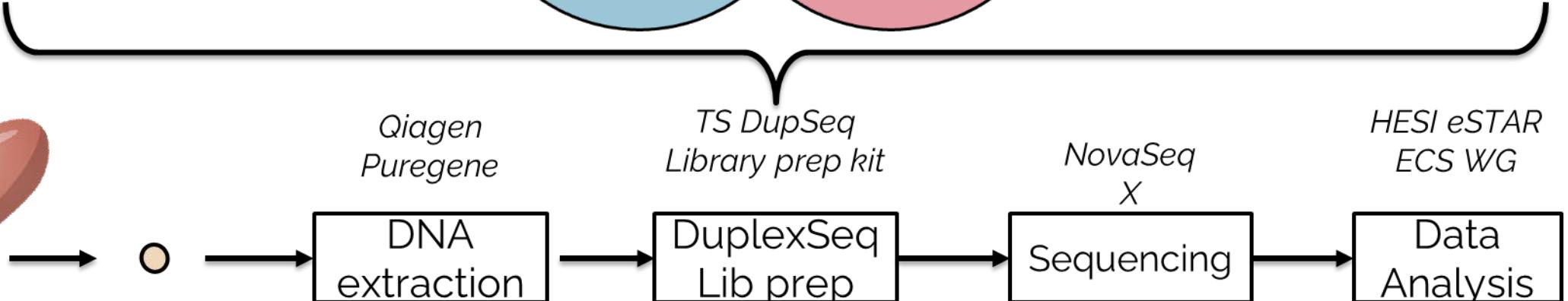
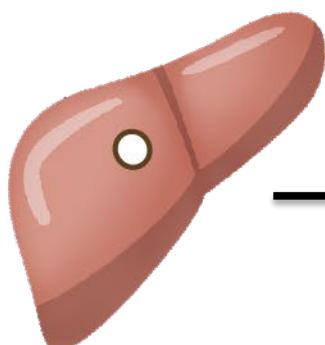
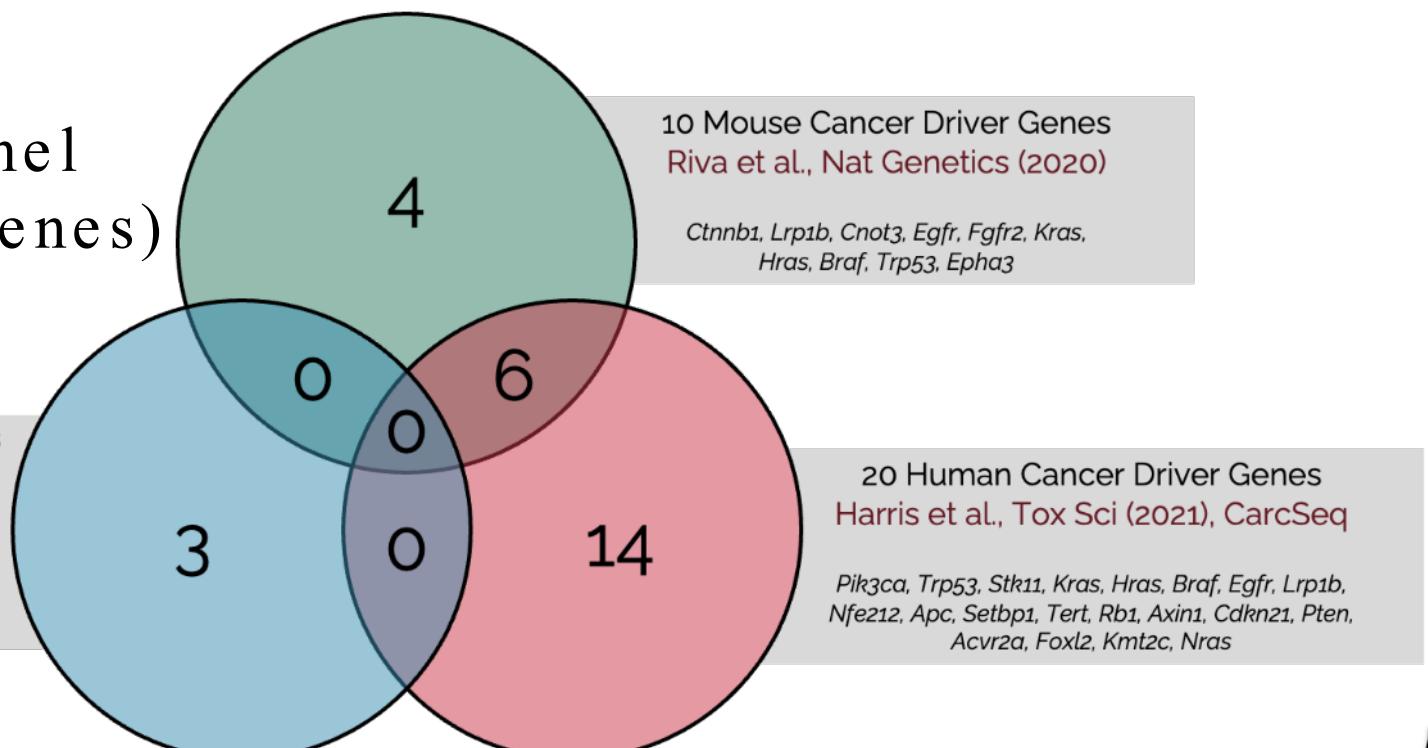
**Goals:** (1) Evaluate if DuplexSeq can detect CDMs  
(2) Establish sample prep and sequencing parameters

- Male CD1 mice
- (OECD 408) 90-day study:  
Liver weight change and  
centrilobular hepatocellular  
hypertrophy
- Vehicle control and 114  
mg/kg (dose higher than the  
carcinogenic dose at 2-year  
bioassay)

\*Syngenta study



# Mouse CDM panel (27 cancer driver genes)

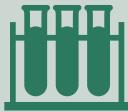


## Overall Study Workflow

# Collaboration is key



Share vision to explore the use of a tool



Multi-site experiment using existing samples



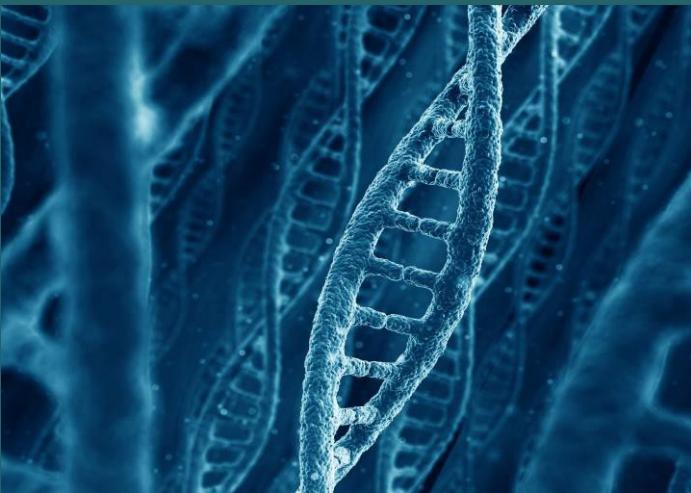
Data Analysis done by multiple groups to agree upon best practices



Email me:  
Connie Mitchell  
(cmitchell@hesiglobal.org)

# Development and Application of Transcriptomics at EPA

Logan J. Everett, Ph.D. – Bioinformatics Scientist, US EPA / ORD

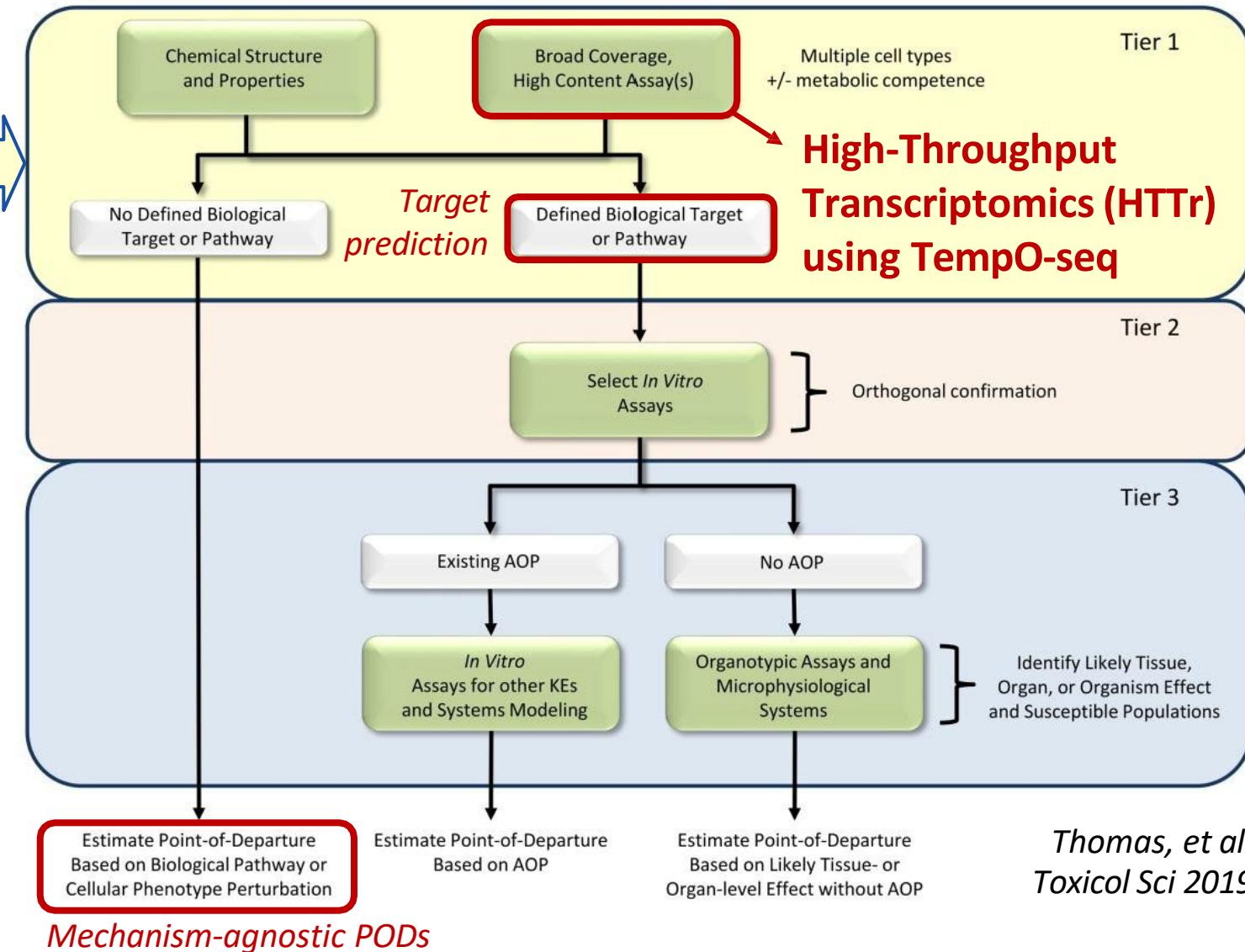


*The views expressed in this presentation are those of the presenter and do not necessarily reflect the views or policies of the U.S. EPA.  
Company or product names do not constitute endorsement by U.S. EPA.*

# Tiered Chemical Safety Testing Strategy

## Tier 1 Primary Goals:

- Prioritize chemicals by bioactivity & potency
- Predict biological targets for chemicals



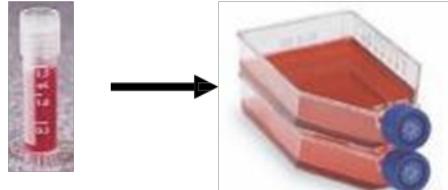
EPA Transcriptomic Assessment Product (ETAP)  
Short-term *in vivo* exposures with transcriptomic profiling



Thomas, et al.  
Toxicol Sci 2019

# Automated *in vitro* Chemical Screening Strategy

Cryopreserved  
Cell Stocks → Cell Expansion



## Cell Line Examples:

MCF-7 Breast epithelium

U-2 OS Bone

HepaRG Liver

## Cell Plating



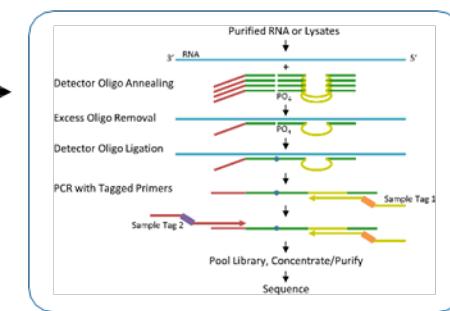
*BioTek*  
*MultiFlo™ FX*

Dispensing Test  
Chemicals



*LabCyte Echo® 550*  
*Liquid Handler*

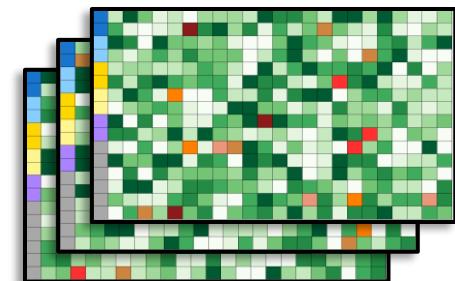
TempO-seq: Targeted sequencing  
for changes in gene expression



Yeakley, et al. *PLoS ONE* (2017)

DOI: [10.1371/journal.pone.0178302](https://doi.org/10.1371/journal.pone.0178302)

## 384-well test plates run in triplicate with:

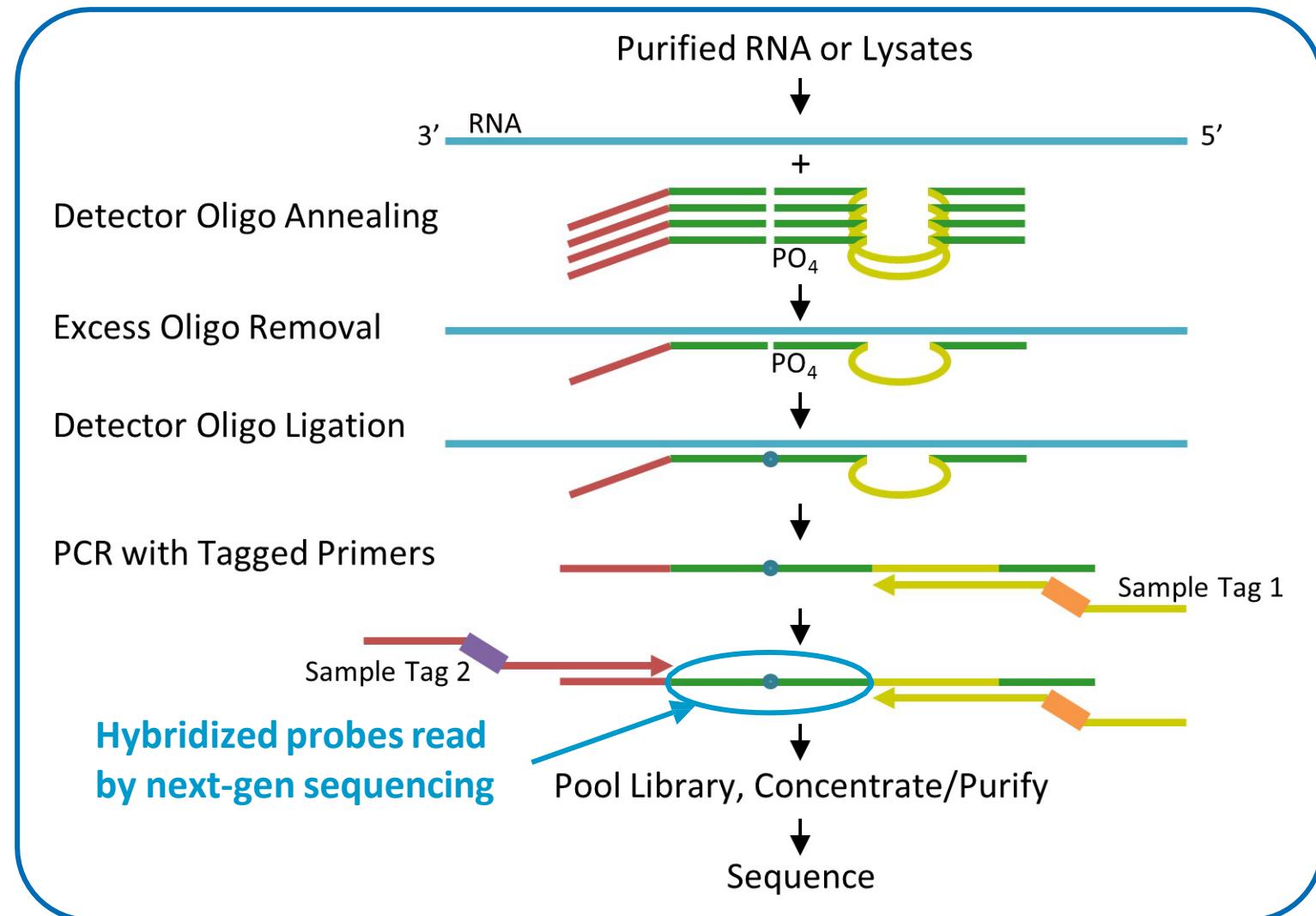


- ~40 test chemicals x 8 concentrations (half-log spacing)
- Multiple vehicle controls, reference chemicals & QC samples on every plate to track assay performance
- Treatment positions randomized on each plate
- Independent culture batch on each plate

See Harrill, et al. *Tox Sci* 2021  
DOI: [10.1093/toxsci/kfab009](https://doi.org/10.1093/toxsci/kfab009)

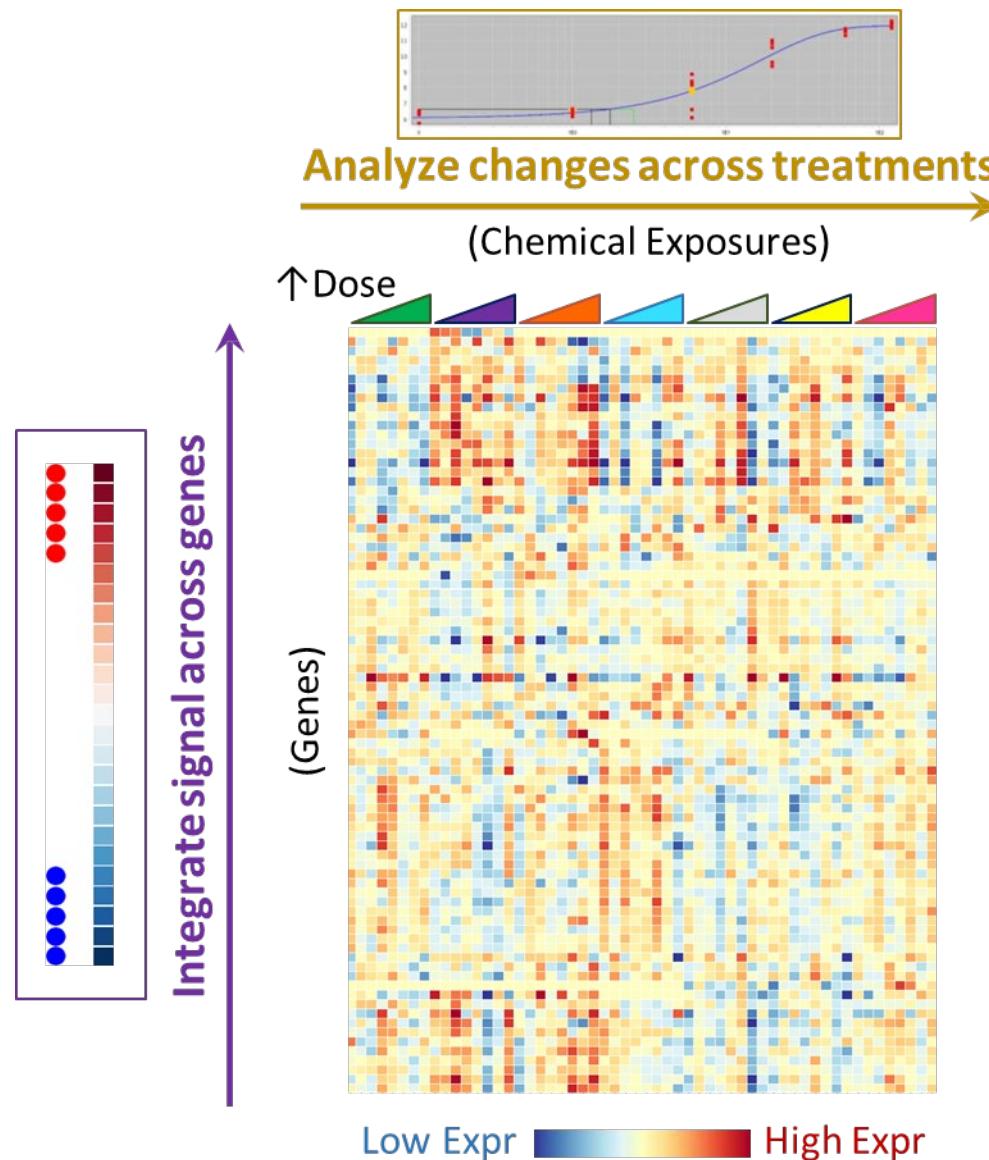
# Targeted RNA-seq Assay (TempO-seq)

- Next-Gen sequencing of targeted probes hybridized to expressed transcripts
- Whole transcriptome coverage (>20,000 genes)
- Captures gene expression at lower cost than RNA-seq or microarrays
- Compatible with raw cell lysates – ***ideal for large-scale screening***



Yeakey, et al. PLoS ONE (2017) DOI: [10.1371/journal.pone.0178302](https://doi.org/10.1371/journal.pone.0178302)

# Transcriptomic Dose-Response Models



- Different genes may respond at different doses of a given exposure!
- Need to analyze both:
  - Dose-responsive trends
  - Coordinated changes in gene expression
- Gene-level data noisier in transcriptomics than targeted measurements (e.g. RT-qPCR)
- Dose-response modeling thousands of features (e.g. mRNA levels) leads to computational & statistical challenges

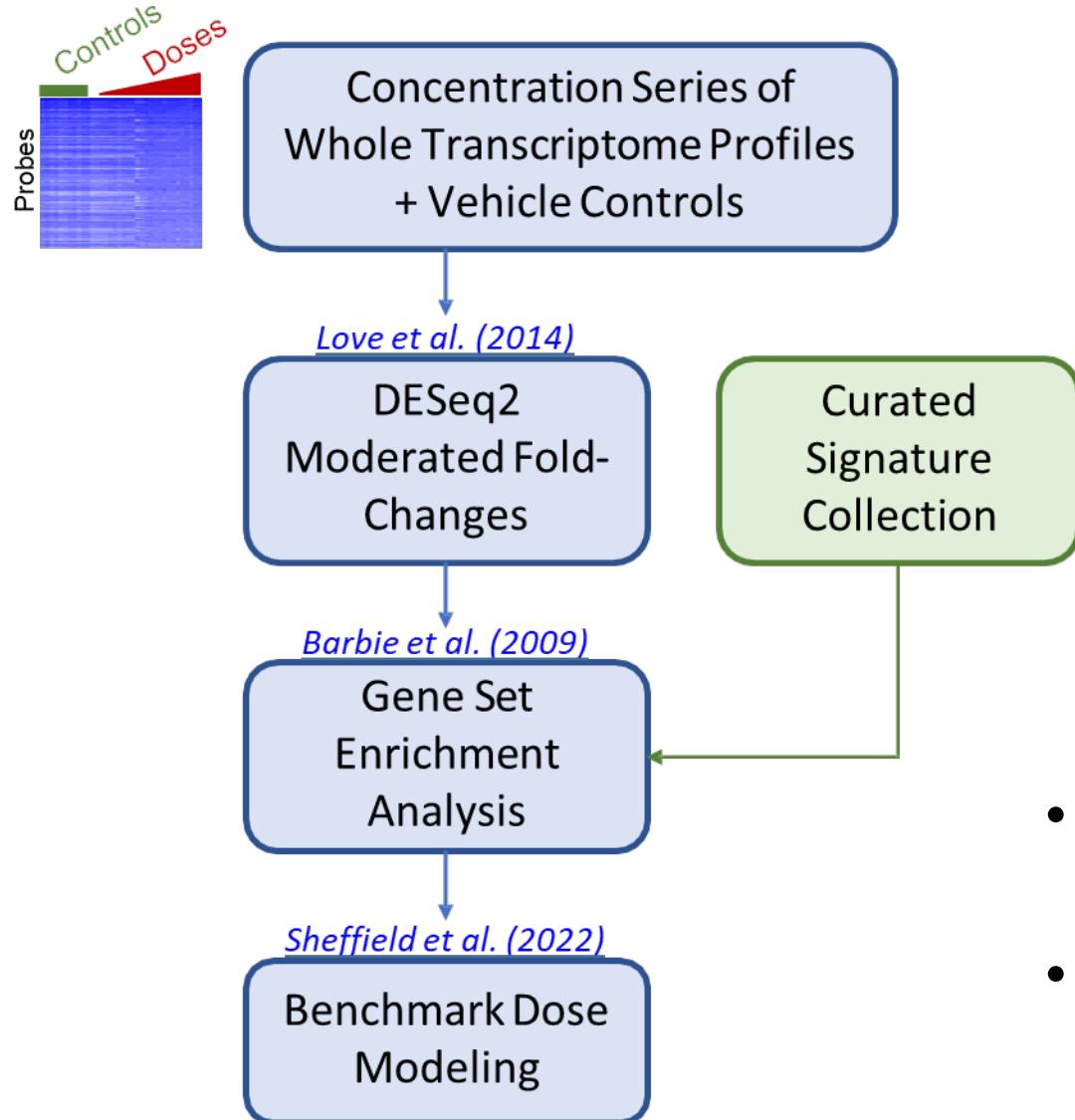
# Many Analysis Choices!



No single “best” method for transcriptomic dose response modeling

- Are you **interested in mechanism**, or just want a threshold for general bioactivity?
- Is it more important to be **predictive** or **protective** of hazard level *in vivo*?
- What other data is available for the same/analogous chemicals?
- Different technologies require different statistical models, quality control, etc.
- Experimental design (*# of replicates, doses, etc.*) impacts analysis choices!

# Dose-Response Modeling of Gene Sets/Signatures



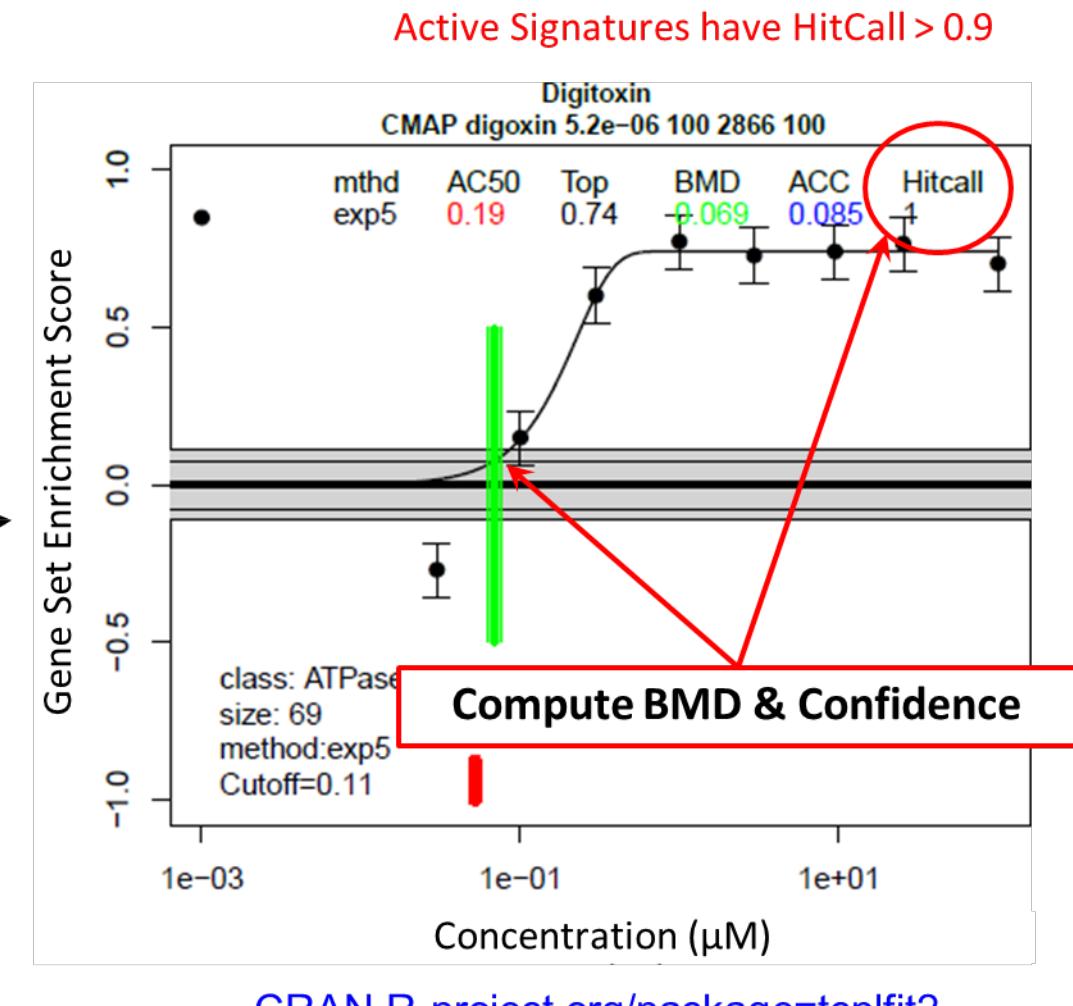
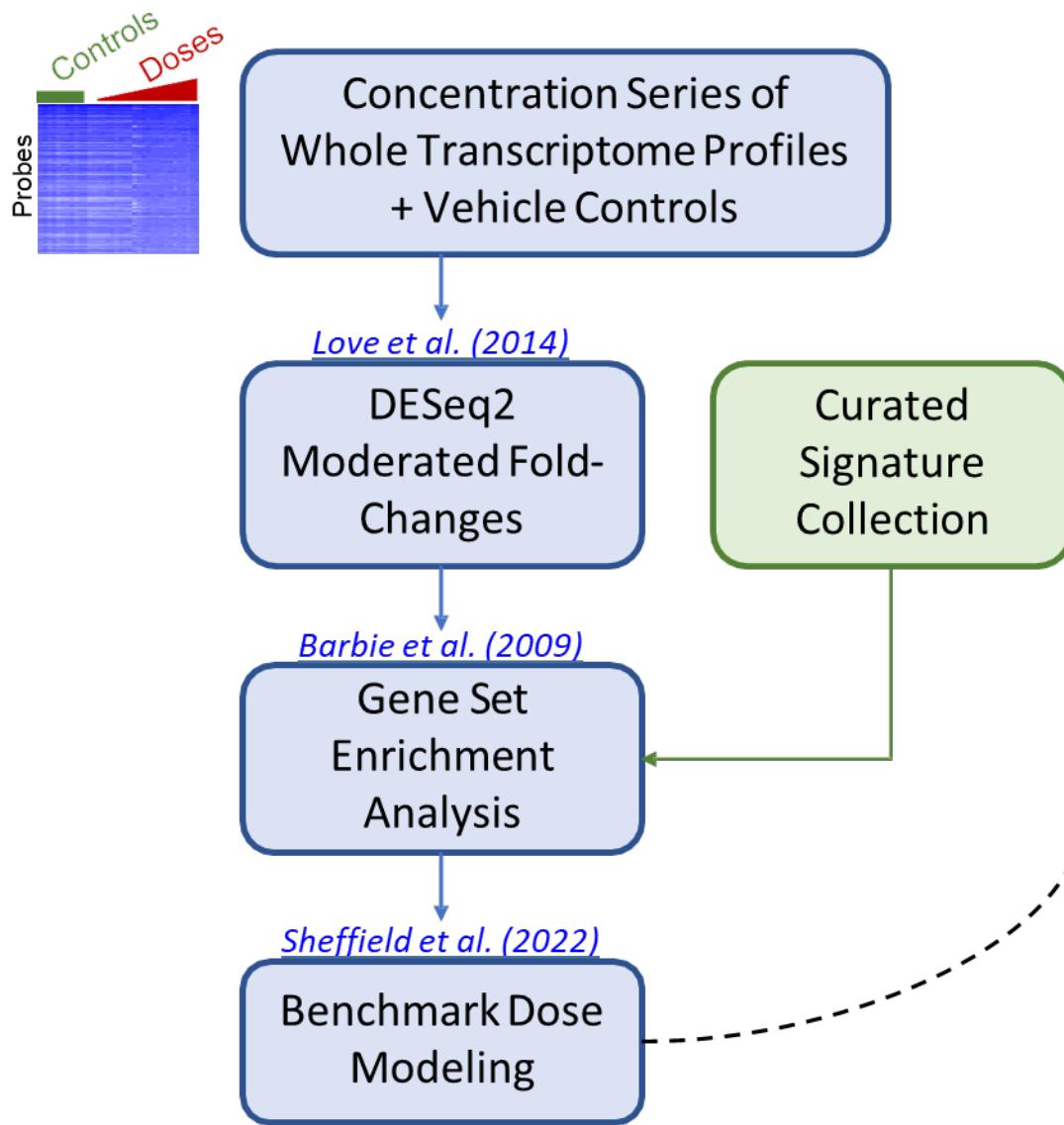
Catalog of gene set signatures with toxicological relevance, annotated for known molecular targets

- [Bioplanet](#) ([Huang, et al. Front Pharmacol 2019](#))
- [CMap](#) ([Subramanian, et al. Cell 2017](#))
- [DisGeNET](#) ([Piñero, et al. Database 2015](#))
- [MSigDB](#) ([Liberzon, et al. Cell Syst 2015](#))

Open Source: [github.com/USEPA/CompTox-httrpathway](https://github.com/USEPA/CompTox-httrpathway)

- EPA/CCTE method for summarizing large-scale transcriptomic screening studies
- Integrates signal across known gene set (a.k.a. signature) **before** dose-response modeling

# Dose-Response Modeling of Gene Sets/Signatures



# Public Release of HTTr Data

Cell Line/Model	Chemicals Screened	Publications
<b>MCF7: Breast adenocarcinoma cell line, sensitive to multiple endocrine disruptors</b>	44 well-characterized chemicals (pilot study)	Harrill, et al. <i>Tox Sci</i> 2021 DOI: <a href="https://doi.org/10.1093/toxsci/kfab009">10.1093/toxsci/kfab009</a> Harrill, et al. <i>Toxicology</i> 2024 DOI: <a href="https://doi.org/10.1016/j.tox.2023.153694">10.1016/j.tox.2023.153694</a>
	1,751 ToxCast chemicals	Harrill, et al. <i>Tox Sci</i> 2024 DOI: <a href="https://doi.org/10.1093/toxsci/kfae108">10.1093/toxsci/kfae108</a>
<b>U-2 OS: Osteosarcoma cell line with complementary Cell Painting data</b>	1,201 ToxCast chemicals	Bundy, et al. <i>TAAP</i> 2024 DOI: <a href="https://doi.org/10.1016/j.taap.2024.117073">10.1016/j.taap.2024.117073</a>
<b>HepaRG: Hepatoma cell line, differentiated into metabolically competent hepatocyte-like cells</b>	1,201 ToxCast chemicals	Rogers, et al. <i>in review</i> Shah, et al. <i>in preparation</i>

# HTTr Results Available on CompTox Chemicals Dashboard

[comptox.epa.gov/dashboard](https://comptox.epa.gov/dashboard)

① Search for a specific chemical –

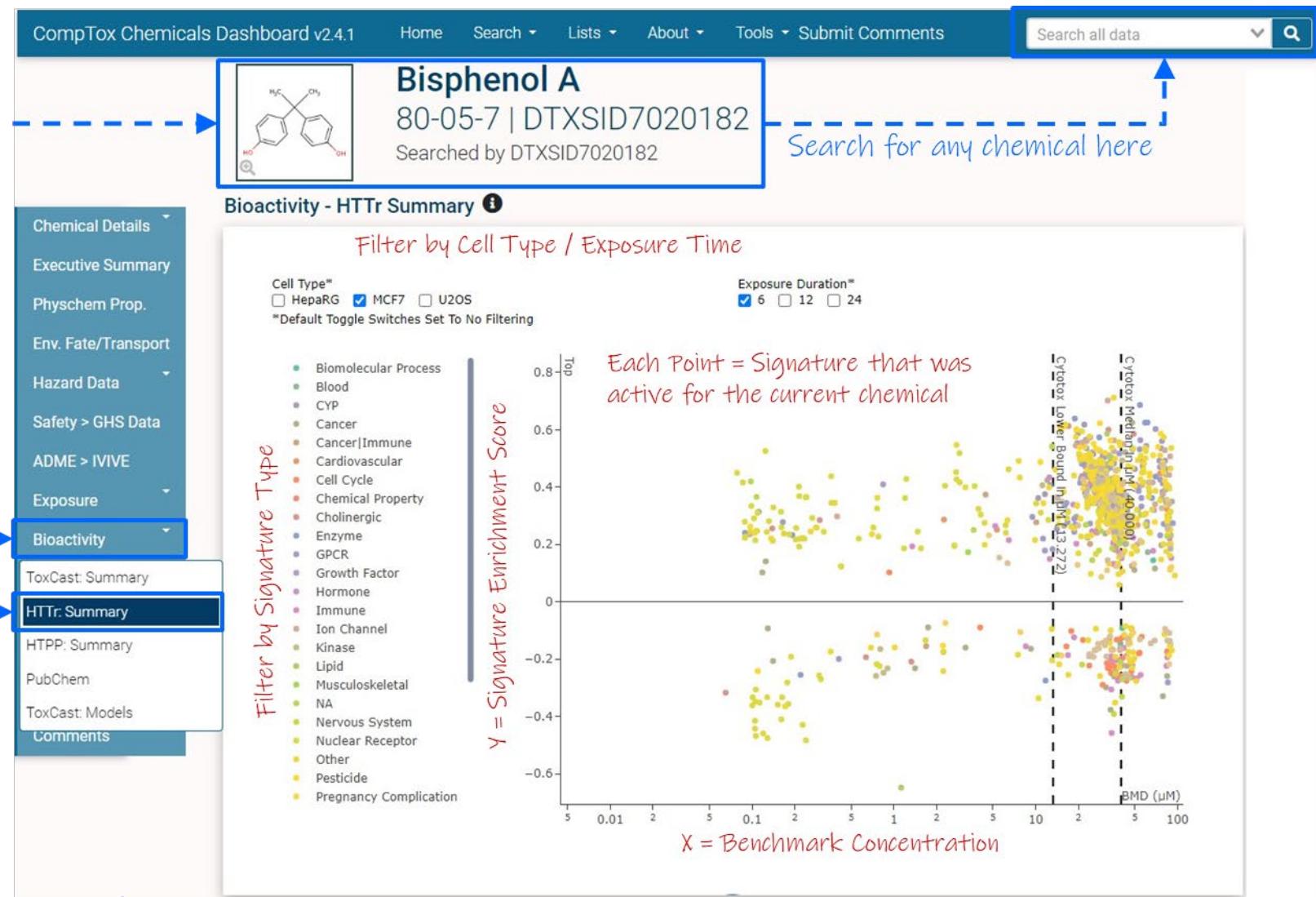
② All *in vitro* NAMs data available here

③ HTTr data available here –



← More Info Available Here

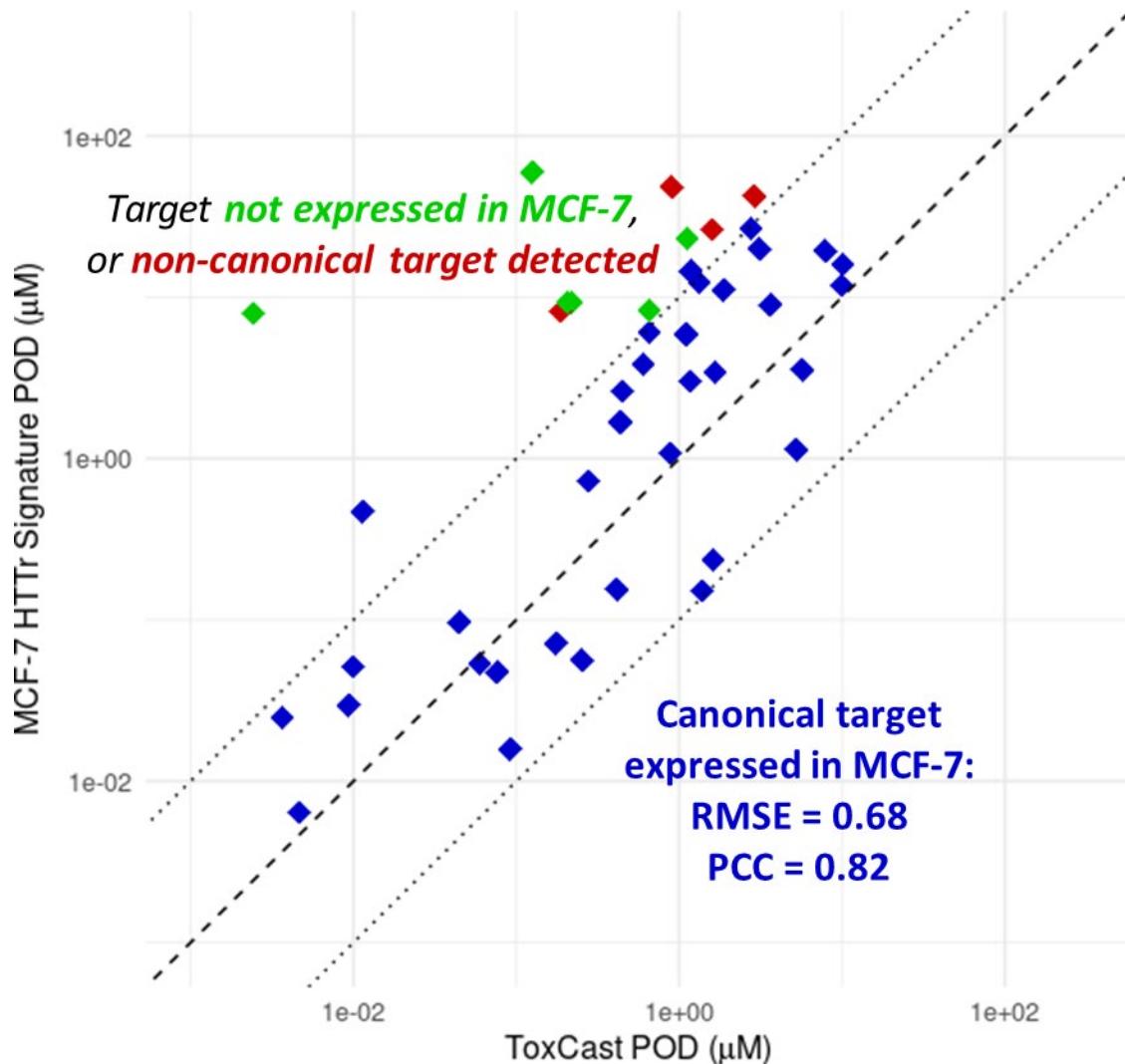
[epa.gov/chemical-research/high-throughput-toxicology](https://epa.gov/chemical-research/high-throughput-toxicology)



Office of Research and Development



# HTTr tPODs Are Concordant With ToxCast PODs



- Computed 5<sup>th</sup> percentile PODs from:
  - Pilot study of 44 well-characterized chemicals in MCF-7 cells, 6h exposure  
Harrill, et al. *Toxicol Sci* (2021)  
DOI: [10.1093/toxsci/kfab009](https://doi.org/10.1093/toxsci/kfab009)
  - ToxCast targeted assay results (*multiple cell types, assays, and exposure lengths*)  
Paul-Friedman, et al. *Toxicol Sci* (2020)  
DOI: [10.1093/toxsci/kfz201](https://doi.org/10.1093/toxsci/kfz201)
- Signature-based PODs are highly concordant with ToxCast results for the majority of test chemicals in pilot study

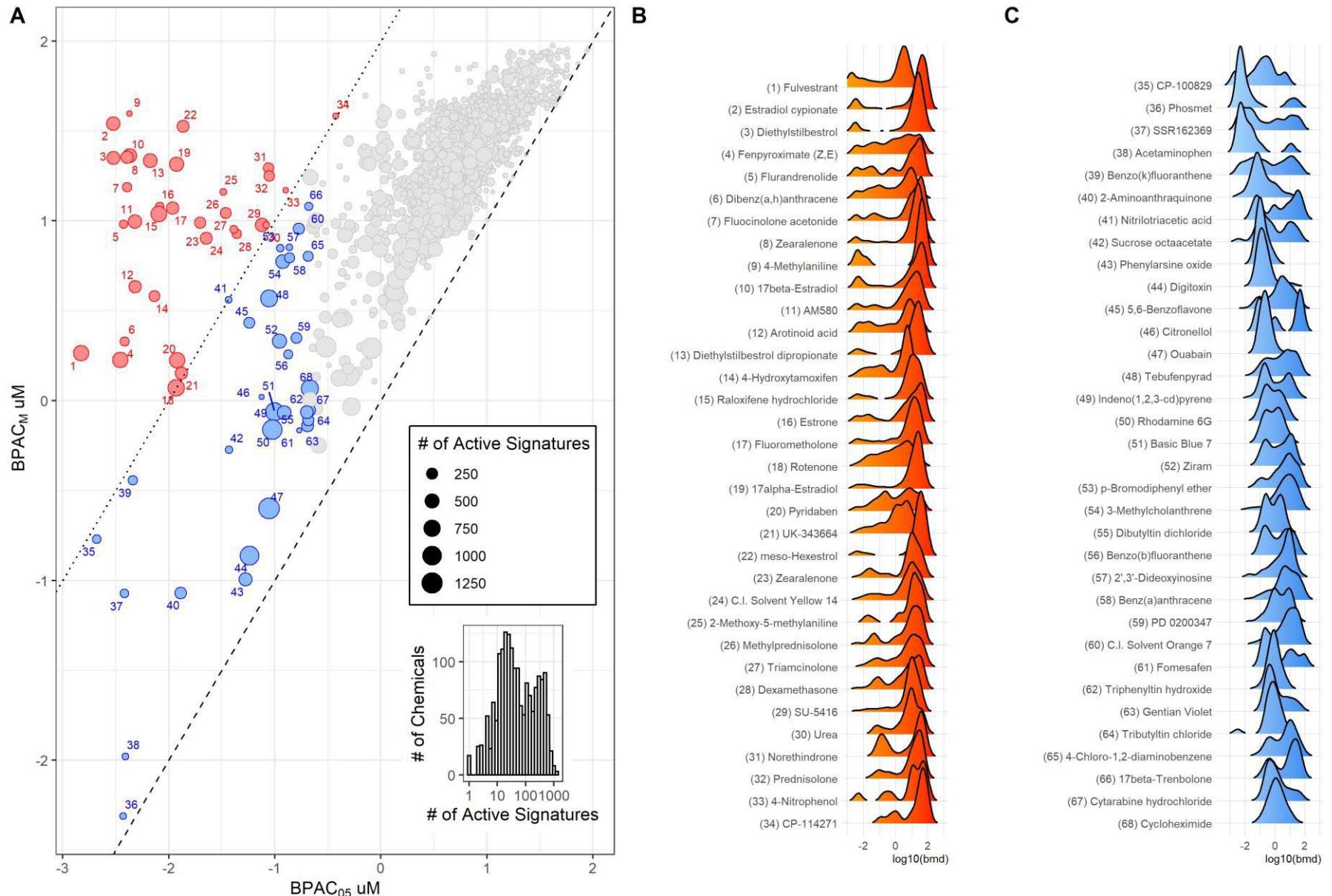
# Signature BMC Distributions Vary Across Chemicals

Results from first large screen of 1,751 chemicals in MCF7 cells

**BPAC** = Biological Pathway Altering Concentration.

Chemicals with known molecular target specificity tended to have  $BPAC_{05}$  much more potent than the median BPAC (red bubbles)

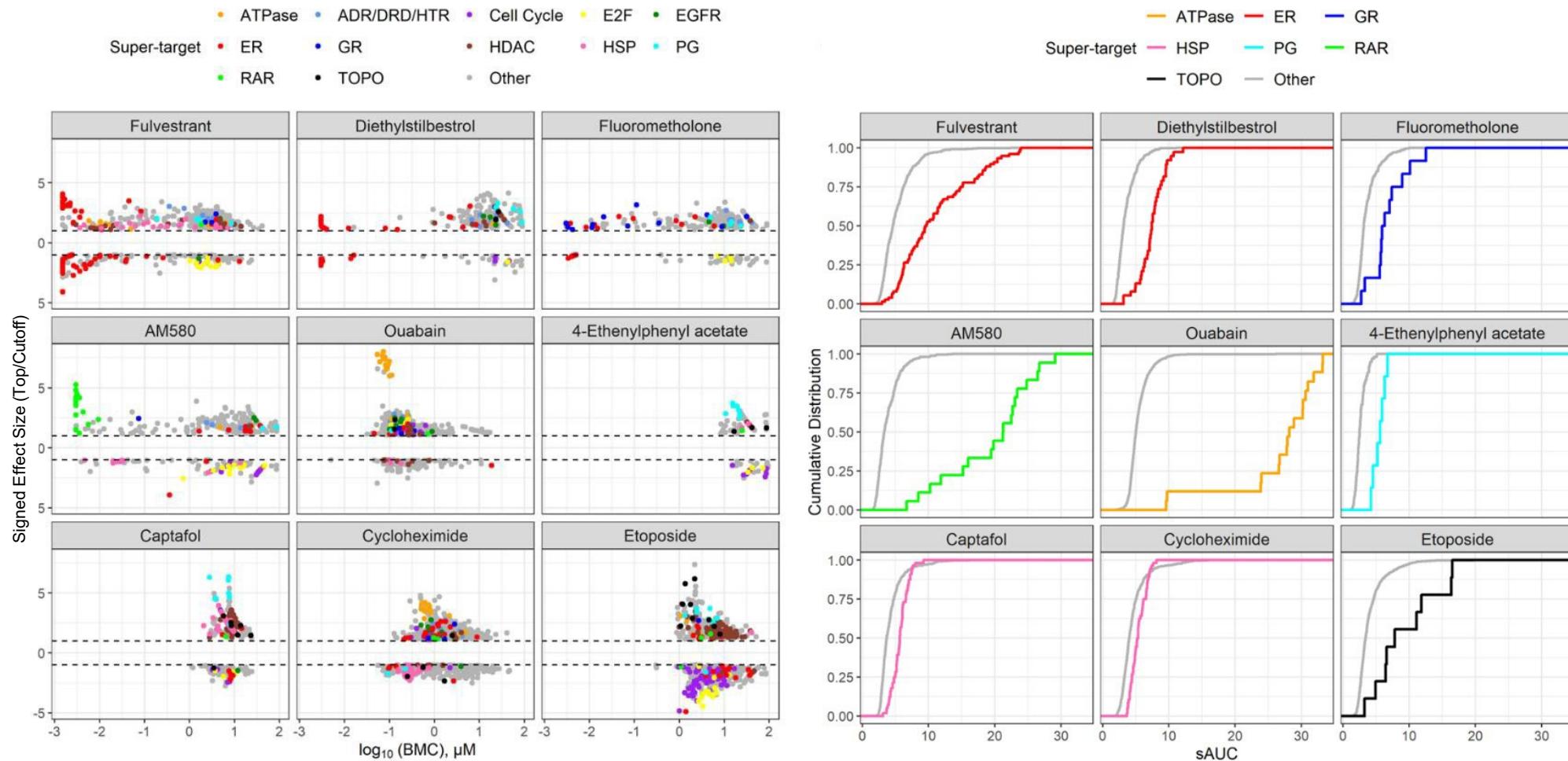
Harrill et al. Tox Sci (2024)  
DOI: [10.1093/toxsci/kfae108](https://doi.org/10.1093/toxsci/kfae108)



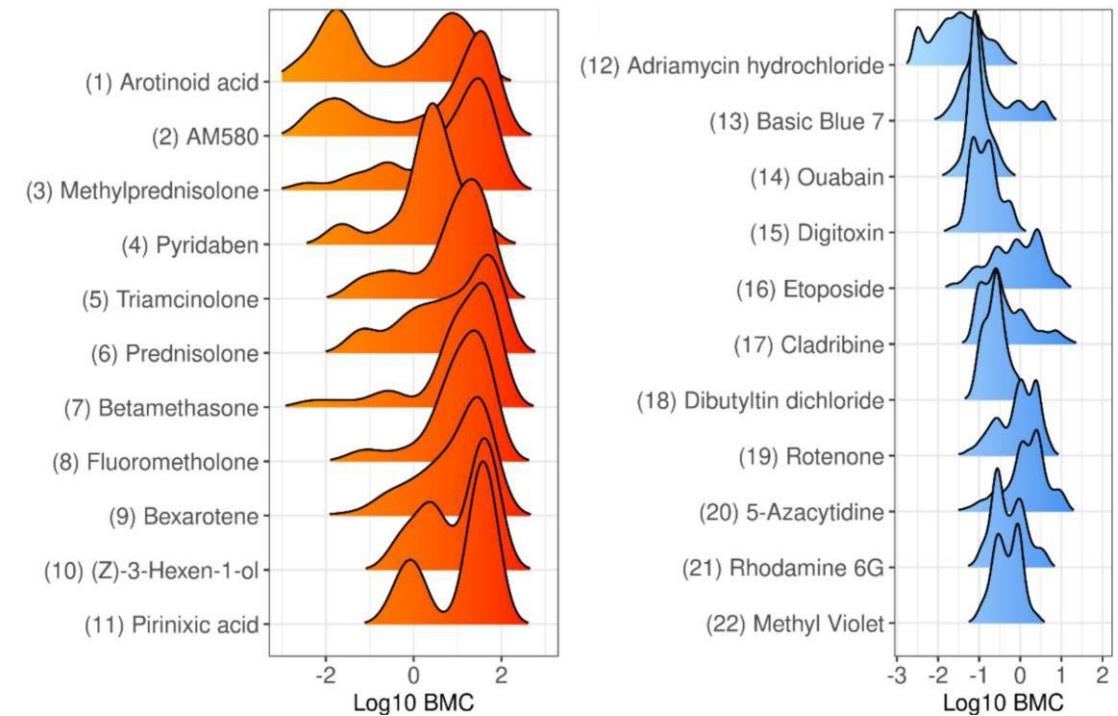
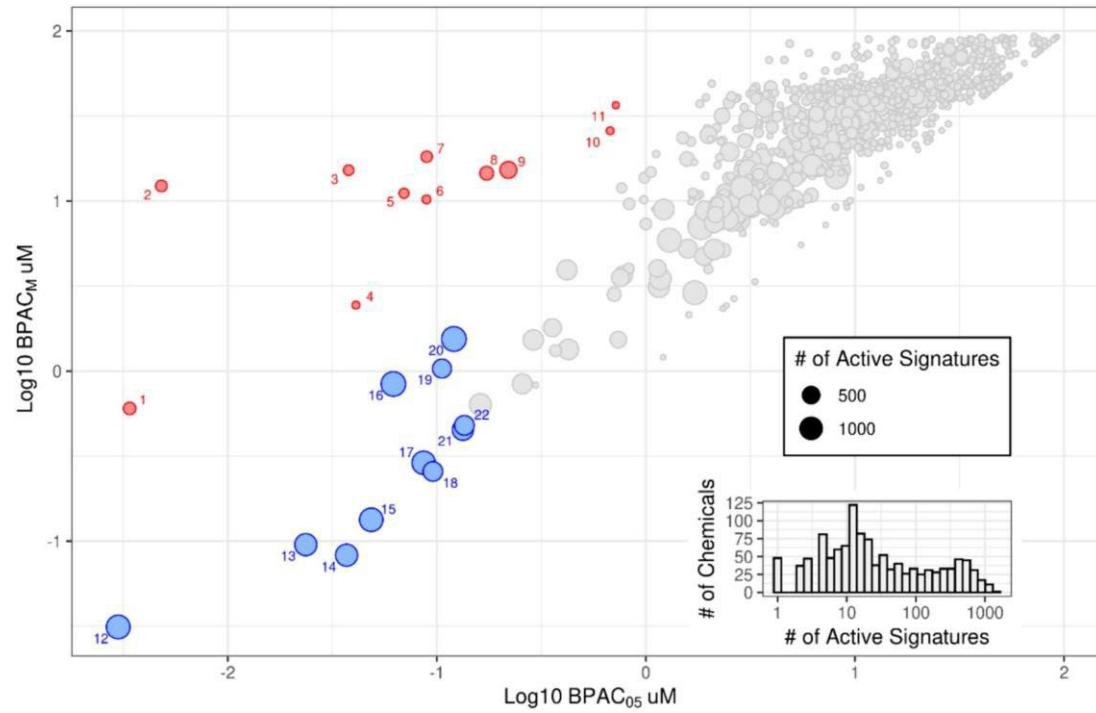
# Distribution of Signature BMCs Indicative of Molecular Target Specificity

Target annotations in existing signature catalog identify target specificity for some chemicals.

Tested for enrichment of signatures with same target annotation by K-S test on AUC values (lower BMD or greater effect size)



# Application of Signature Analysis Method to Additional Cell Types



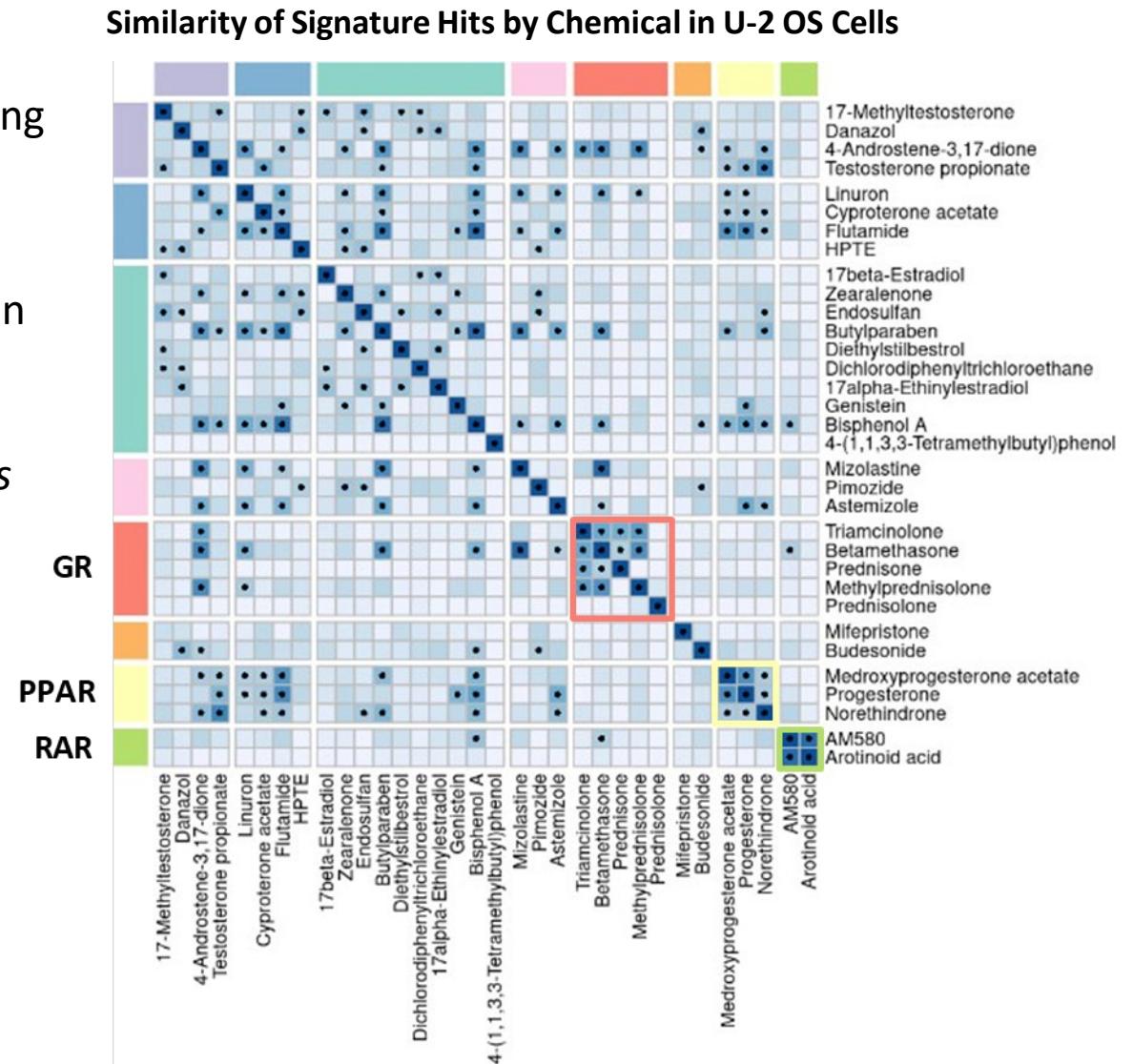
Observed similar distributions of BPAC05 and BPACM in **U-2 OS cells** as in MCF7:

- Some chemicals have clear delineation between target-specific and non-specific activity (red)
- Other chemicals have potent but non-specific activity (blue)

# Predicting Molecular Targets Using Signature Modeling and Reference Chemicals

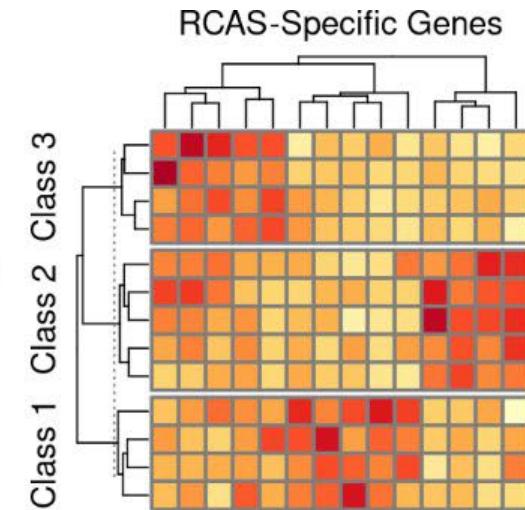
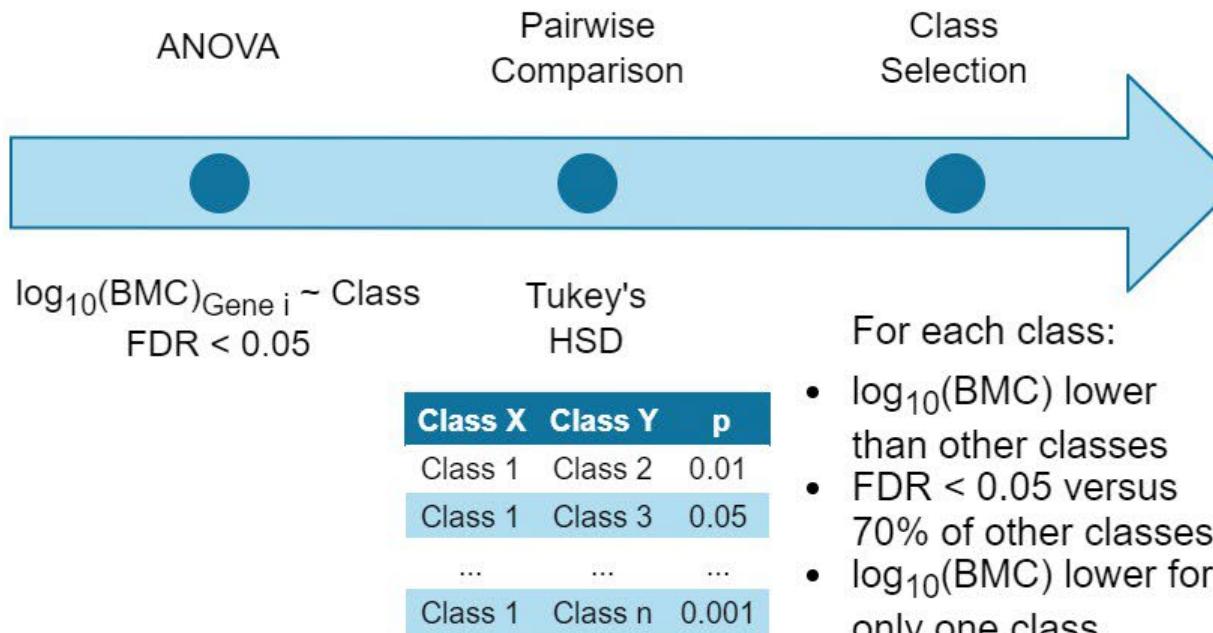
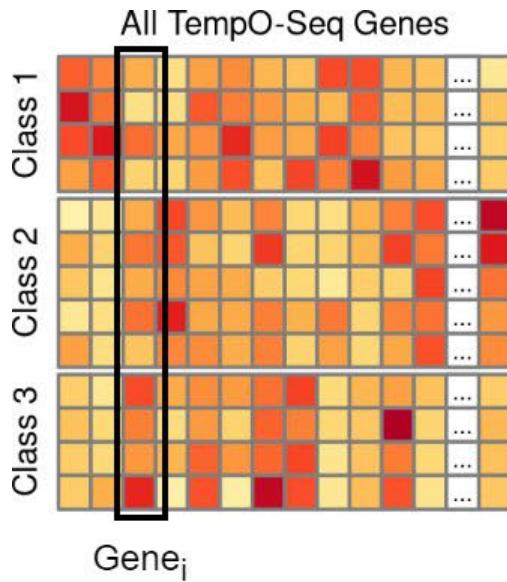
- Existing signature annotations are less effective at inferring chemical target in other cell types  
*(many signatures derived from studies in MCF-7)*
- However, similarity of overall signature activity profile can still be used to group chemicals sharing certain common targets

*Pairwise similarity = Jaccard Index of active signature hits  
Results shown from U-2 OS (osteosarcoma cell line)*

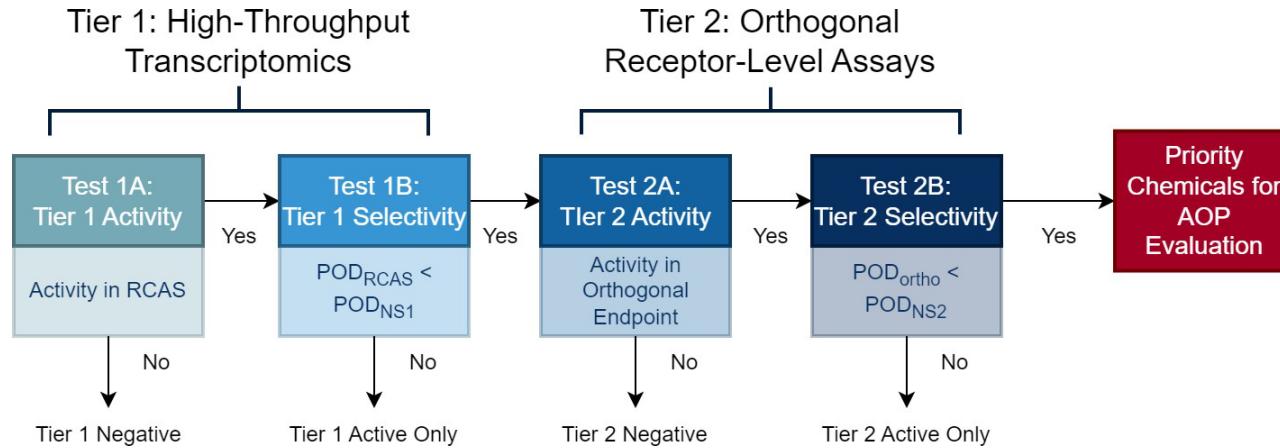


# Reference Class-Associated Signatures (RCAS) for Profiling Chemical Mechanisms-of-Action

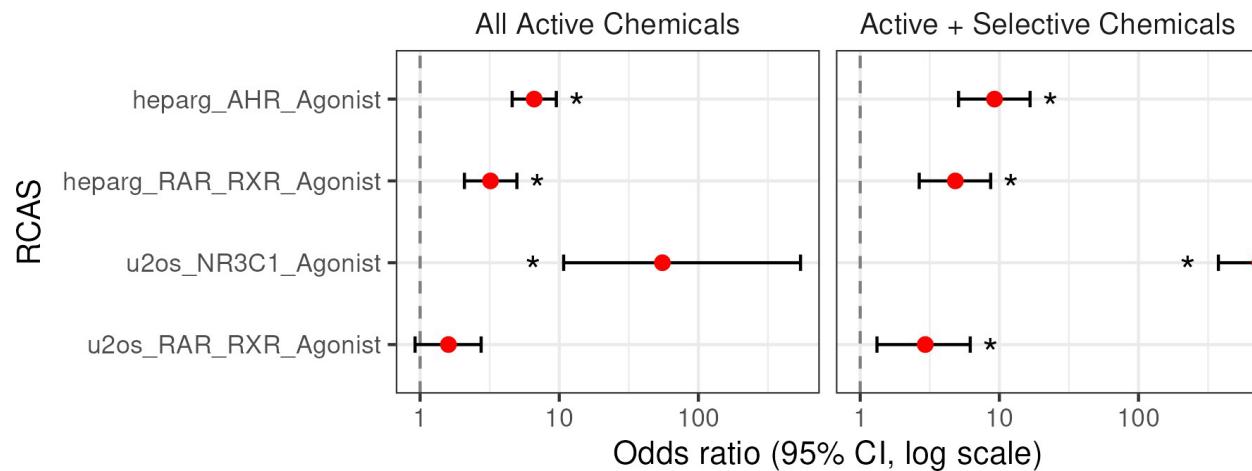
- Goal: Develop new target-specific signatures tailored to each cell line (applied to U-2 OS and HepaRG so far)
- Genes uniquely sensitive for individual molecular targets selected via univariate analysis
- Tailored signatures identify selective chemical effects during Tier 1 screening



# Integration of TempO-Seq Readouts for Chemical Prioritization Framework



RCAS-derived potencies compared to non-selective point of departure (POD<sub>NS</sub>) estimated from public signature collection



Chemicals with selective effects on molecular targets identified via RCAS and validated using high-throughput screening assay data from US EPA's ToxCast program

Rogers, et al. Manuscript submitted

# Summary

- High-Throughput Transcriptomics (HTTr) applied to human cells *in vitro* captures a wide variety of biological perturbations and can be used to:
  - Derive an overall “mechanism-agnostic” point of departure (POD/BPAC)
  - Detect perturbation of specific targets/mechanisms
- Ongoing work at US EPA:
  - Screening in additional cell types (e.g. hTERT lines)
  - Benchmarking/harmonization of analysis methods in specific contexts
  - Development of additional signatures/models for mechanistic inference

# Acknowledgements

## CCTE Leadership

Rusty Thomas  
Sid Hunter  
Kimberly Slentz-Kesler  
Katie Paul Friedman  
John Cowden

## HTTr Team

Joshua Harrill  
Richard Judson  
Imran Shah  
Derik Haggard  
Beena Vallanat  
Joseph Bundy  
Jesse Rogers  
Jacob Fredenburg  
Sarah Davidson-Fritz

Bryant Chambers  
Laura Taylor  
James Johnson  
Felix Harris  
Clinton Willis  
Thomas Sheffield  
Khan Inan  
Monique Hazemi  
Joshua Witten



## Questions?

[everett.logan@epa.gov](mailto:everett.logan@epa.gov)



# Use of 'Omics to Inform Development and Safety of New Pesticide Active Ingredients

JESSICA LAROCCA  
CORTEVA AGRISCIENCE

# Global Regulatory Agencies



Only 1 in 139,000  
chemicals successfully  
progresses through the  
regulatory process from the  
laboratory to the field

# Agrochemical *In Vivo* Mammalian Toxicology Requirements

Task Description	OECD Guidelines	Species	Males		Females	
			Min	Max	Min	Max
Global Toxicokinetic Registration PK/Met (Default study)	OECD 417 (2010)	Rat	8	32	8	32
Acute Oral Toxicity (Default Rat)	OECD 423 (2001)	Rat	0	0	3	24
Acute Dermal Toxicity	OECD 402 (2017)	Rat	0	0	1	7
Acute Inhalation Toxicity	OECD 403 (2009)	Rat	3	15	3	15
Acute Dermal Irritation	OECD 404 (2015)	Rabbit	0	0	1	3
Acute Eye Irritation	OECD 405 (2017)	Rabbit	0	0	1	3
Dermal Sensitization (LLNA)	OECD 429 (2010)	Mouse	0	0	25	33
Acute Neurotoxicity (Rat)	OECD 424 (1997)	Rat	40	40	40	40
28-Day Mouse Oral + Palatability	OECD 407 (2008)	Mouse	20	30	20	30
28-Day Rat Oral	OECD 407 (2008)	Rat	20	40	20	40
13-Week Dog Oral	OECD 409 (2018)	Dog - Beagle	16	24	16	24
13-Week Mouse Oral	OECD 408 (2018)	Mouse	40	50	40	50
13-Week Rat Oral	OECD 408 (2018)	Rat	40	50	40	50
SubChronic Neurotoxicity	OPPTS 870.6200	Rat	120	150	120	150
28-Day Rat Dermal	OECD 410 (1981)	Rat	20	25	20	25
1-Year Chronic Dog	OECD 452 (2018)	Dog - Beagle	16	21	16	21
In vivo Mouse Micronucleus	OECD 474 (2016)	Mouse	25	35	25	35
2-Year Chronic Rat/Carcinogenicity	OECD 453 (2018)	Rat	220	310	220	310
18-Month Carcinogenicity Mouse	OECD 451 (2018)	Mouse	200	265	200	265
Dev Screen OECD 421	OECD 421 (2016)	Rat	68	68	68	68
Reproduction Study (2 gen)	OECD 416 (2001)	Rat	120	120	120	120
Rat Dev Tox Full (Teratogenicity)	OECD 414 (2018)	Rat	0	0	88	88
Rabbit Dev Tox Full (Teratogenicity)	OECD 414 (2018)	Rabbit	0	0	25	25
28-Day Stand-Alone Immunotoxicity (Sheep Red Blood Cell)	OPPTS 870.7800	Mouse or Rat	0	50	50	50



# NAMs for Predictive Toxicology Studies to Reduce Animal Use

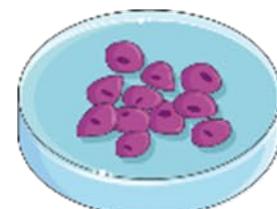


## New Approach Methodologies (NAMs)

In Silico



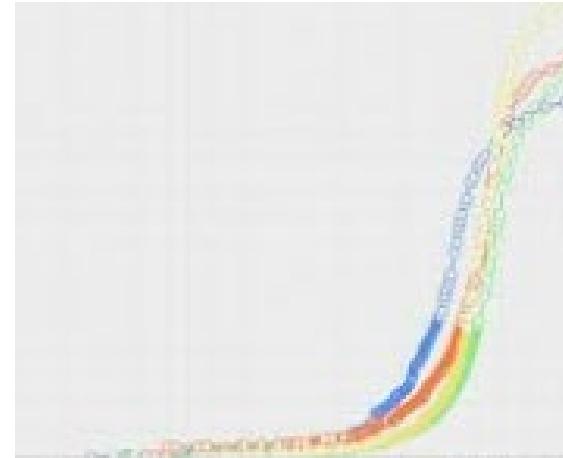
In Vitro and  
Aquatic



In Vivo



# Transcriptomics to Predict Points of Departure

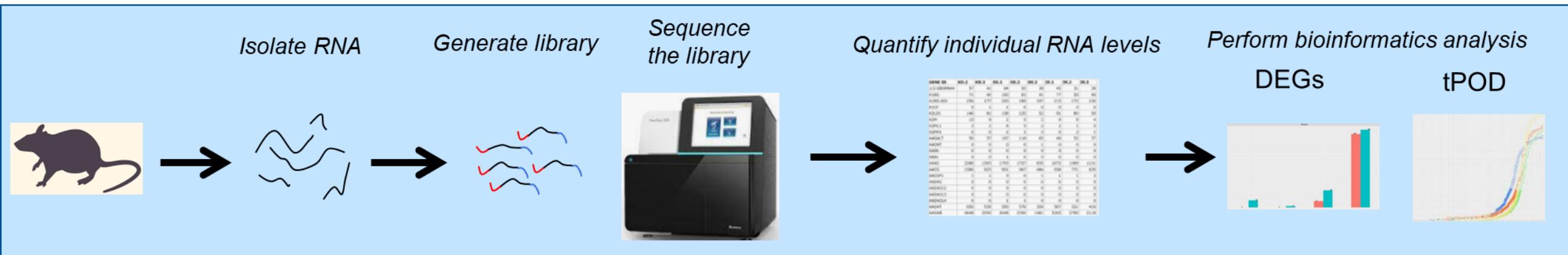


# Transcriptomics Is a Tool to Comprehensively Examine Change in Gene Expression (RNA levels)

*Central Dogma of Molecular Biology*



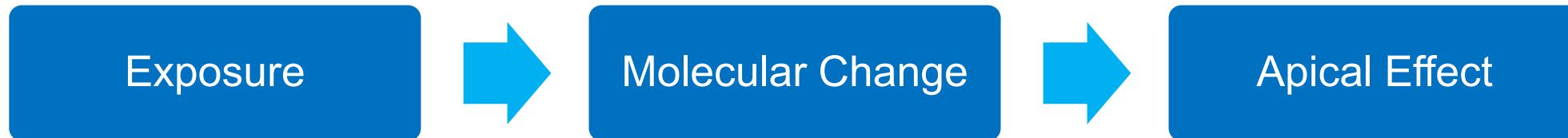
**Typically, 13,000 – 16,000 unique RNA molecules are observed per organ**



DEG: Differentially Expressed Gene

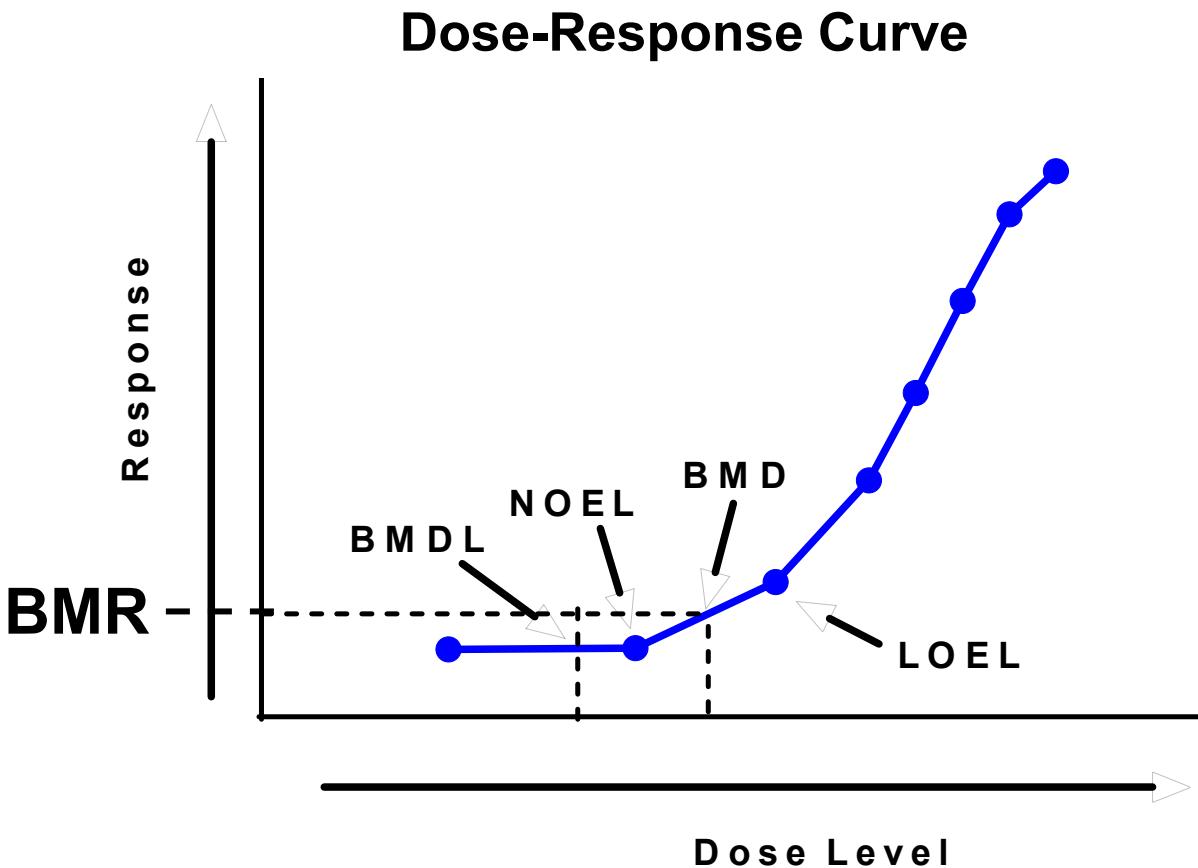
# All Apical Effects Result From A Prior Change At The Molecular Level

## Generic Adverse Outcome Pathway



Therefore, changes observed in **comprehensive** molecular data (like toxicogenomics) will detect any apical effect (however, the identity of the apical effect is unknown).

# A Point of Departure Can Be Determined Using NOEL or BMD Approaches



**NOEL: No Observed Effect Level**

**LOEL: Lowest Observed Effect Level**

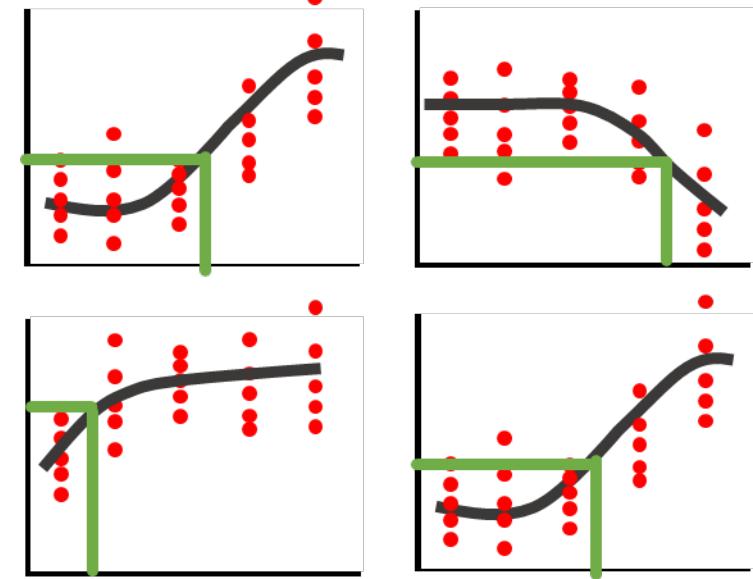
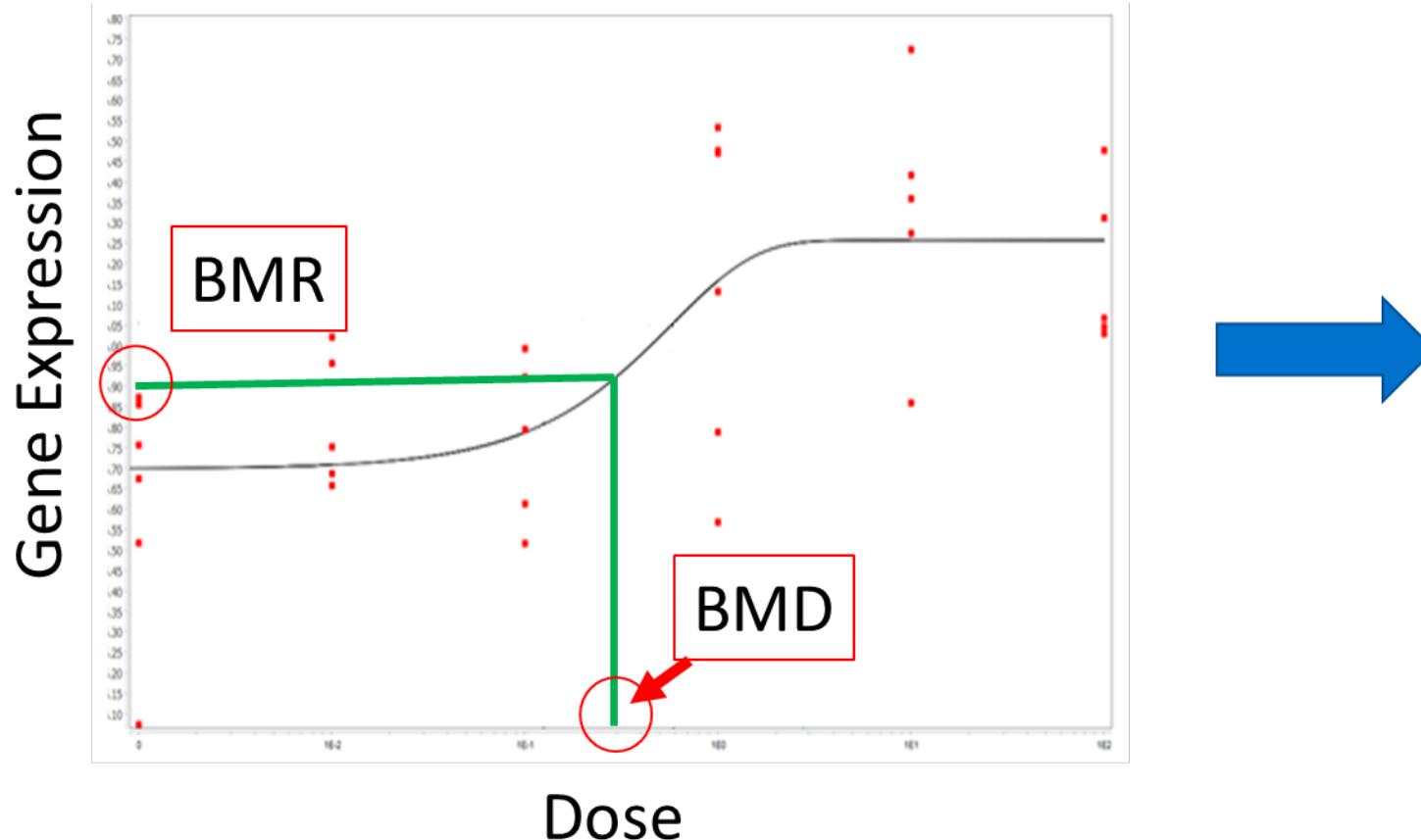
**BMD: Benchmark Dose**

**BMDL: Benchmark Dose Lower Confidence Limit**

**BMR: Benchmark Response**

# What is a **Transcriptome Point of Departure (tPOD)**?

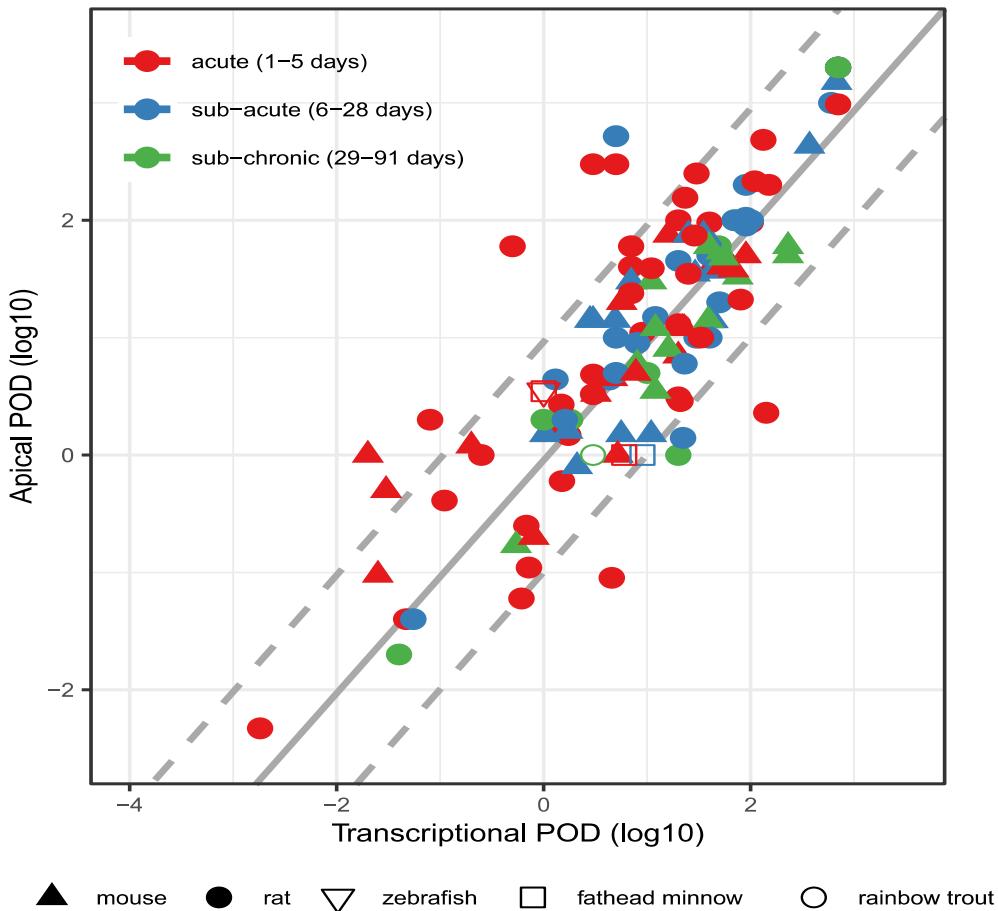
- Estimates the lowest dose for a “concerted” change in gene expression



# A tPOD Accurately Estimates a Traditional Apical Endpoint POD

Systematic literature review of >100 articles (from Jason O'Brien)

- $R^2 > 0.80$
- 95% within 10-fold of traditional POD
- >88% within 5-fold of traditional POD
- High correlation after 1-5 days!!

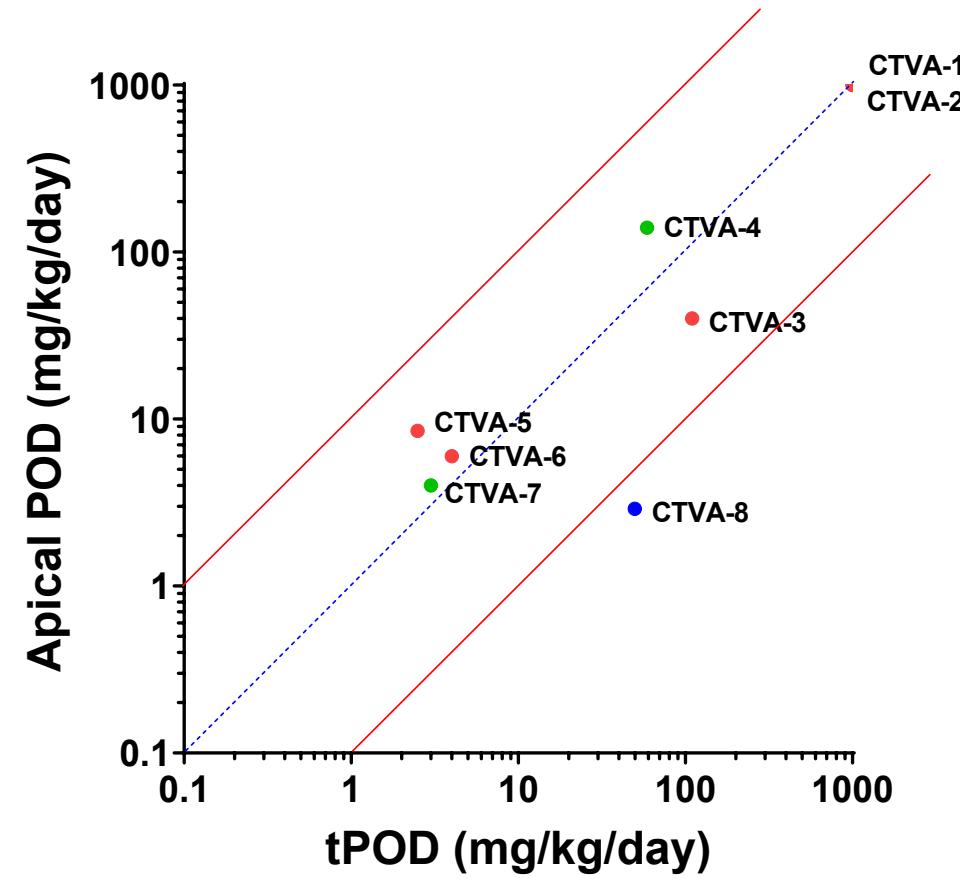


- Concordance of tPOD and a traditional POD is similar to the concordance of traditional PODs from repeating a guideline study

# tPods Accurately Predict Apical PODs for Corteva Agrochemicals

Short-term transcriptomic studies with tPODs can approximate the apical POD from long-term studies (90-day rat, cancer bioassay, two-gen, etc).

**tPOD vs Apical POD**



# 5-day Toxicogenomic Study Design to Inform Target Organs and PoDs

- Standardized study design that can be used for novel compounds with unknown hazard and PoDs
- Short-term study can be used to inform PoDs resulting from chronic exposure (and inform risk assessments)
- **Rat 5-Day TGx Study Design** Dietary exposure
  - Males only (animal reduction purposes) (5/group)
  - Organ weights collected from all organs in TGx study and perform histopathology on liver, kidney, and expected target organs, if known
  - Seven default organs analyzed via RNAseq – can be modified based upon prior information
  - 6 – 7 dose levels tested

Organs collected for RNAseq			
Liver (49%)	Kidney (19%)	Thyroid (14%)	Testis (3%)
Adrenal (3%)	Spleen (2%)	Epididymis	Pituitary

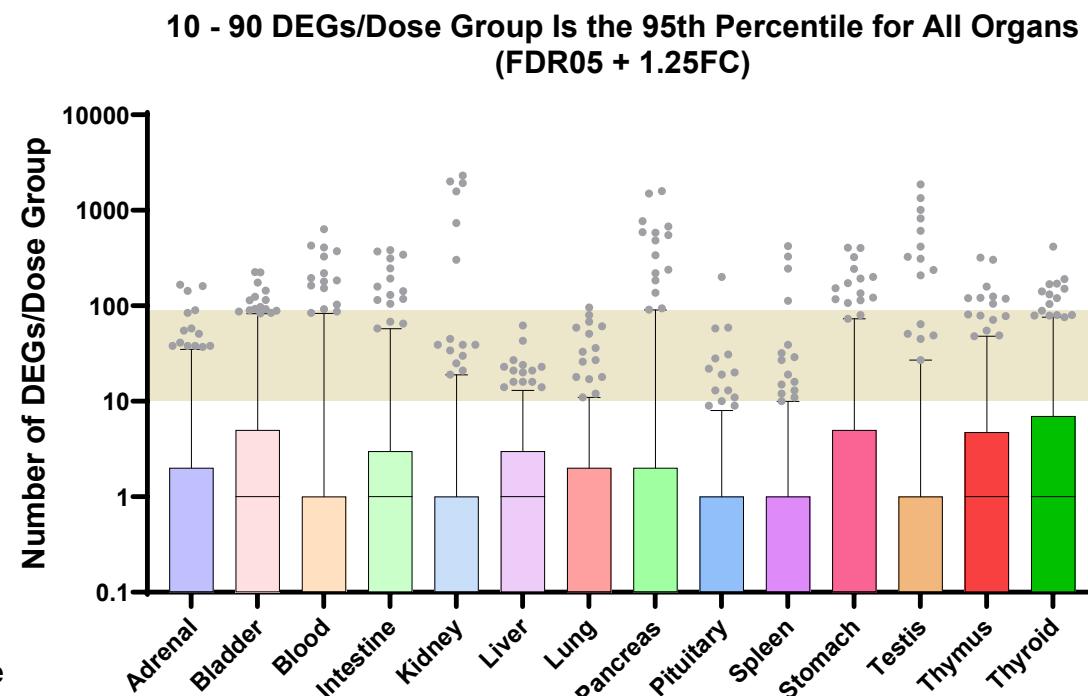
Number in parentheses is the % of 435 pesticides that target that organ.

# Rat 5-Day Toxicogenomics Study Control Data Analysis

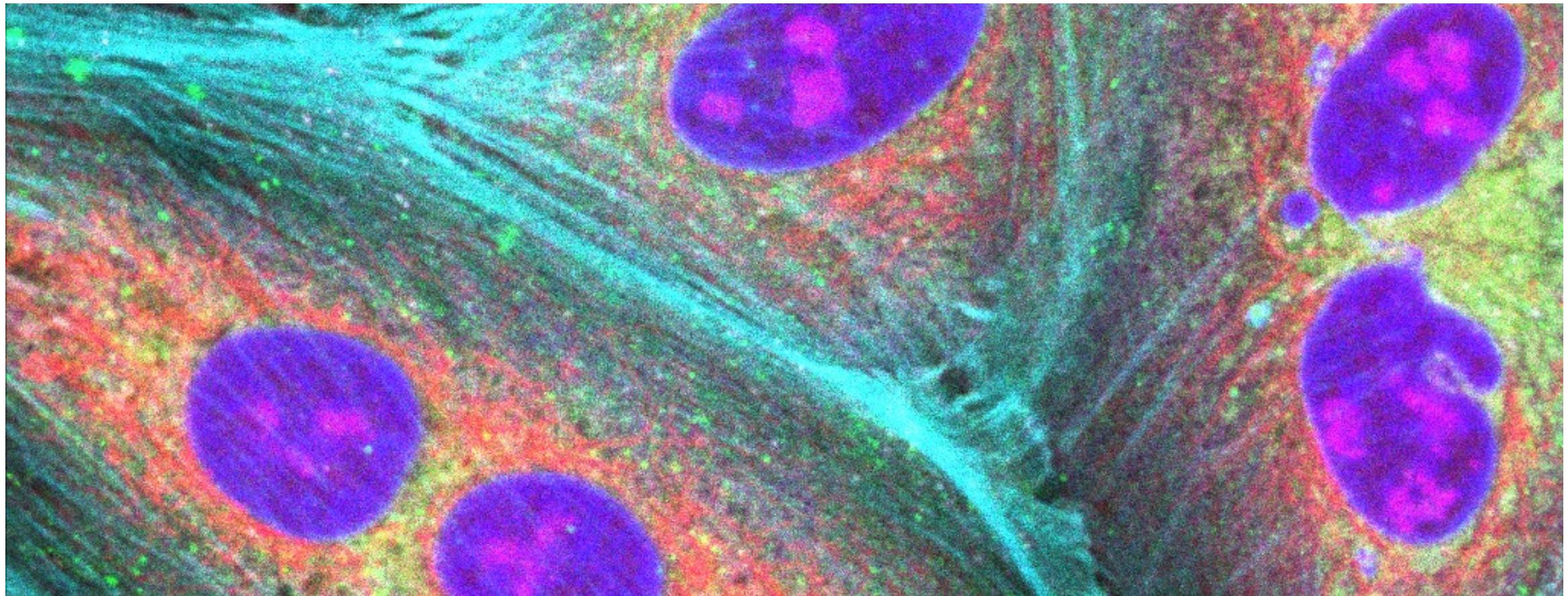
If a tPOD is identified in an organ, it will be concluded that an apical effect (adaptive or adverse) is present after 5-days of exposure or will occur with a longer duration of exposure.

A tPOD will be derived if more than 90 DEGs are identified in an organ at the highest dose level tested.

- Data in graph below are from a control (unexposed) rat multi-organ RNAseq study (i.e. no exposure) performed at Corteva.
- In all organs, identification of 90 DEGs was the 95<sup>th</sup> percentile upper bound in control rat organ RNAseq

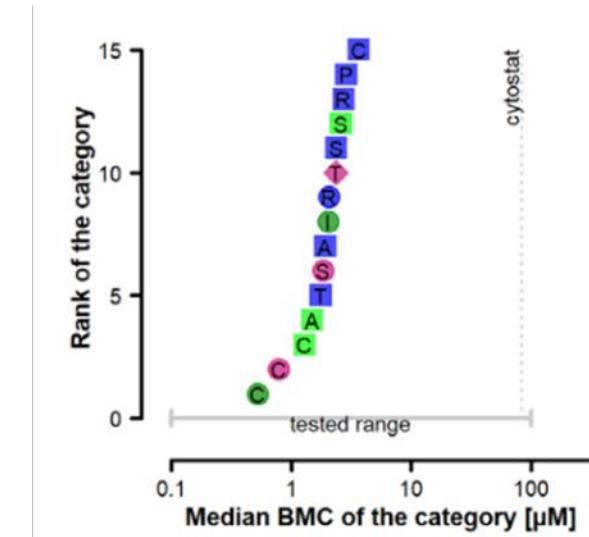
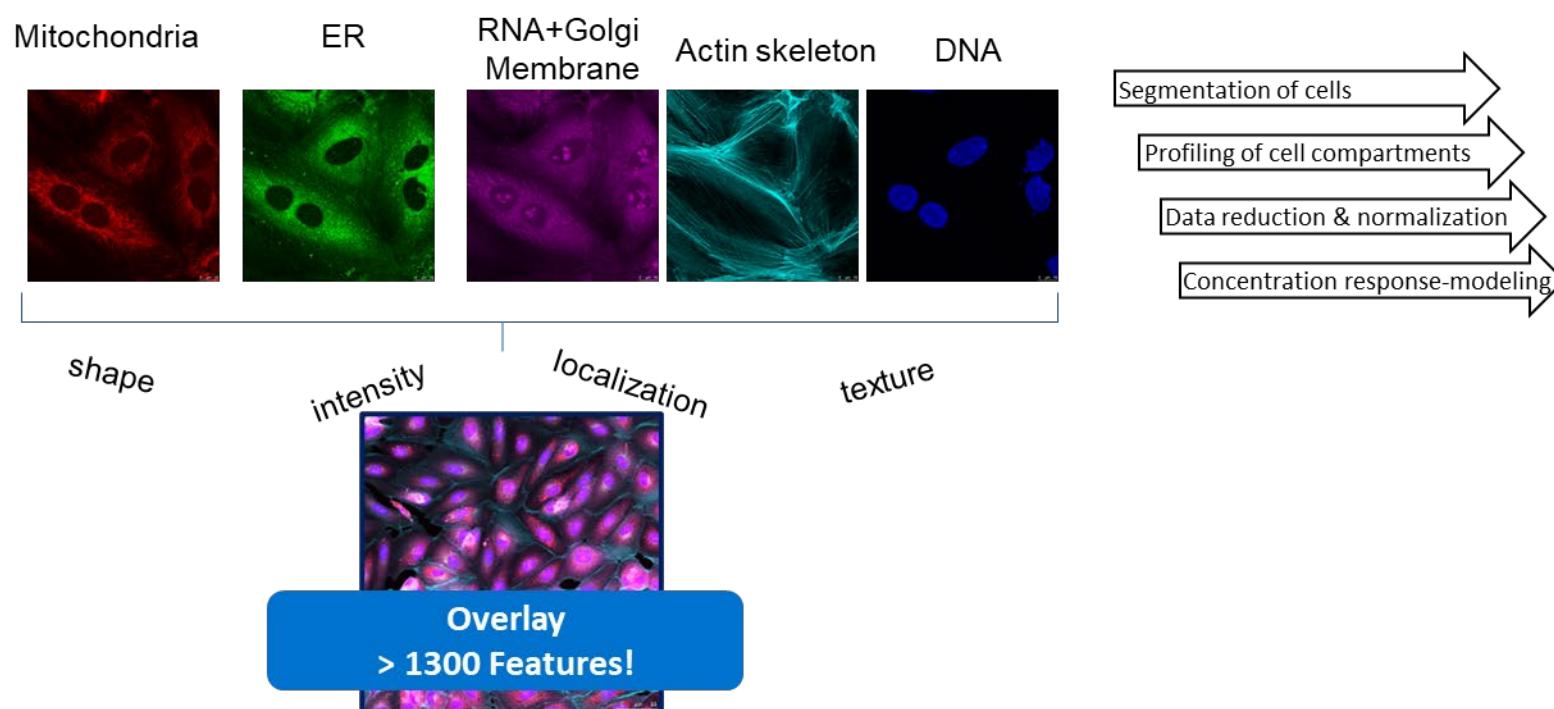


# Cell Painting Toxicophenomics to Predict PODs



# Cell Painting: A high-throughput method to inform toxicological-induced cellular perturbations

- Cell painting is a HCl technique that involves staining of various cell organelles with fluorescent dyes.
- Developed by Broad Institute *Nat Protoc. 2016*



Adapted from Willis et al. SLAS DISCOVERY. 2020;25(7):755-69.

# Liver Cell Painting

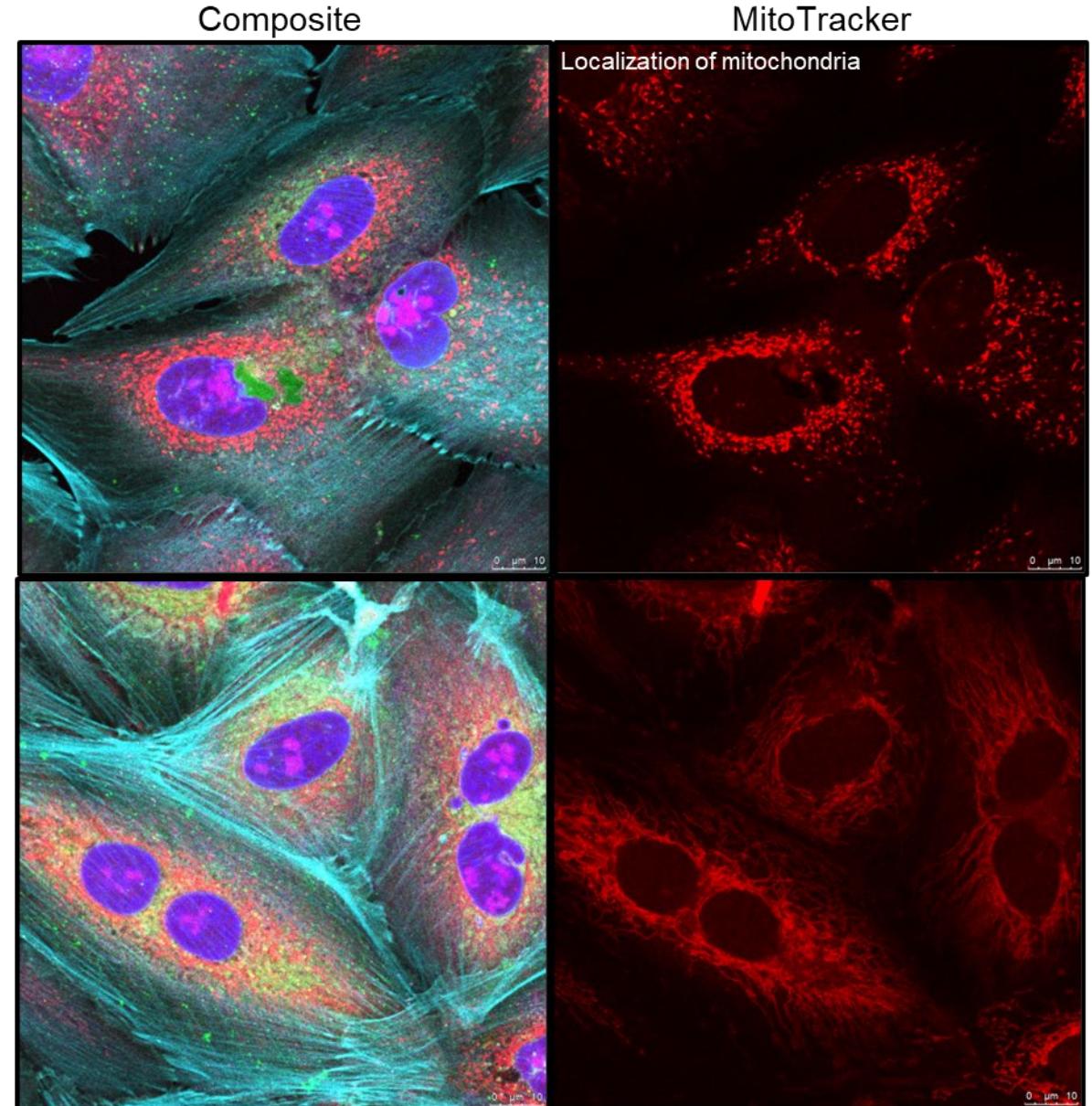
- Liver is the #1 target organ for agrochemicals.
- Many agrochemicals are extensively metabolites, therefore metabolically competent cells are more relevant.
- Therefore, HepaRG liver cells were selected for predictive tox

Channel	Stain	Organelles
405/435-480	Hoechst 33342	Nucleus
488/500-550	Concanavalin A; SYTO14	ER, Nucleoli/RNA
561/570-630	Wheat Germ Agglutinin, Phalloidin	Actin skeleton, Golgi, Plasma membrane
640/650-760	MitoTracker™ Deep Red FM	Mitochondria

Berberine chloride

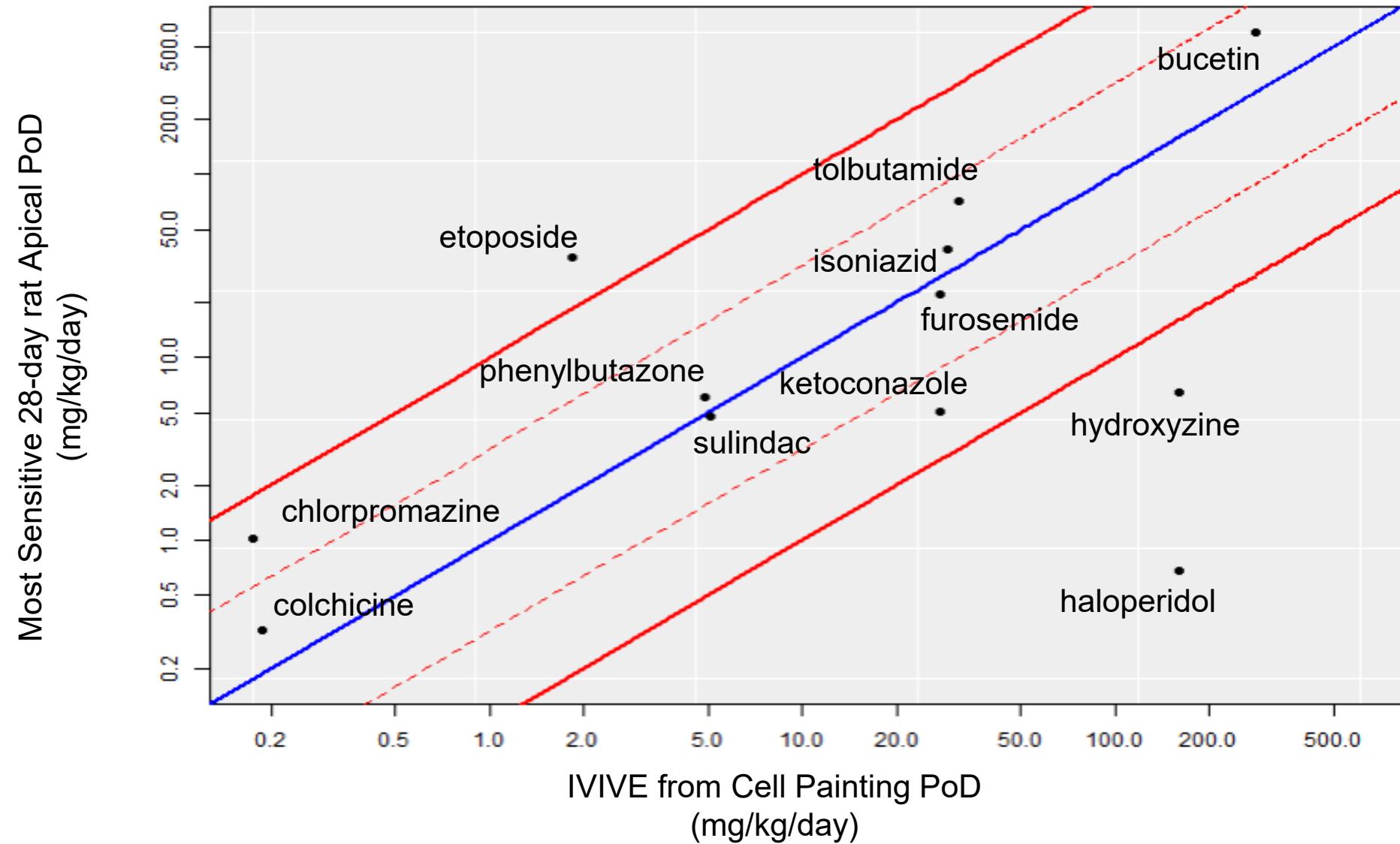
UT Control

Mitochondria  
DNA; ER; Actin  
Golgi; Membrane; RNA



# HepaRG Cell Painting IVIVE Approximates In Vivo Pharmaceutical PODs

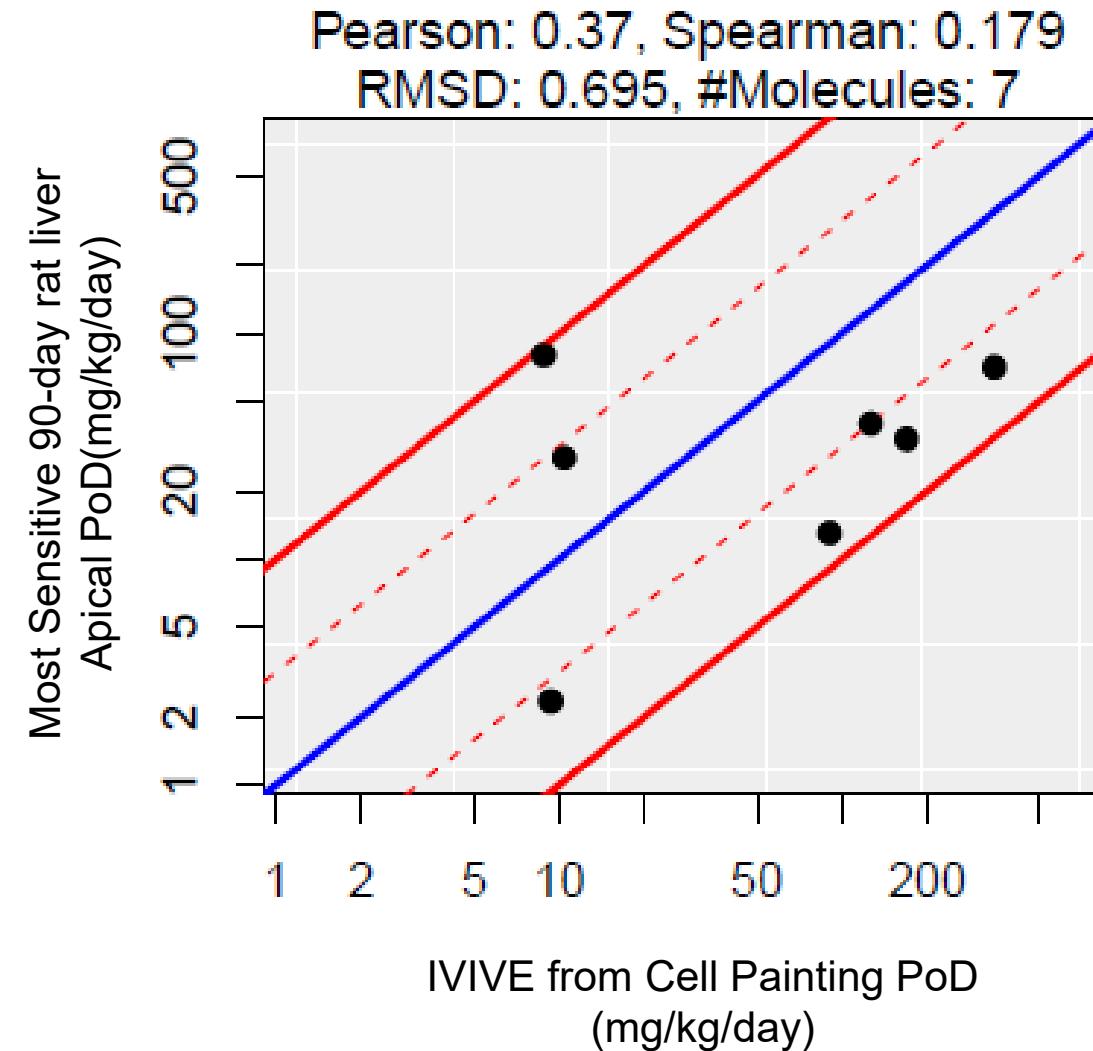
Comparison of cell paint POD with IVIVE with most sensitive 29-day apical endpoint POD (mg/kg/day)



# HepaRG Cell Painting IVIVE Approximates In Vivo Agchem Subchronic PODs

Comparison of cell paint  
POD with IVIVE with most  
sensitive liver 90-day apical  
endpoint POD (mg/kg/day).

Molecules with cell paint  
POD > highest concentration  
tested were excluded

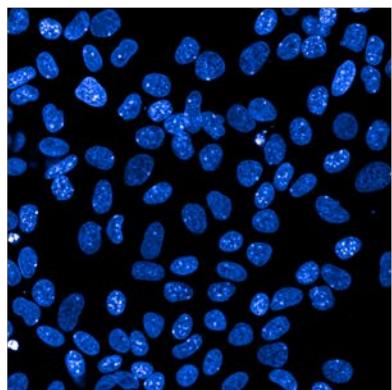


# Machine Learning with Cell Painting to Predict Hazard

# Can We Use Machine Learning to Predict Specific Toxicity Mechanisms/Adverse Outcomes?

Solvent Control

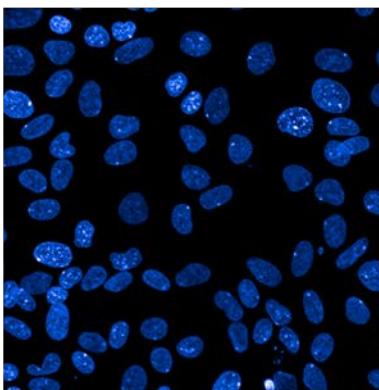
0.5% DMSO



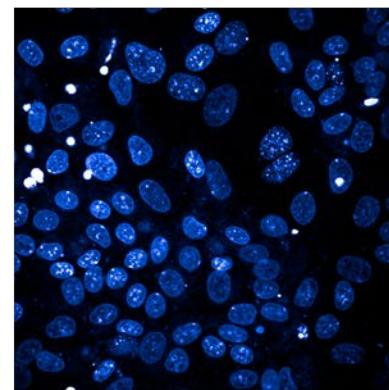
Nuclei (Hoechst)

Ames Negative

50 $\mu$ M Berberine Chloride

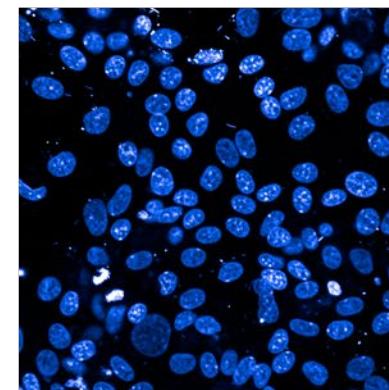


15.8 $\mu$ M Benzo[a]pyrene

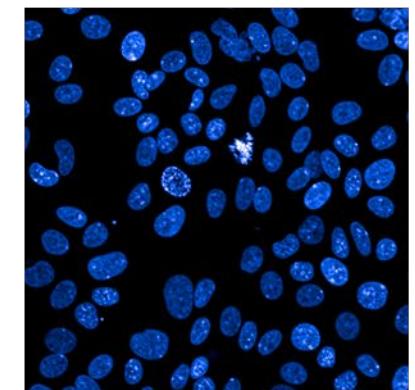


Ames Positive

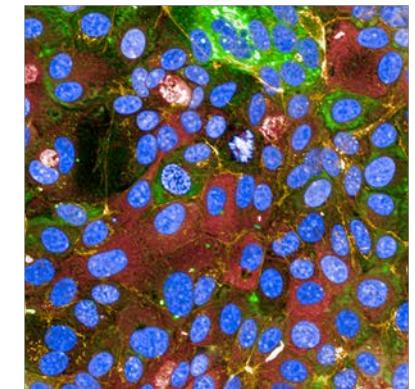
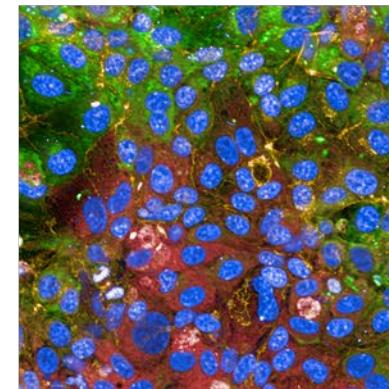
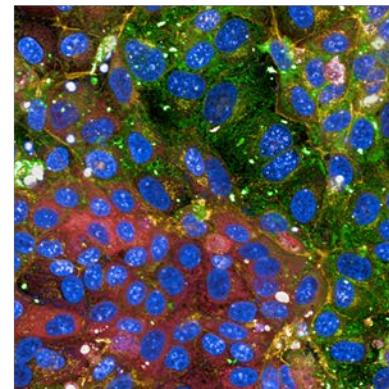
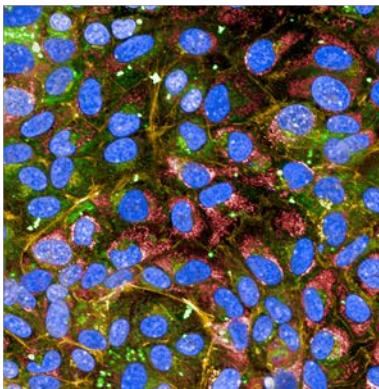
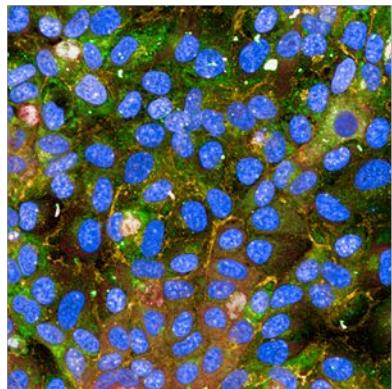
50 $\mu$ M Dibenz[a,h]anthracene



0.5 $\mu$ M 4-Nitroquinoline 1-oxide



Composite



# Cell Painting Machine Learning Can Predict Ames Classification

Training data (well level): 918 Positive V.S. 899 Negative  
(compound level: 104 Pos V.S. 107 Neg)

		Reference	
		Positive	Negative
Predicted	Positive	84	27
	Negative	20	80

Sensitivity/Recall	Specificity	Precision	F1	Balanced Accuracy
0.808	0.748	0.757	0.781	0.777

# HESI OASIS

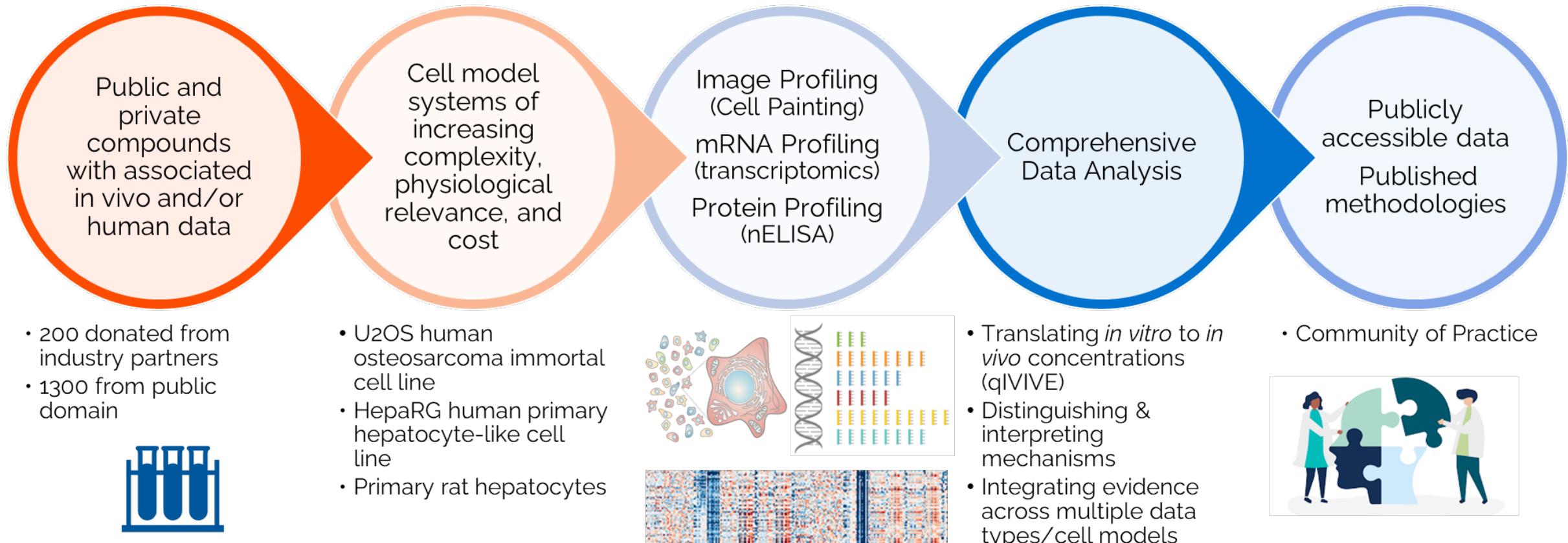
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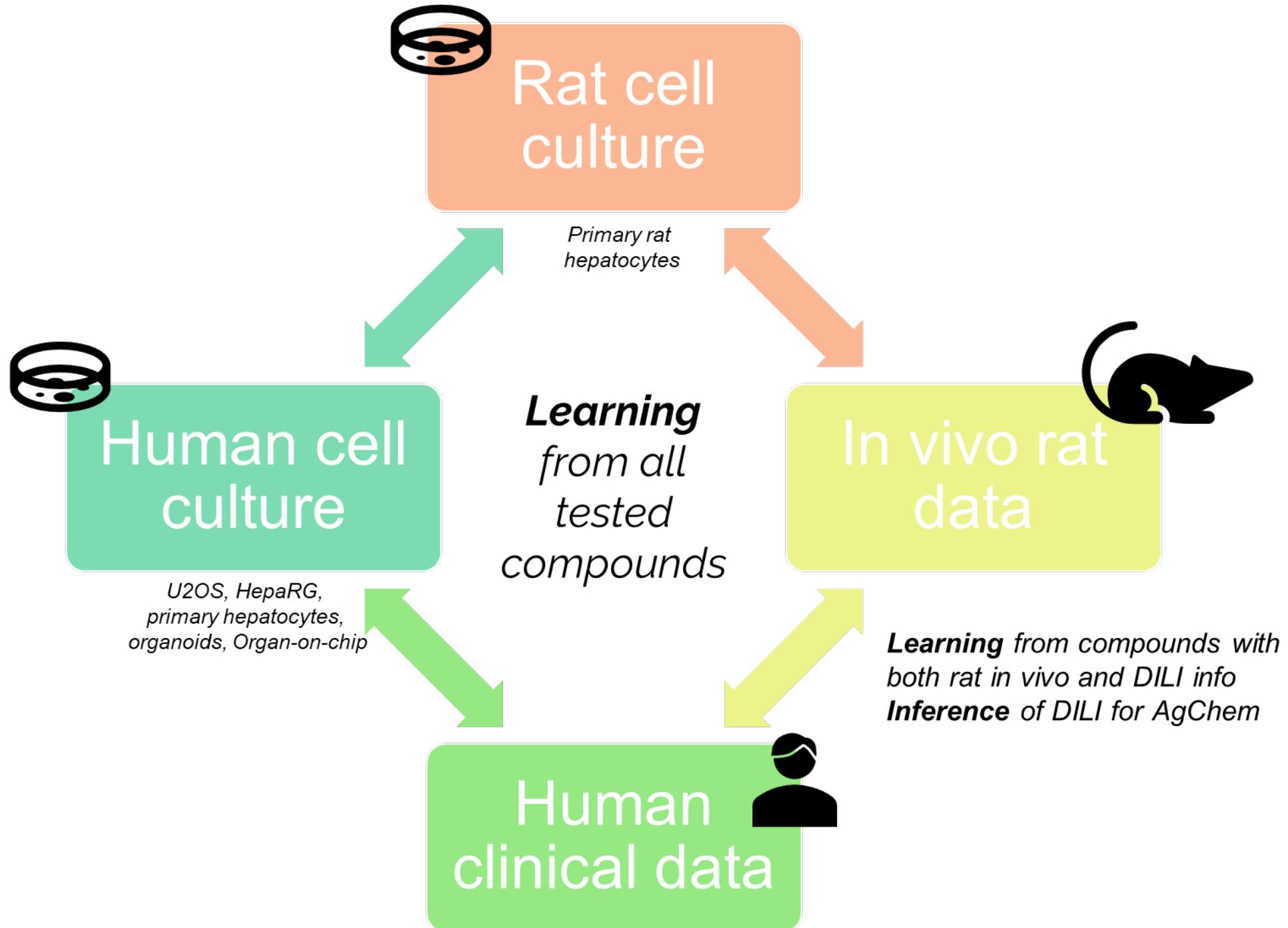
**Mission:** Gain confidence in the combination of Cell Painting, transcriptomics and proteomics for safety assessment using hepatotoxicity as use-case.

---

**Objective:** Benchmark *in vivo* hepatotoxicity (from rodent or clinical trial data) induced by a series of compounds against informatically aligned molecular & phenotypic cell-based assays.







Discovery

Pre-Development

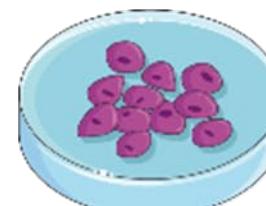
Development

## New Approach Methodologies (NAMs)

In Silico



In Vitro and  
Aquatic



In Vivo



# Acknowledgements

**Corteva**

**Kamin Johnson**

**Wei Chen**

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Eduardo Costa

Zachary Sutake

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Jiaqi Xu

Yang Li

Sagar Ksheera

Dillon Aberasturi

**US EPA**

Joshua Harrill

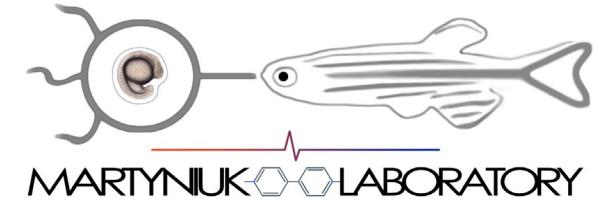
Jo Nyffeler

**Broad Institute**

Anne Carpenter

Shantanu Singh

**HESI OASIS Steering Team**



# Knowledge gaps and opportunities for omics data in environmental assessments

**Chris Martyniuk, Ph.D.**

Department of Physiological Sciences

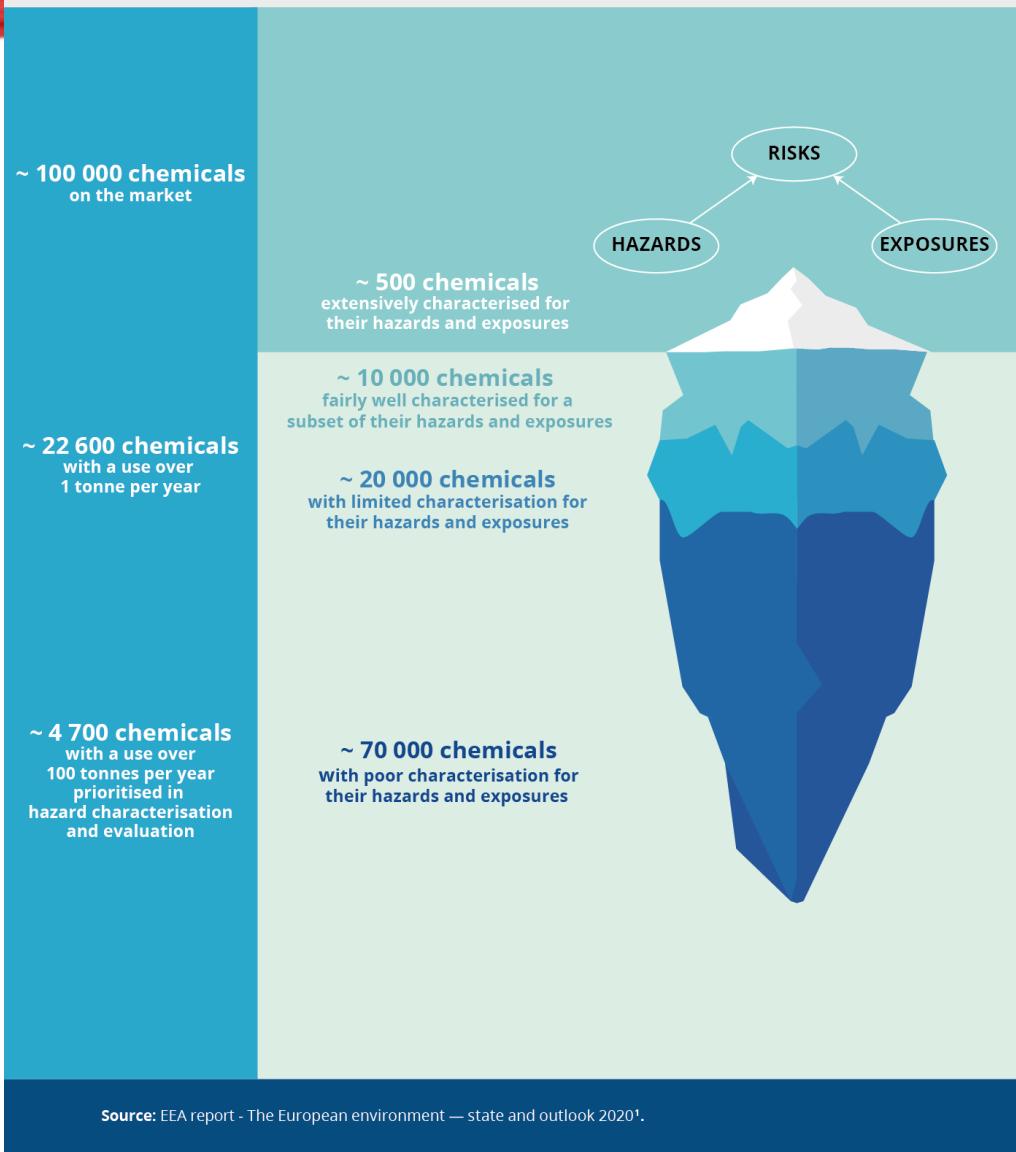
College of Veterinary Medicine

EPA NAMs workshop, November 5-6, 2024

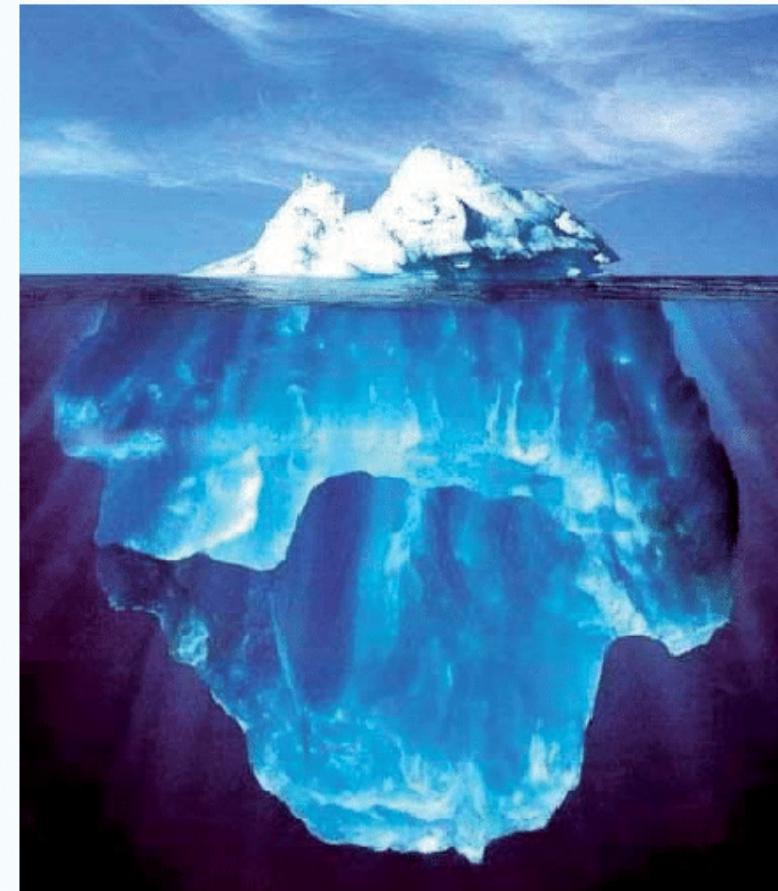


## The unknown territory of chemical risks

There are many chemicals on the market and only a small fraction of these have been extensively studied for their risks. Designing safe products with a smaller number of different chemicals is one way of reducing potential risks.

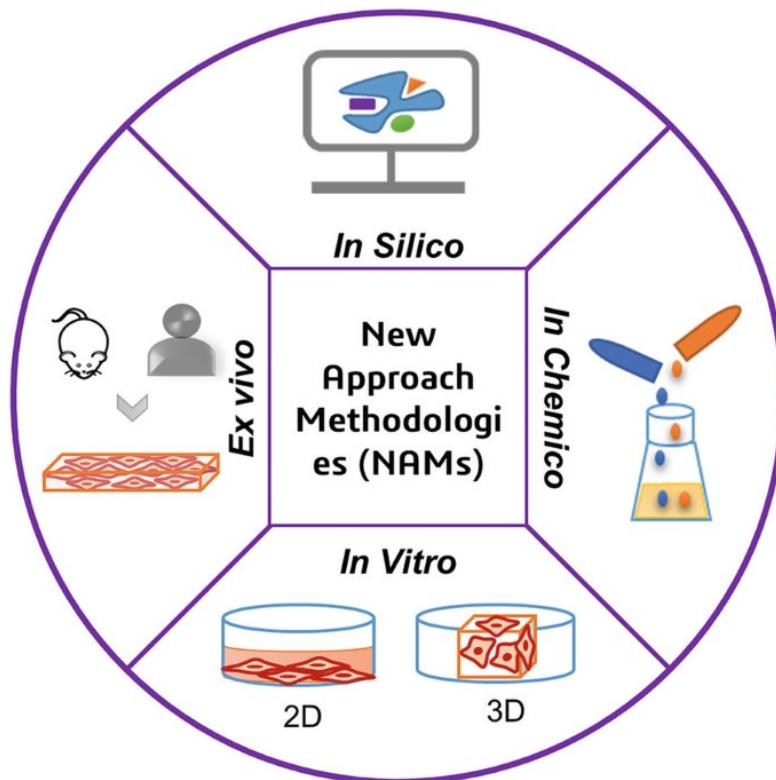


# THE CHALLENGE FOR REGULATORY AGENCIES



# Why important?

## Risk Evaluations for Existing Chemicals



## Prioritization

Which one do we test?



## Evaluation

How do we measure the toxicity?



## Management

Which ones do we regulate?

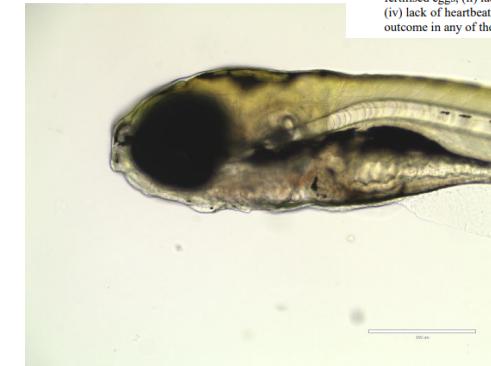
When: \_\_\_\_\_  
22 & 23 November

## Current Status of New Approach Methodologies (NAMs) for Regulatory Purposes

Online  
[Lhasalimited.org/events](http://Lhasalimited.org/events)

# THE ZEBRAFISH MODEL FOR HIGH THROUGHPUT TOXICITY TESTING

- Relatively cheap to house and readily accessible
- Rapid development with transparent developmental stages
- Replication: High numbers of individuals can be examined at once
- Amendable to mechanistic studies with chemicals of concern (AOPs)



Nan Wu photo credit

OECD/OCDE

236

Adopted:  
26 July 2013

### OECD GUIDELINES FOR THE TESTING OF CHEMICALS

#### Fish Embryo Acute Toxicity (FET) Test

##### INTRODUCTION

1. This Test Guideline (TG) 236 describes a Fish Embryo Acute Toxicity (FET) test with the zebrafish (*Danio rerio*). This test is designed to determine acute toxicity of chemicals on embryonic stages of fish. The FET-test is based on studies and validation activities performed on zebrafish (1)(2)(3)(4)(5)(6)(7)(8)(9)(10)(11)(12)(13)(14). The FET-test has been successfully applied to a wide range of substances exhibiting diverse modes of action, solubilities, volatilities, and hydrophobicities (reviewed in 15 and 16).

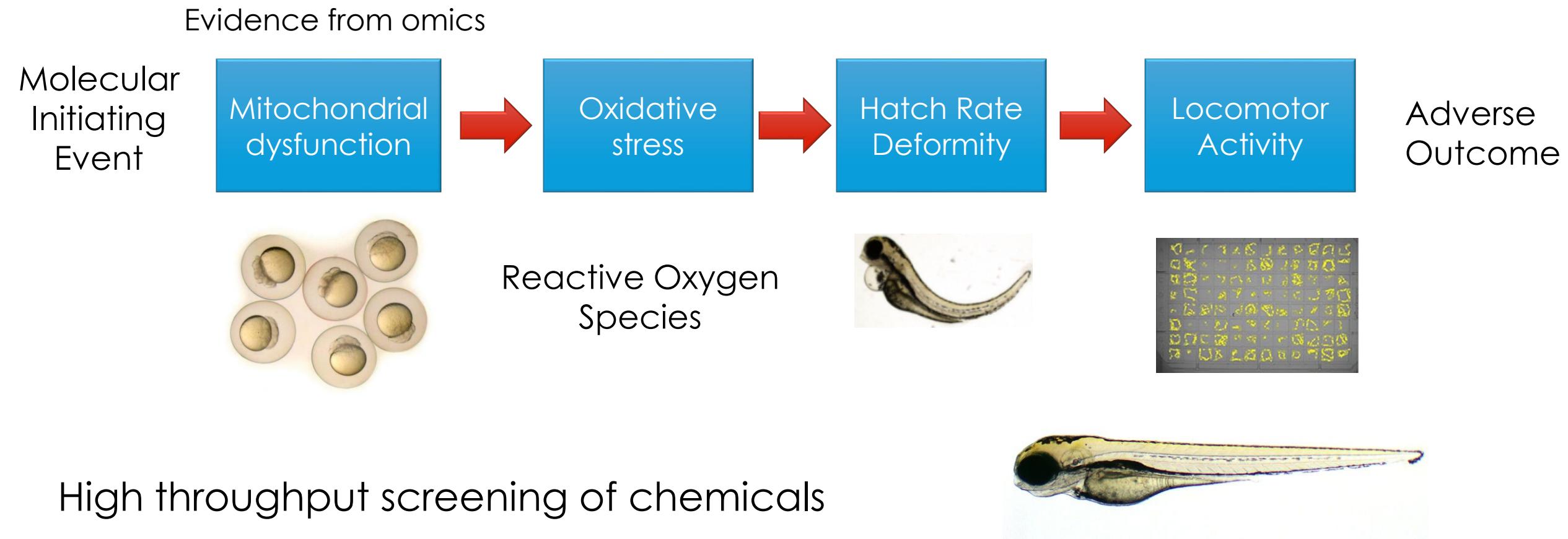
2. Definitions used in this Test Guideline are given in Annex 1.

##### PRINCIPLE OF THE TEST

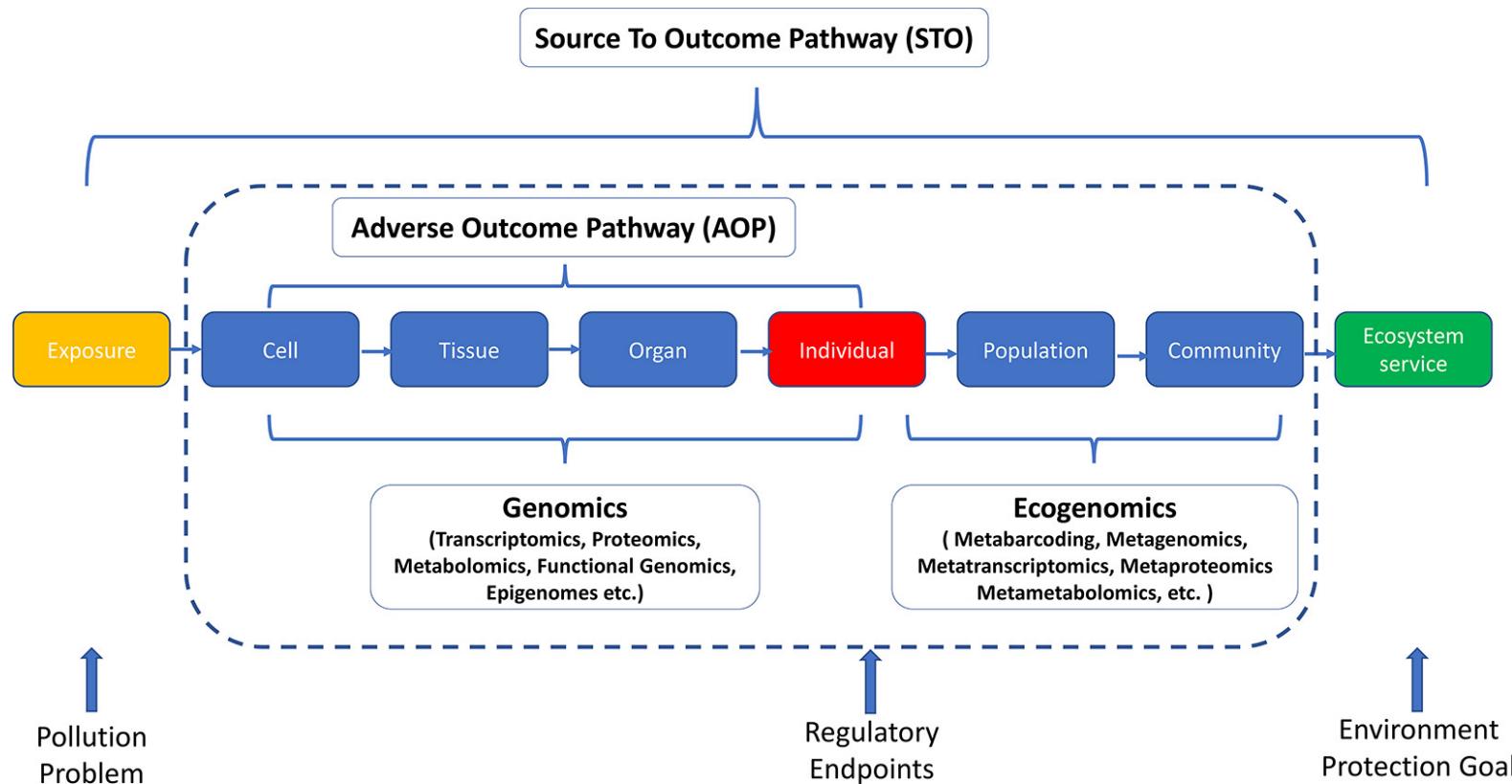
3. Newly fertilised zebrafish eggs are exposed to the test chemical for a period of 96 hrs. Every 24 hrs, up to four apical observations are recorded as indicators of lethality (6): (i) coagulation of fertilised eggs, (ii) lack of somite formation, (iii) lack of detachment of the tail-bud from the yolk sac, and (iv) lack of heartbeat. At the end of the exposure period, acute toxicity is determined based on a positive outcome in any of the four apical observations recorded, and the  $LC_{50}$  is calculated.



# Long term efforts: Defining AOPs for contaminants of concern



# Opportunities for **omics** in Risk Characterization and Assessment?



TOXICOLOGICAL SCIENCES, 158(2), 2017, 252–262

doi: 10.1093/toxsci/kfx097  
Advance Access Publication Date: May 19, 2017  
Forum

## FORUM

### The Role of Omics in the Application of Adverse Outcome Pathways for Chemical Risk Assessment

Erica K. Brockmeier,<sup>\*</sup> Geoff Hodges,<sup>†,1</sup> Thomas H. Hutchinson,<sup>‡</sup> Emma Butler,<sup>†</sup> Markus Hecker,<sup>§</sup> Knut Erik Tollefsen,<sup>¶</sup> Natalia Garcia-Reyero,<sup>¶,||</sup> Peter Kille,<sup>|||</sup> Dörthe Becker,<sup>#</sup> Kevin Chipman,<sup>#</sup> John Colbourne,<sup>#</sup> Timothy W. Collette,<sup>\*\*</sup> Andrew Cossins,<sup>\*</sup> Mark Cronin,<sup>††</sup> Peter Graystock,<sup>¶</sup> Steve Gutsell,<sup>†</sup> Dries Knapen,<sup>¶</sup> Ioanna Katsiadaki,<sup>¶</sup> Anke Lange,<sup>¶</sup> Stuart Marshall,<sup>†</sup> Stewart F. Owen,<sup>¶</sup> Edward J. Perkins,<sup>¶</sup> Stewart Plaistow,<sup>\*</sup> Anthony Schroeder,<sup>¶</sup> Daisy Taylor,<sup>¶</sup> Mark Viant,<sup>#</sup> Gerald Ankley,<sup>¶</sup> and Francesco Falciani<sup>\*</sup>

# Prioritization

Which one do we test?



# Evaluation

How do we measure the toxicity?

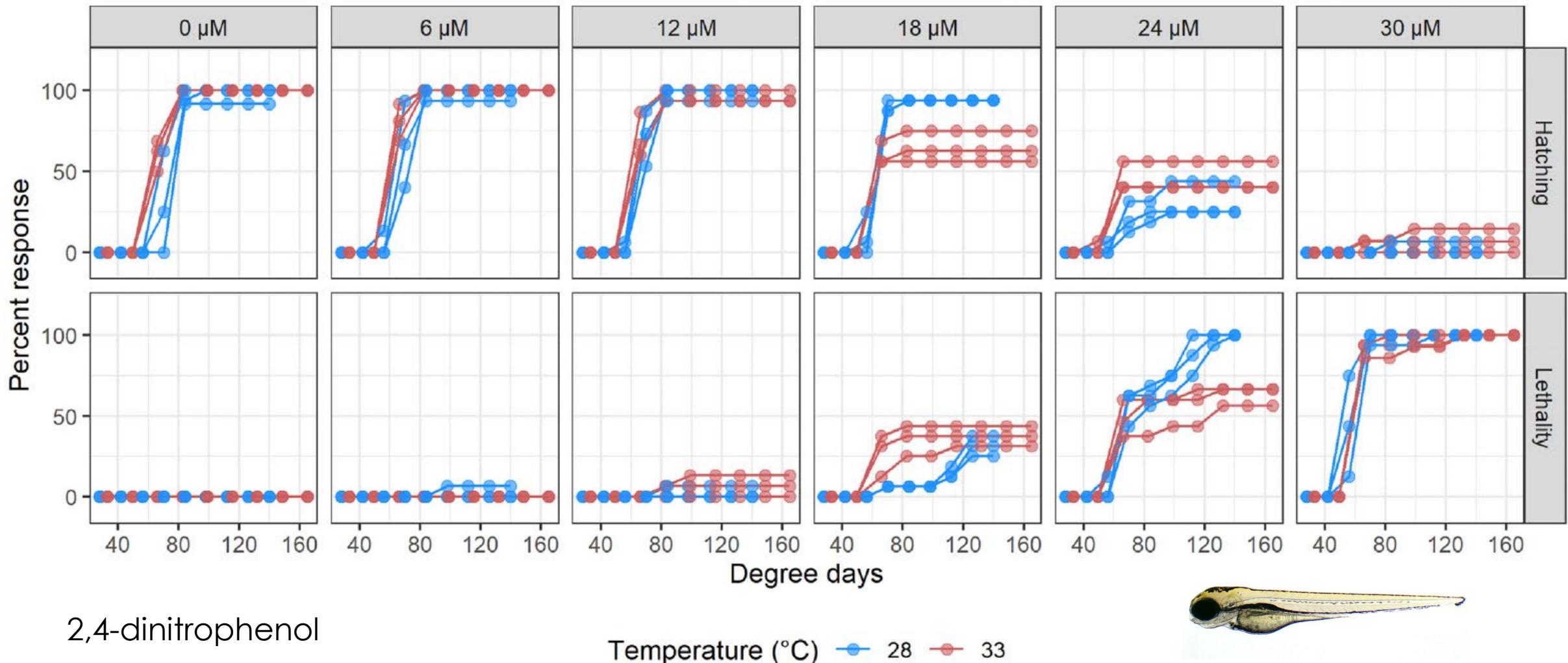


# Management

Which ones do we regulate?

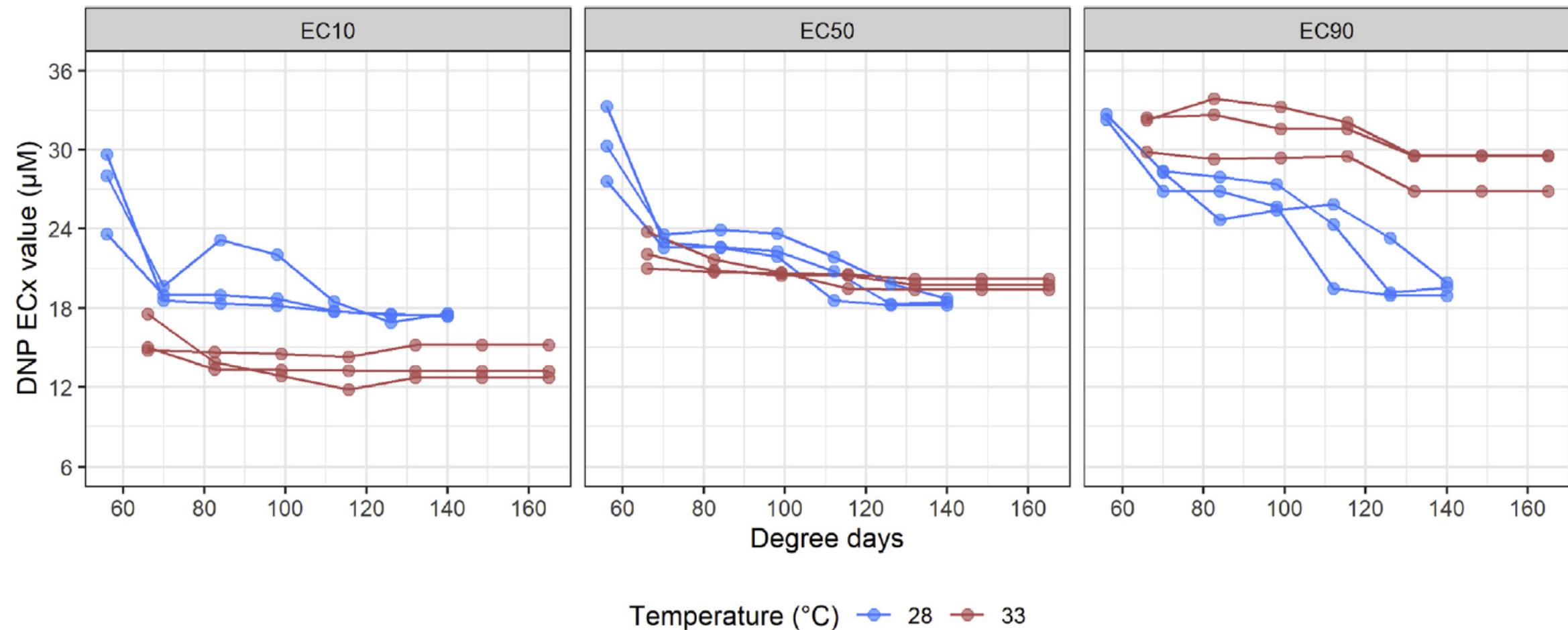


# Temperature, omics, and a mitochondrial toxicant

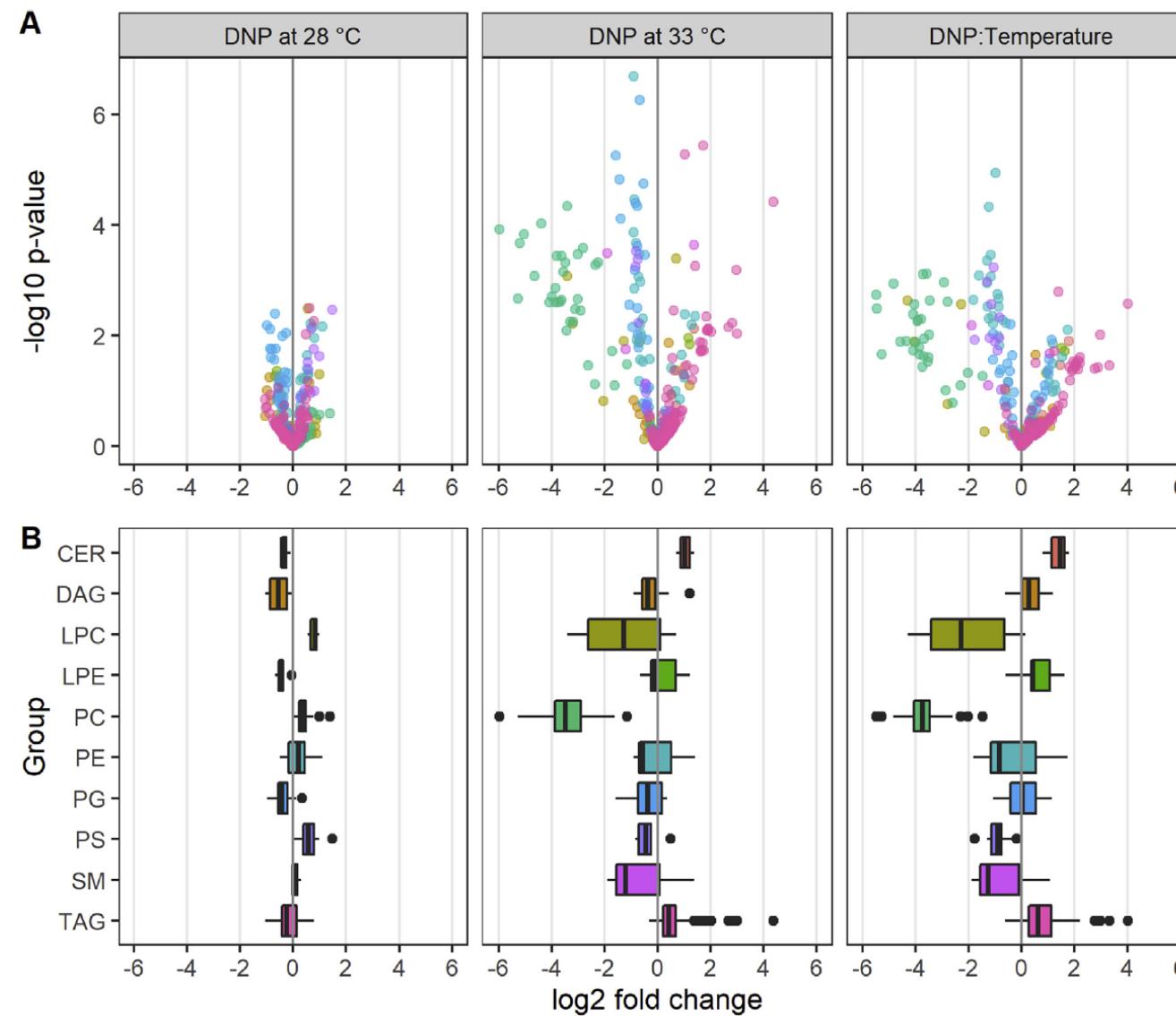


**Fig. 1.** Percent hatching and lethality as a function of exposure duration (degree days) for each clutch, temperature, and 2,4-Dinitrophenol concentration.

Dreier DA, Nouri MZ, Denslow ND, Martyniuk CJ. Lipidomics reveals multiple stressor effects (temperature  $\times$  mitochondrial toxicant) in the zebrafish embryo toxicity test. *Chemosphere*. 2021 Feb;264(Pt 1):128472. doi: 10.1016/j.chemosphere.2020.128472. Epub 2020 Sep 30. PMID: 33039916.



**Fig. 2.** 2,4-Dinitrophenol (DNP) EC<sub>x</sub> values as a function of exposure duration (degree days) for each clutch (connected points) and temperature.

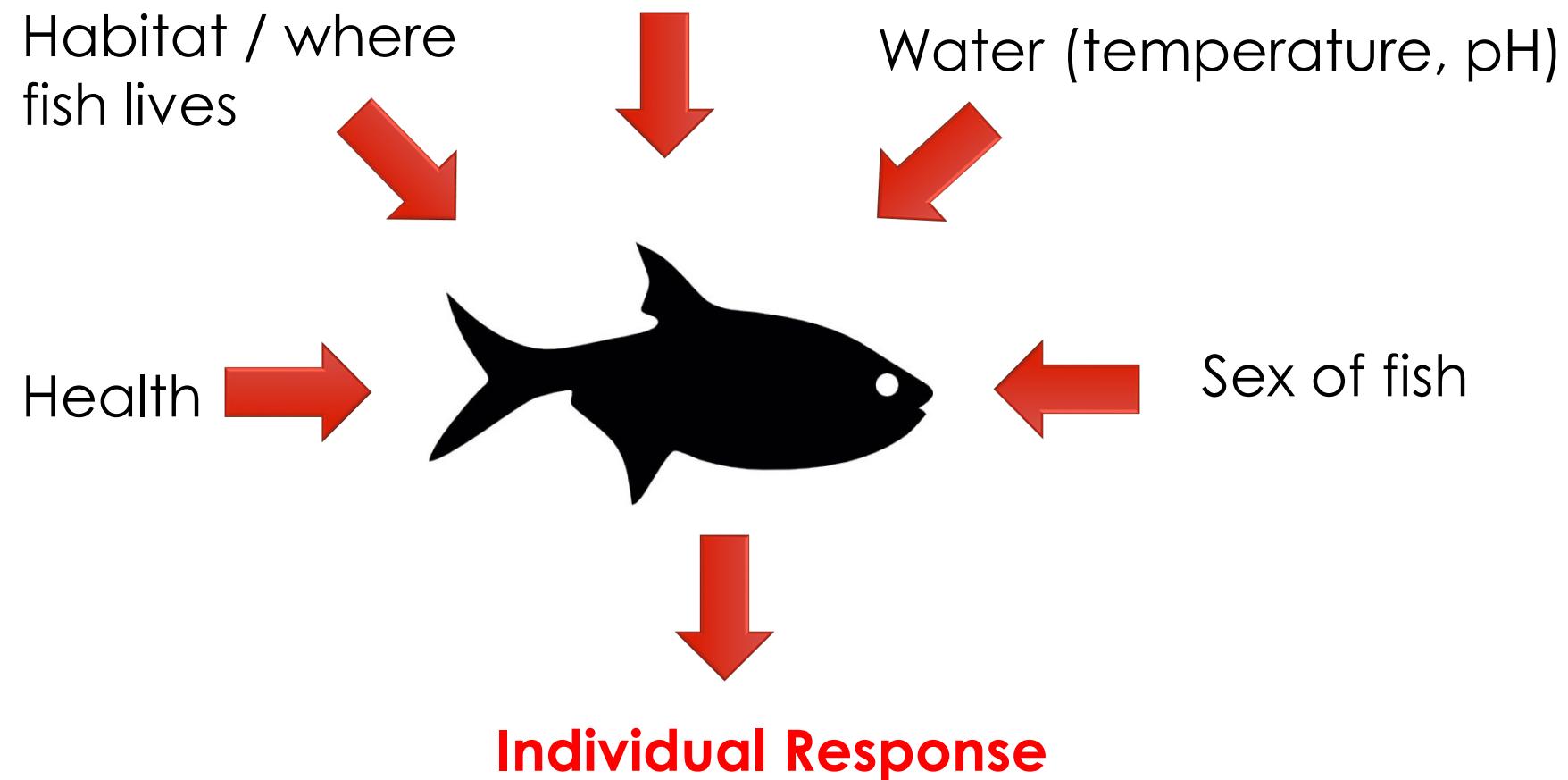


Lipidomics data set

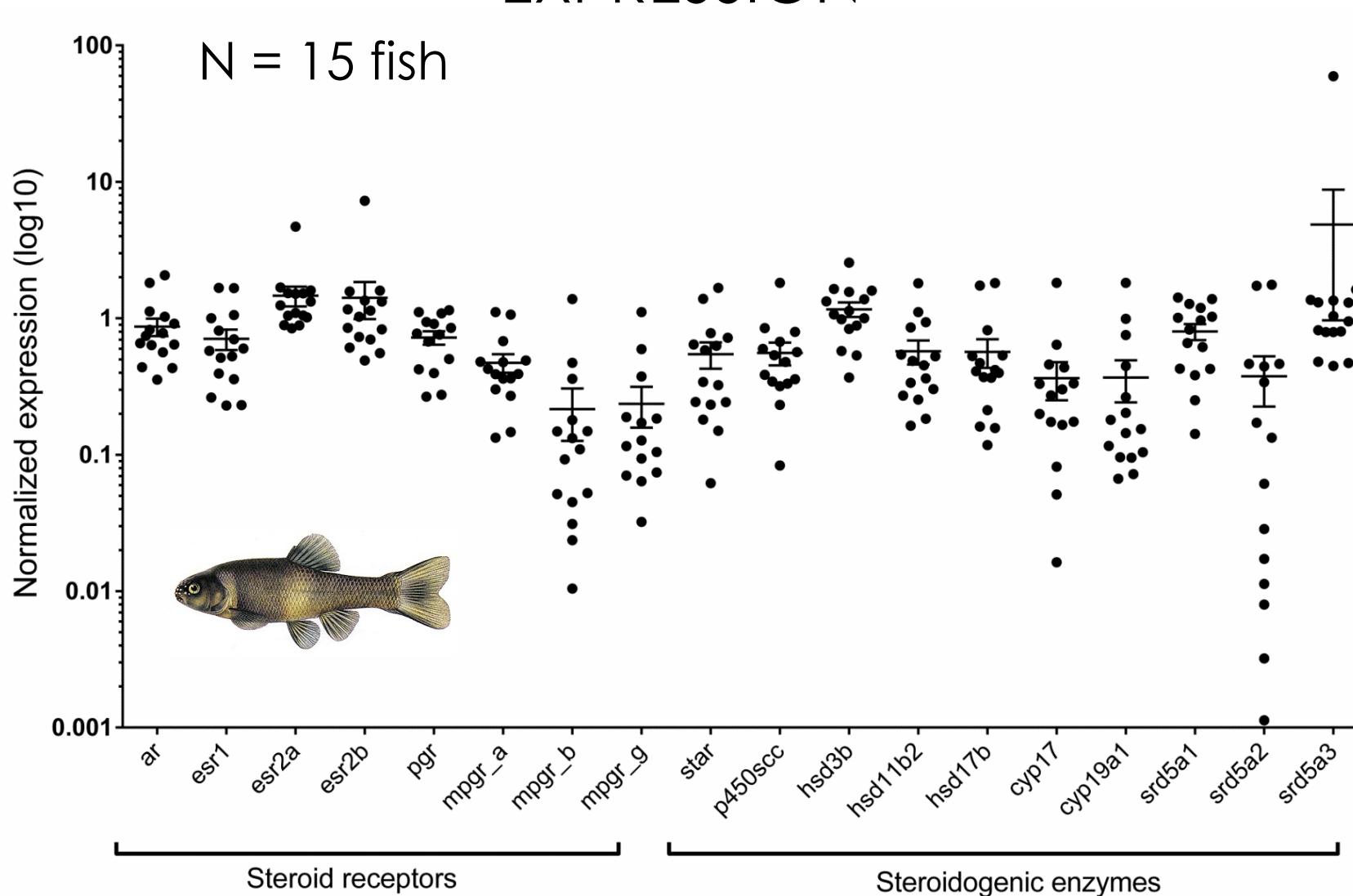
**Fig. 4.** Differential lipid composition for linear contrasts of interest (DNP at 28 °C, DNP at 33 °C, DNP:Tempearture) visualized as a (A) volcano plot and (B) boxplot.

# THE CHALLENGE MOVING INTO THE ENVIRONMENT...

## Ecological Interactions



# INDIVIDUAL VARIABILITY IN GENE EXPRESSION



Largest watershed in southern Ontario, Canada

This system receives discharges from **30 Sewage Plants**

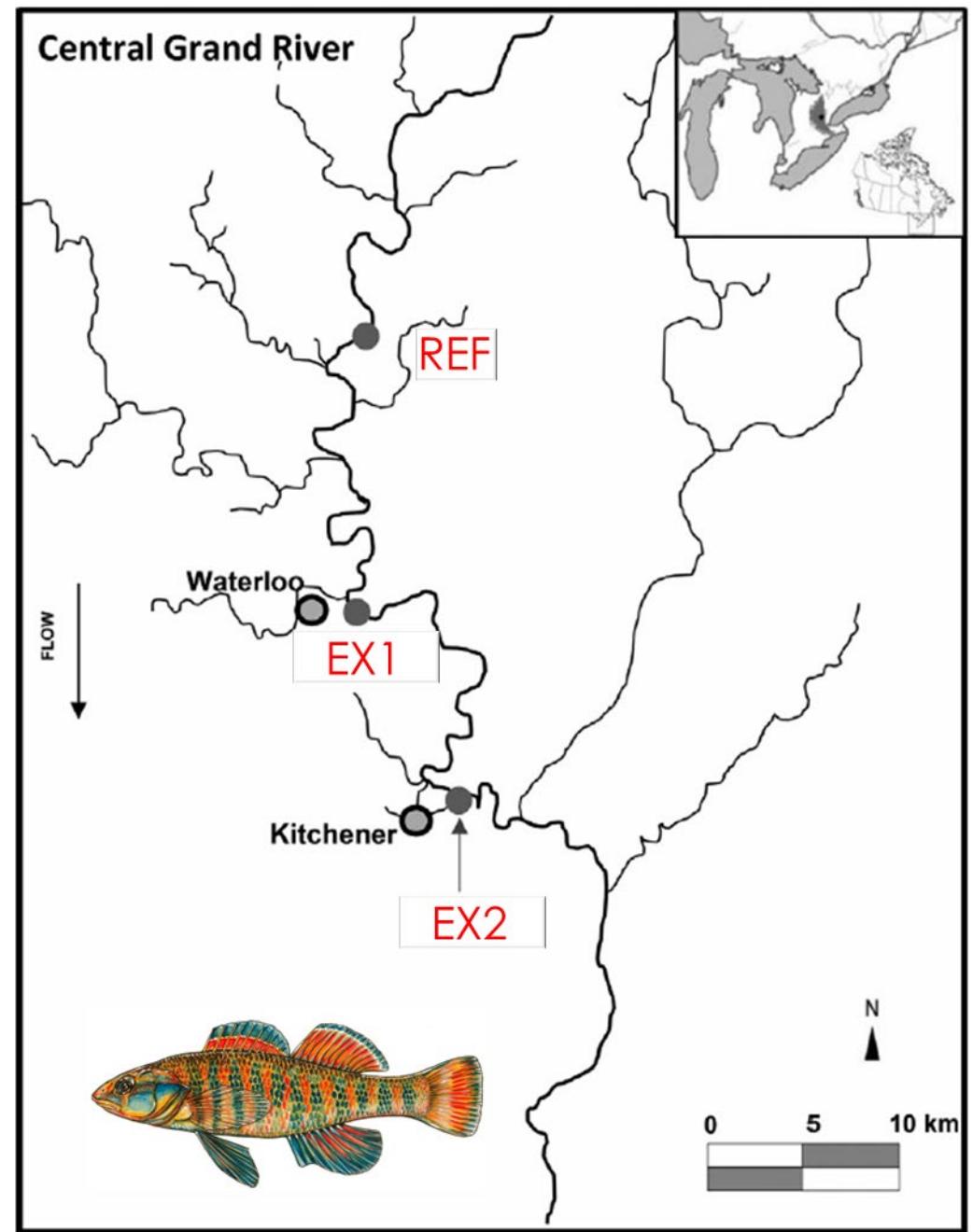
Upgrades at the Kitchener Plant

Ref = upstream

EX1 = downstream Waterloo

EX2 = downstream Kitchener

Marjan P, Bragg LM, MacLatchy DL, Servos MR, Martyniuk CJ. How Does Reference Site Selection Influence Interpretation of Omics Data?: Evaluating Liver Transcriptome Responses in Male Rainbow Darter (*Etheostoma caeruleum*) across an Urban Environment. *Environ Sci Technol*. 2017 Jun 6;51(11):6470-6479. doi: 10.1021/acs.est.7b00894. Epub 2017 May 17. PMID: 28489360.

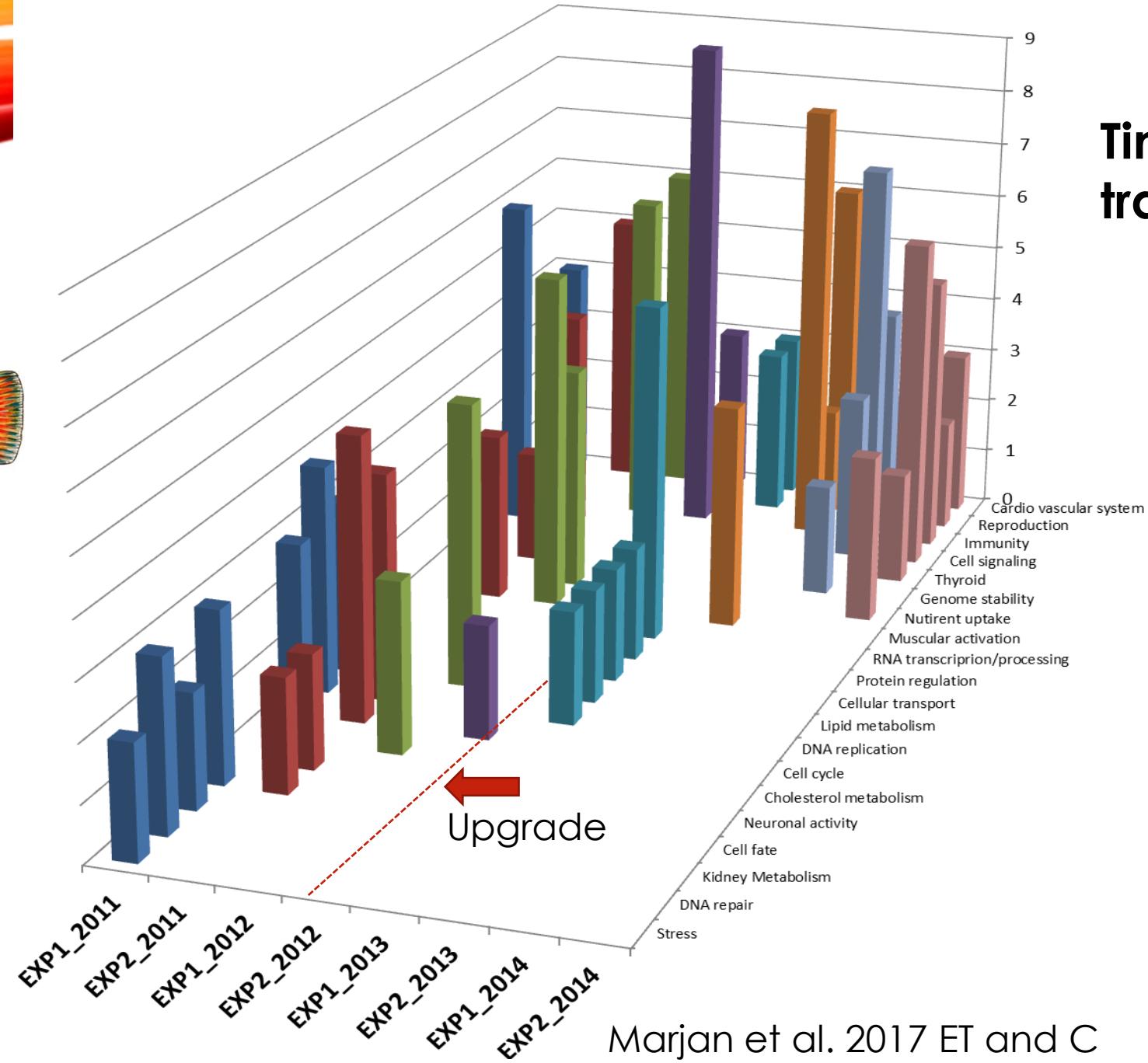




## Cell Fate

## DNA Repair

## Stress



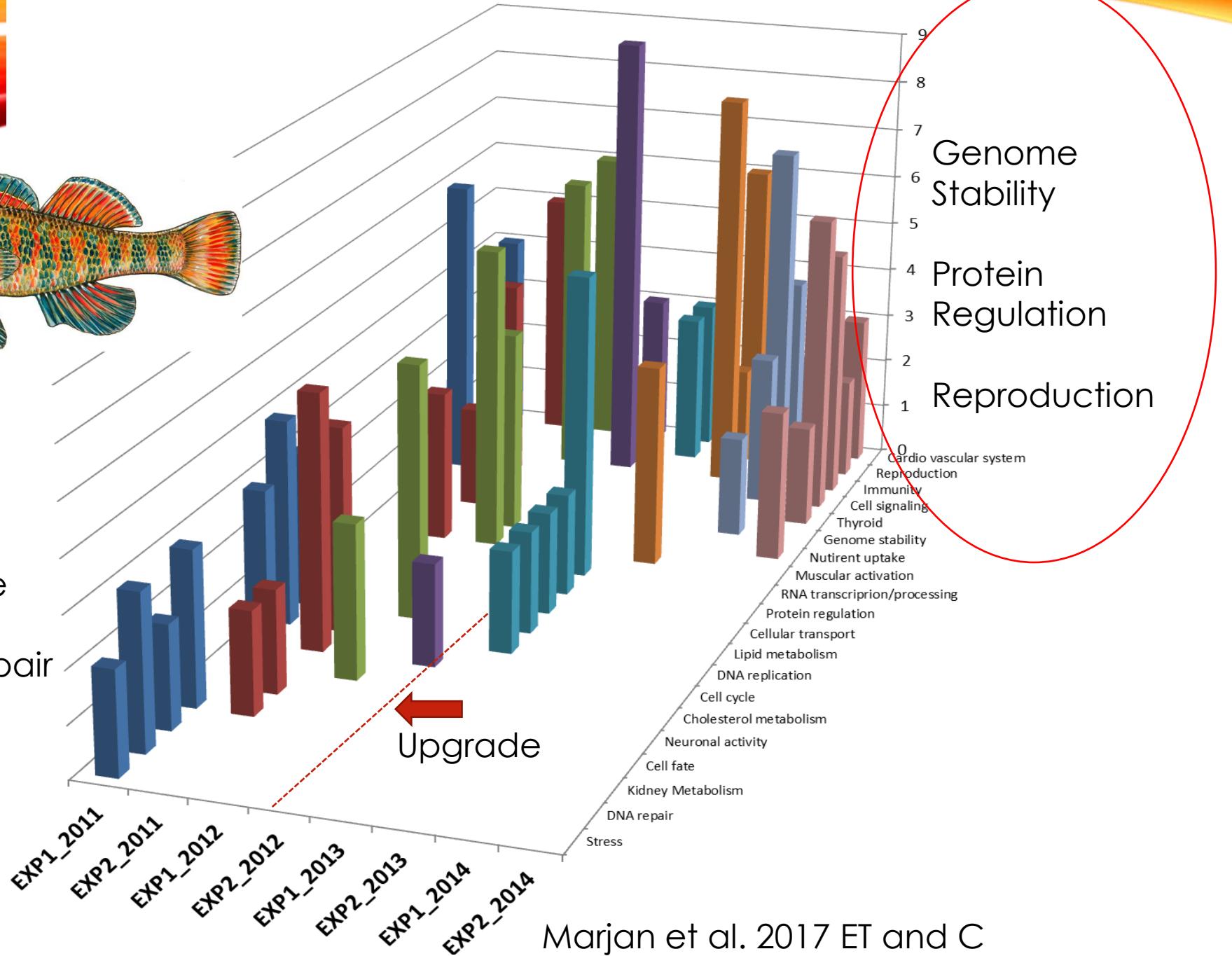
# Time and transcriptomes

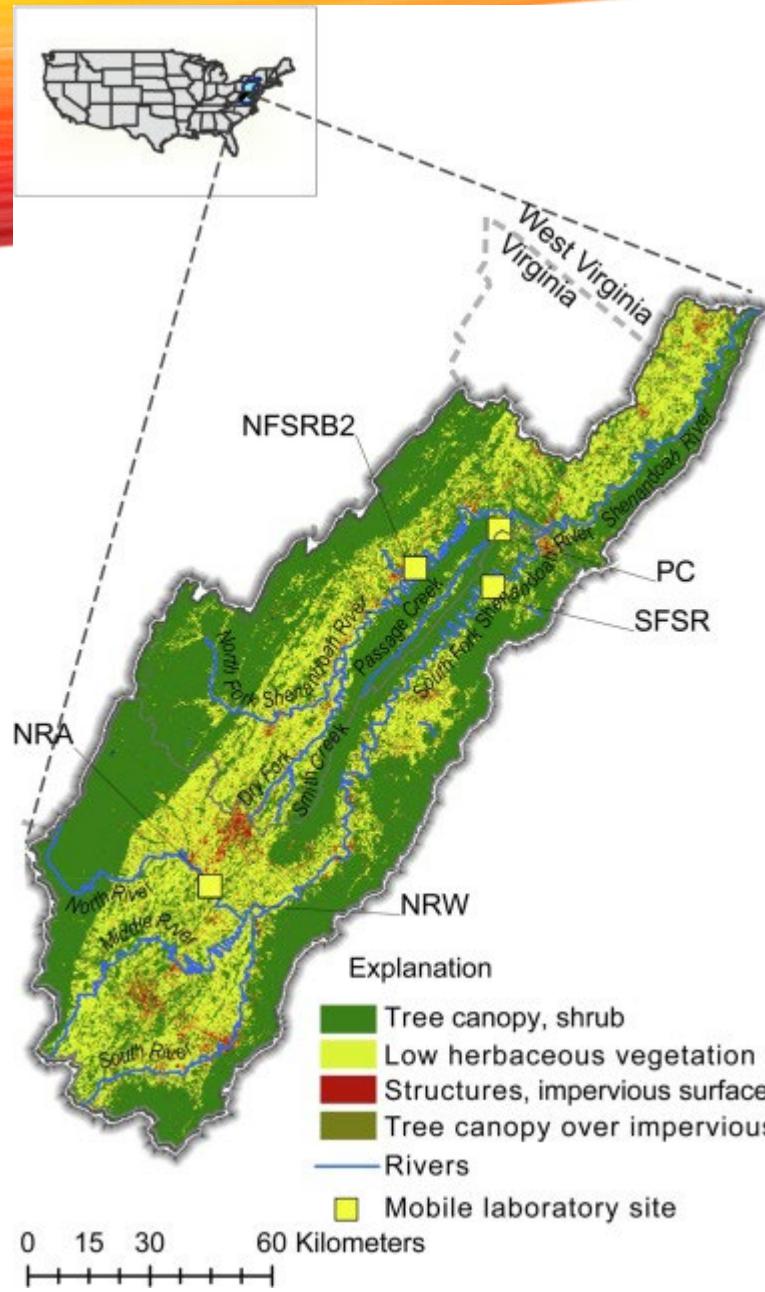
Marjan et al. 2017 ET and C

# Time and transcriptomes



Cell Fate  
DNA Repair  
Stress





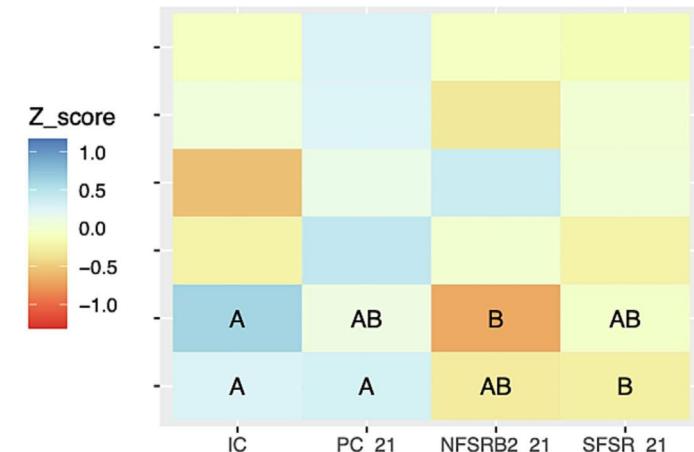
0 15 30 60 Kilometers

Bertolatus DW, Barber LB, Martyniuk CJ, Zhen H, Collette TW, Ekman DR, Jastrow A, Rapp JL, Vajda AM. Multi-omic responses of fish exposed to complex chemical mixtures in the Shenandoah River watershed. *Sci Total Environ.* 2023 Dec 1;902:165975. doi: 10.1016/j.scitotenv.2023.165975. Epub 2023 Aug 1. PMID: 37536598; PMCID: PMC10592118.

A) 2014 Male

Condition factor	AB	BC	A	AB	C
GSI	A	AB	B	B	B
Nuptial tubercles	A	B	AC	C	AB
Sperm abundance	A	AB	AB	B	AB
Plasma Vtg	AB	A	AB	B	AB
vtg mRNA	A	AB	AB	B	AB
	IC	PC_21	NRA_21	NRW_21	SFSR_21

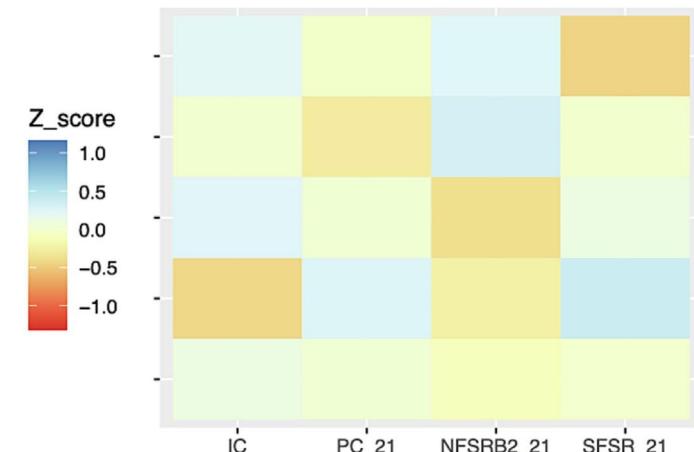
B) 2015 Male

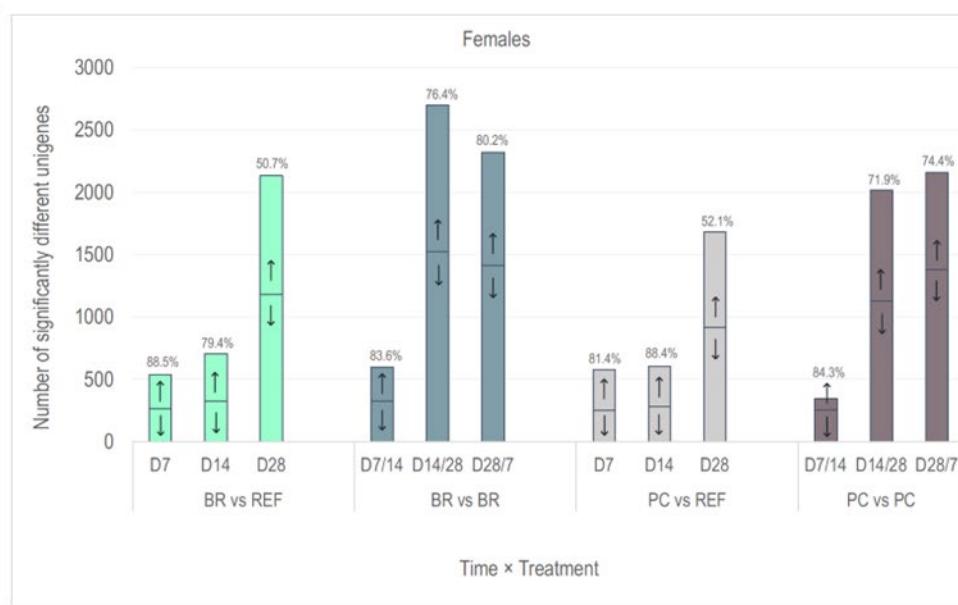
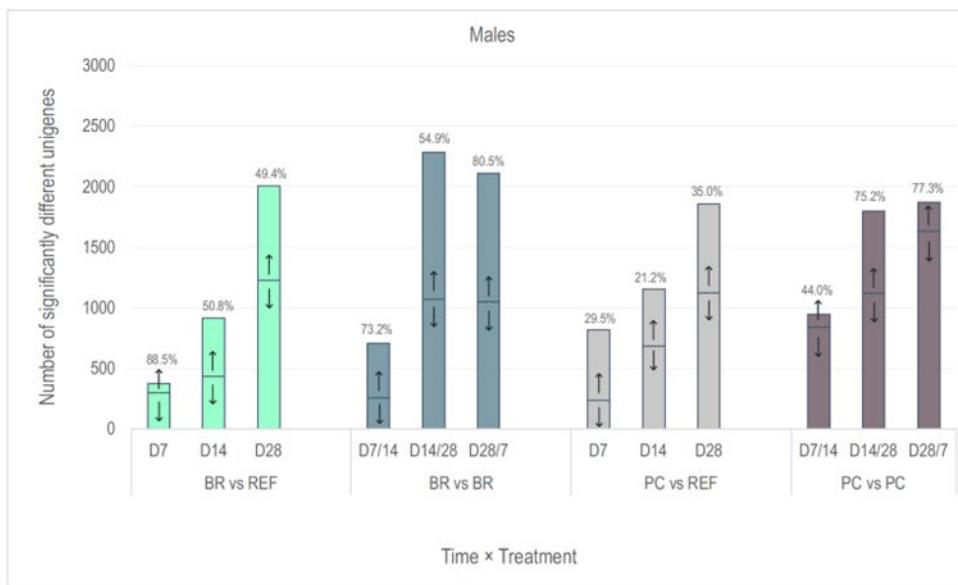


C) 2014 Female

Condition factor	A	AB	AB	B	A
GSI	A	A	B	B	A
Ovarian stage	AB	A	AB	AB	B
Plasma Vtg	A	B	A	AB	B
vtg mRNA	A	A	B	C	A
	IC	PC_21	NRA_21	NRW_21	SFSR_21

D) 2015 Female





*“While there are several putative biomarkers identified in hubs related to gene sets, temporal responses in the hepatic transcriptome made it challenging to elucidate definitive response patterns that could be used in field-based ecotoxicogenomic studies on the impacts of well-treated MWWE.”*

DEFINING THE ROLE OF OMICS IN ASSESSING ECOSYSTEM HEALTH:  
PERSPECTIVES FROM THE CANADIAN ENVIRONMENTAL MONITORING PROGRAM

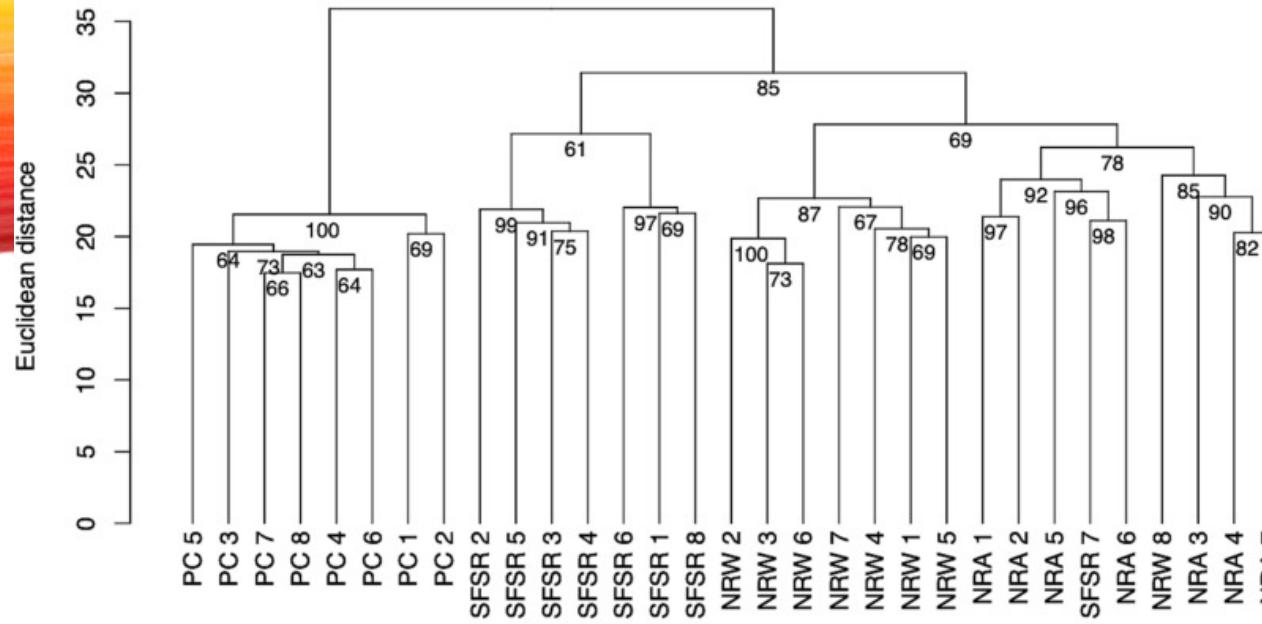
PAULINA A. BAHAMONDE, APRIL FESWICK, MEGHAN A. ISAACS, KELLY R. MUNKITTRICK, and  
CHRISTOPHER J. MARTYNIUK\*

Canadian Rivers Institute and Department of Biology, University of New Brunswick, Saint John, New Brunswick, Canada

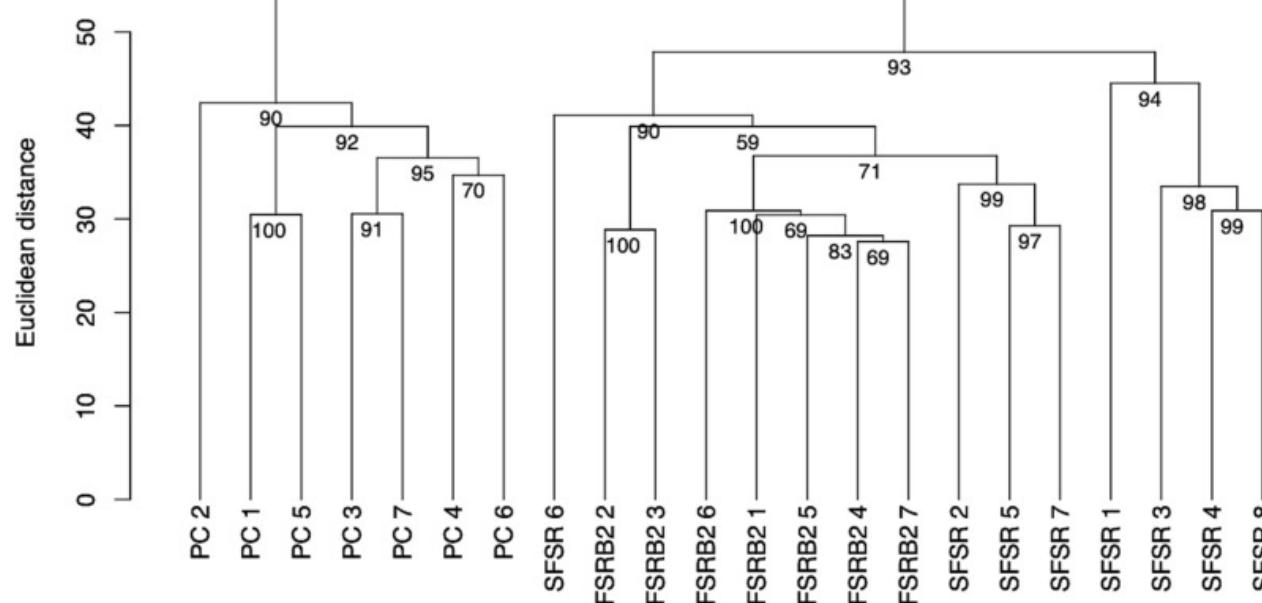
*(Submitted 28 March 2015; Returned for Revision 16 May 2015; Accepted 20 August 2015)*

- DEFINING NORMAL: AN IMPORTANT EEM CONSIDERATION
- Sampling period and temporal variability
- Spatial scale variation in field studies
- Understanding magnitude of abiotic, non-chemical factors
- Physiological responses

A) 2014



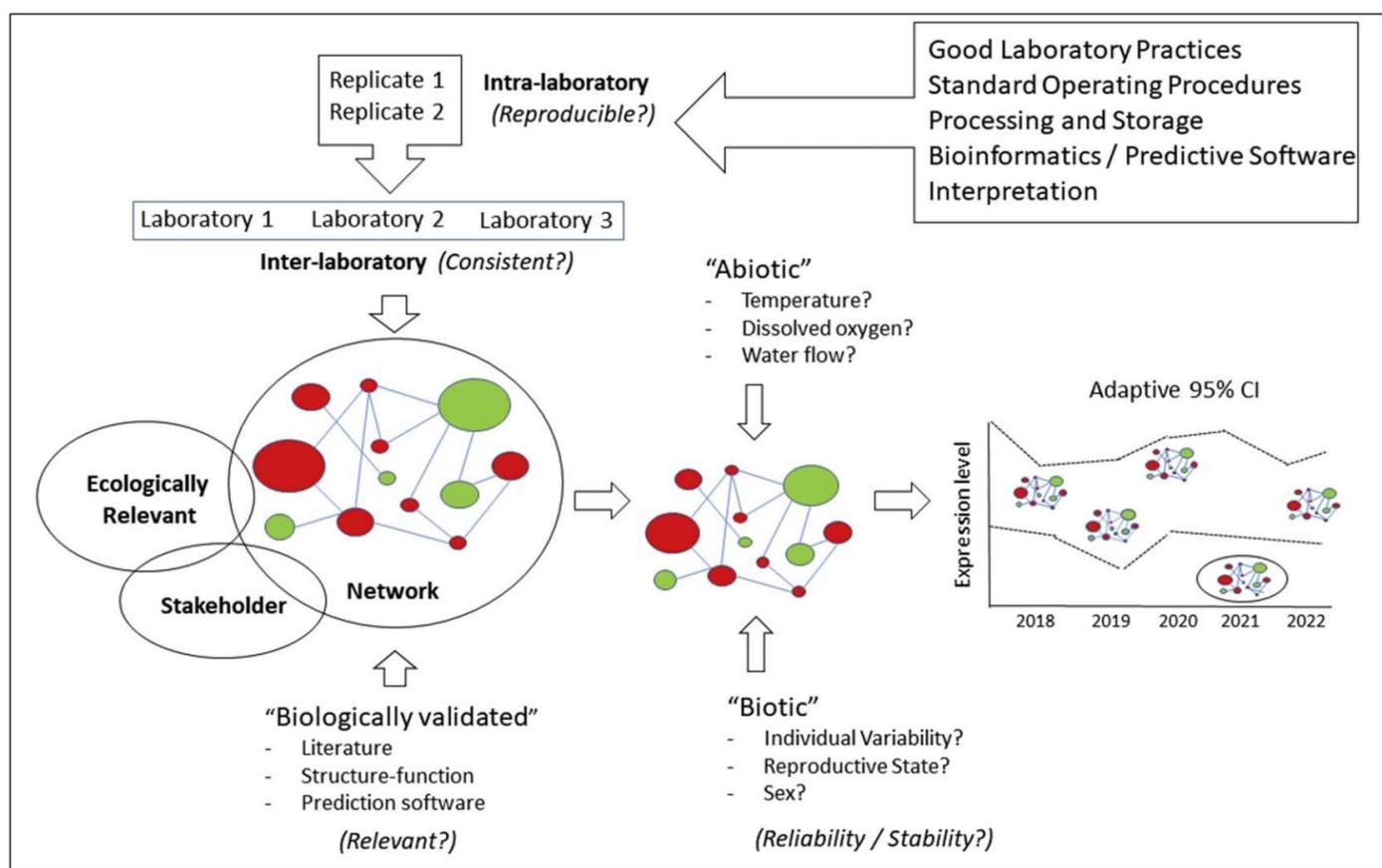
B) 2015



Omics: Good at predicting individual habit

But....

Relative response due to chemical vs. other variables?



Key points to be addressed prior to implication of omics technologies into a monitoring program.

#### Key Points

- Baseline data for omics responses in ecologically relevant species to be assessed.
- Data to identify the range of what is biologically normal within a given system to be monitored.
- Develop a level of standardization, consistency, and rigor that will allow interpretation of the relevance of response across broader scales.
- Leverage the AOP framework to bridge the gap between molecular responses and apical endpoints relevant for environmental monitoring.
- Discussion on what constitutes a meaningful change in a molecular network.

# Prioritization

Which one do we test?



# Evaluation

How do we measure the toxicity?



# Management

Which ones do we regulate?

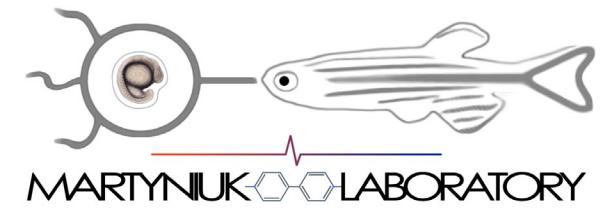


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Kelly R. Munkittrick, U Calgary

David Bertolatus, Adams State  
Alan Vajda, U. Colorado Denver



**UF** | Physiological Sciences  
College of Veterinary Medicine



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FLORIDA  
College of  
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Health  
Canada

Santé  
Canada

Canada

# **Fostering transparency and reproducibility using the OECD Omics Reporting Framework**

Presented at the 4<sup>th</sup> U.S. Environmental Protection Agency NAMs Conference:

State of Science on Development and Use of NAMs for Chemical Safety Testing (November 5-6, 2024)

**Matthew J. Meier**, Research Scientist, Health Canada

# Contributors to the OORF

- **Members of EAGMST, ESCA, and Omics Expert Group for valuable feedback**
- **Case study partners**
- **Tim Gant (Univ. of Leicester)**
- **Carole Yauk (Univ. of Ottawa)**
- **Joshua Harrill (US EPA)**
- **Mark Viant (Univ. of Birmingham)**
- **Ksenia Groh (Eawag)**
- **Matthew Meier (Health Canada)**
- **Magda Sachana (OECD secretariat)**
- Scott S. Auerbach
- Richard D. Beger
- Mounir Bouhifd
- Jason O'Brien
- Lyle Burgoon
- Florian Caiment
- Donatella Carpi
- Tao Chen
- Brian N. Chorley
- John Colbourne
- Raffaella Corvi
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- Alberto Martini
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- Serge Rudazyz
- Prem Kumarathasan
- Alexandra Schaffert
- Oliver Schmitz
- Tomasz Sobanski
- Volker Strauss
- Russell S. Thomas
- Monica Vaccaria
- Vikrant Vijay
- Ralf J.M. Weber
- Antony J. Williams
- Andrew Williams
- Maurice Whelan
- David Crizer
- Tom Lawson
- N. Cabaton
- M. Trembley-Franco
- C. Canlet
- T. Schock
- K. Hagiwara
- Leah Wehmas
- Sarah Davidson
- Alison Harrill
- Beena Vallanat
- Nil Basu/Doug Crump

# Evaluating thousands of chemicals for potential toxicity: easier said than done

- Disadvantages of historically used single-endpoint tests:

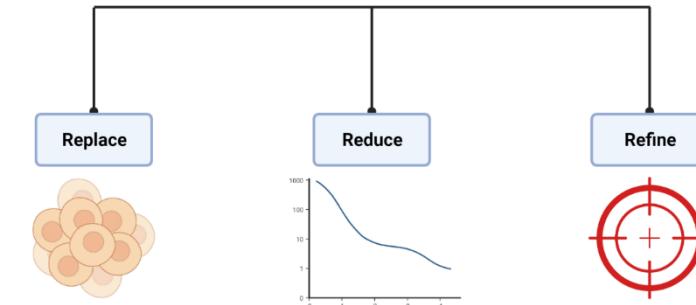
- Slow
- Expensive
- Use animals



- New Approach Methods (NAMs)
  - Regulatory agencies worldwide are reducing reliance on tests in animals (EU and cosmetics, 2013; US EPA; Canada has legislated changes in CEPA)
- Omics plays a critical role in NAM-based tests
  - Compatibility with non-animal tests: e.g., human cell lines, micro-physiological systems – increasingly complex *in vitro* assays



## The "3 Rs" of animal testing



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français

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MENU ▾

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> [Consultation: Draft strategy to replace, reduce or refine vertebrate animal testing](#)

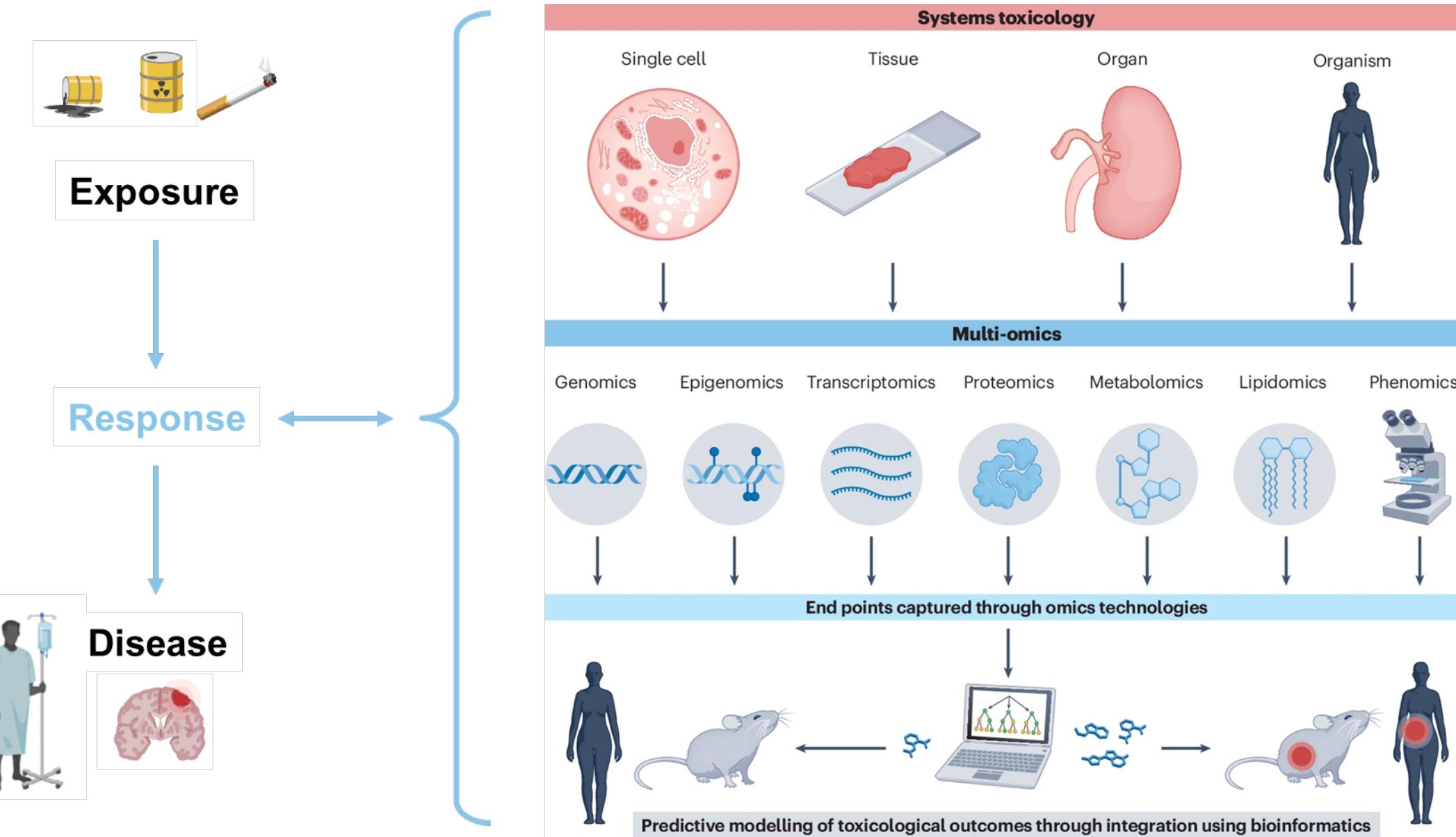
## Draft Strategy to Replace, Reduce or Refine Vertebrate Animal Testing under the Canadian Environmental Protection Act, 1999

Environment and Climate Change Canada

Health Canada

September 2024

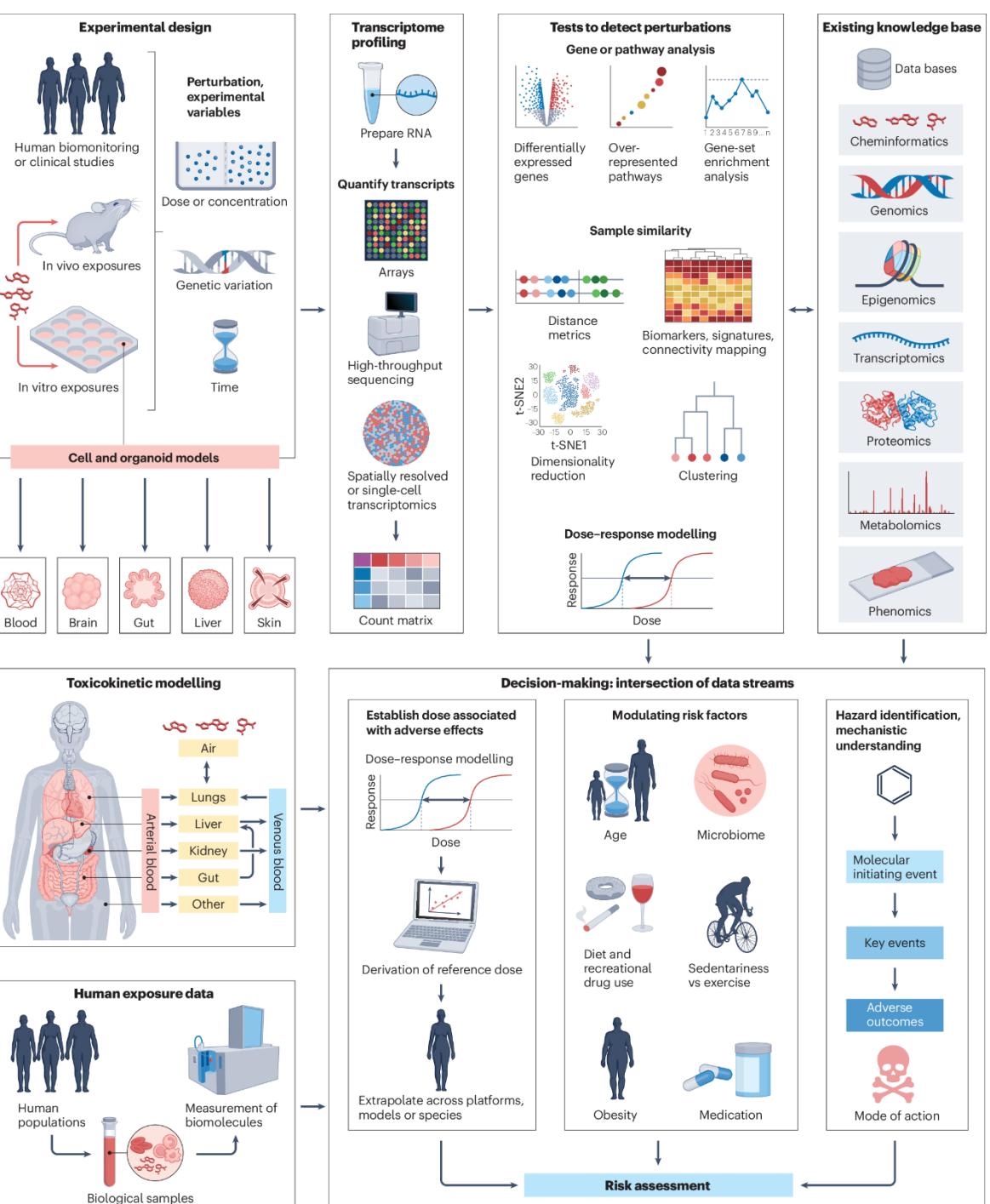
# Using omics to explore biological responses



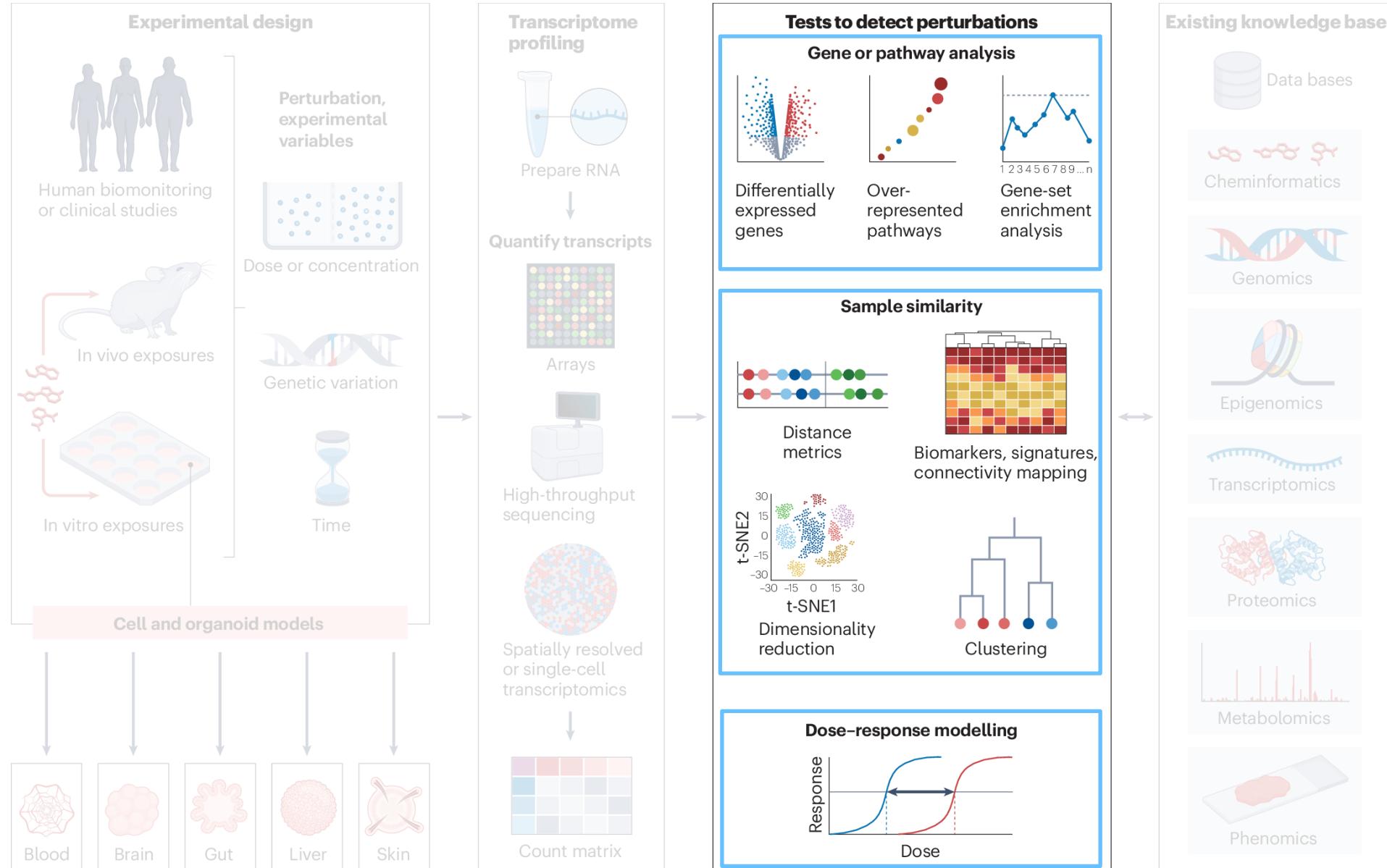
Meier, M.J., Harrill, J., Johnson, K., Thomas, R.S., Tong, W., Rager, J.E., Yauk, C.L. *Nat Rev Genet* (2024).

# The promise of toxicogenomics for human health protection

- Molecular alterations occur before changes in apical endpoints
- Omics provides significant advantages over traditional toxicology tests
  - Rapid & cost-effective data generation
  - Reduction in animal use

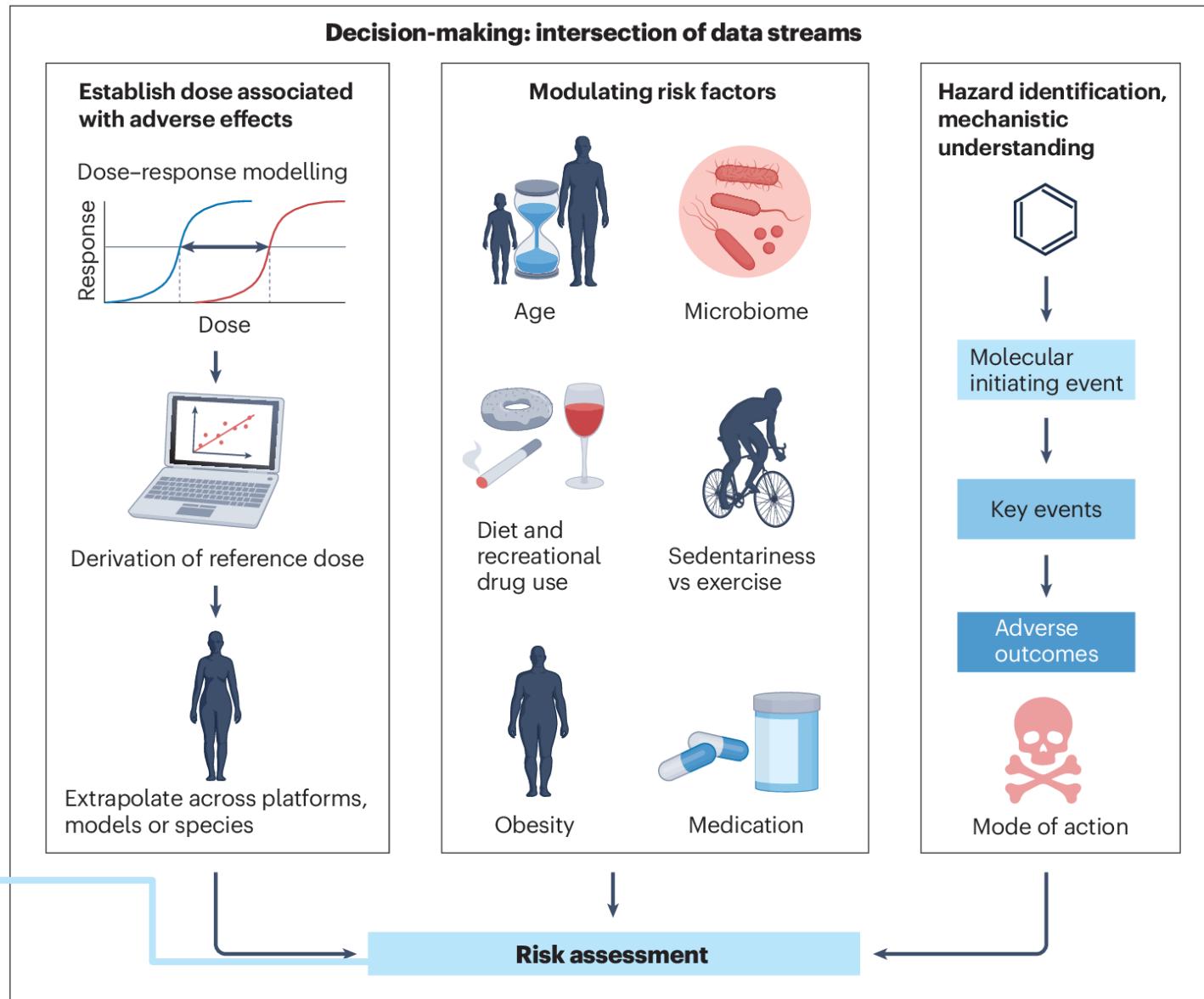


# The promise of toxicogenomics for human health protection



# Implementing toxicogenomics in decision-making

- Data streams converge on risk assessment



“Torture the data, and it will confess to anything.”

-Ronald Coase, British economist



# Challenges applying omics in risk assessment



Contents lists available at [ScienceDirect](#)

Regulatory Toxicology and Pharmacology

journal homepage: [www.elsevier.com/locate/yrtpb](http://www.elsevier.com/locate/yrtpb)



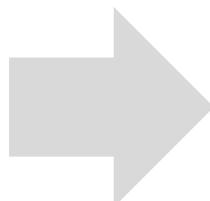
Commentary

Progress towards an OECD reporting framework for transcriptomics and metabolomics in regulatory toxicology

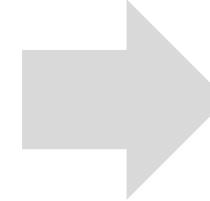


Joshua A. Harrill <sup>a,1,\*</sup>, Mark R. Viant <sup>b,c,\*,1</sup>, Carole L. Yauk <sup>d,1,\*\*\*</sup>, Magdalini Sachana <sup>e</sup>,  
Timothy W. Gant <sup>f</sup>, Scott S. Auerbach <sup>g</sup>, Richard D. Beger <sup>h</sup>, Mounir Bouhifd <sup>i</sup>, Jason O'Brien <sup>j</sup>,  
Lyle Burgoon <sup>k</sup>, Florian Caiment <sup>l</sup>, Donatella Carpi <sup>m</sup>, Tao Chen <sup>h</sup>, Brian N. Chorley <sup>a</sup>,  
John Colbourne <sup>b,c</sup>, Raffaella Corvi <sup>m</sup>, Laurent Debrauwer <sup>n,o</sup>, Claire O'Donovan <sup>p</sup>, Timothy M.  
D. Ebbels <sup>q</sup>, Drew R. Ekman <sup>r</sup>, Frank Faulhammer <sup>s</sup>, Laura Gribaldo <sup>m</sup>, Gina M. Hilton <sup>t</sup>,  
Stephanie P. Jones <sup>j</sup>, Aniko Kende <sup>u</sup>, Thomas N. Lawson <sup>c</sup>, Sofia B. Leite <sup>m</sup>, Pim E.G. Leonards <sup>v</sup>,  
Mirjam Luijten <sup>w</sup>, Alberto Martin <sup>i,2</sup>, Laura Moussa <sup>x</sup>, Serge Rudaz <sup>y,z,aa</sup>, Oliver Schmitz <sup>ab</sup>,  
Tomasz Sobanski <sup>i</sup>, Volker Strauss <sup>s</sup>, Monica Vaccari <sup>ac</sup>, Vikrant Vijay <sup>h</sup>, Ralf J.M. Weber <sup>b,c</sup>,  
Antony J. Williams <sup>a</sup>, Andrew Williams <sup>ad</sup>, Russell S. Thomas <sup>a</sup>, Maurice Whelan <sup>m</sup>

- Lack of transparency in data generation and processing
- Lack of standardisation in study parameters and reporting of results
- Lack of case studies and guidance describing acceptable (and ultimately best) practices



Impacts on  
experimental or  
analytical  
reproducibility



Reduced  
regulatory utility

# OECD Omics Reporting Framework (OORF)

ENV/CBC/MONO(2023)41 | 3

OECD Environment, Health and Safety Publications

SERIES ON TESTING AND ASSESSMENT

NO. 390



OECD Omics Reporting Framework (OORF): Guidance on reporting elements for the regulatory use of omics data from laboratory-based toxicology studies

- Recognition in international community that omics provides significant advantages over traditional tests
  - However - regulators don't want to drink from a fire hose
- Growing interest in regulatory applications of omics must be supported by guidelines and frameworks
- The OECD Omics Reporting Framework (OORF) is now publicly available for use (Nov. 2023)
  - Began under the Extended Advisory Group for Molecular Screening and Toxicogenomics (EAGMST), now the OORF and related projects are within the Omics Expert Group, reporting to the Advisory Group on Emerging Science in Chemicals Assessment (ESCA), under the Working Party on Hazard Assessment (WPHA)

# OECD Omics Reporting Framework (OORF)

## Objective

Develop a framework to standardize **reporting** of omics data generation and analyses, so regulators can **understand and interpret** omics studies.

## Why do we need the OORF?

To ensure that sufficient information is available to **enable an evaluation of the quality of the experimental data and interpretation, and support reproducibility**.

(The 5 Rs: Regulatory Ratification Requires Reproducible Research)

- ✗ **NOT to stipulate the methods of data analysis or interpretation**
- ✓ **Rather, provide guidance on reporting of information that fosters transparency and reproducibility**

# Modular structure of the OORF

**Study Summary Reporting Module (SSRM)**

**Toxicology Experiment Reporting Module (TERM)**

# Expanding the scope of the OORF

- Early OORF development focused on transcriptomics and metabolomics
- There is a need to expand and re-evaluate language to encompass all omics
- Ksenia Groh (Eawag) and Alexandra Schaffert (Medical University Innsbruck) are co-leading the integration of proteomics modules, and reviewing what components should be updated, creating new DAPRMs



## Development of Proteomics Modules for the OECD Omics Reporting Framework



Ksenia Groh<sup>1</sup>, Premkumari Kumarathasan<sup>2</sup>, Steve U. Ayobahan<sup>3</sup>, Davide Degli-Esposti<sup>4</sup>, Verónica I. Dumit<sup>5</sup>, Sebastian Eilebrecht<sup>3</sup>, Vera Engelbrecht<sup>6</sup>, Salvador Fernandez Arroyo<sup>7</sup>, Nico Jehmlich<sup>8</sup>, Predrag Kukic<sup>9</sup>, Isabel Karkossa<sup>8</sup>, Thibaut Leger<sup>10</sup>, Arnaud Salvador<sup>11</sup>, Johannes Schmidt<sup>12</sup>, Kristin Schubert<sup>8</sup>, Joshua Harrill<sup>13</sup>, Magdalini Sachana<sup>14</sup> and Alexandra Schaffert<sup>15</sup>

<sup>1</sup>Eawag, Swiss Federal Institute of Aquatic Science and Technology, 8600 Dübendorf, Switzerland; <sup>2</sup>Analytical Biochemistry and Proteomics Laboratory, Mechanistic Studies Division, Environmental Health Science and Research Bureau, Healthy Environments and Consumer Safety Branch, Health Canada, Canada; <sup>3</sup>Fraunhofer Institute for Molecular Biology and Applied Ecology IME, Department Ecotoxicogenomics, 57392 Schmallenberg, Germany; <sup>4</sup>INRAE (National Research Institute for Agriculture, Food and Environment), Centre de Lyon-Grenoble Auvergne-Rhône-Alpes, Unité de recherches RiverLy, Ecotox team, 69625 Villeurbanne Cedex, France; <sup>5</sup>Federal Institute for Risk Assessment (BfR), Berlin, Germany; <sup>6</sup>PETA Science Consortium International e.V.; <sup>7</sup>EURECAT, Center for Omics Sciences, Reus, Tarragona, Spain; <sup>8</sup>Helmholtz Centre for Environmental Research (UFZ), Leipzig, Germany; <sup>9</sup>Unilever, Safety and Environmental Assurance Centre, Computational and in vitro Toxicology, Colworth Science Park, Bedfordshire, UK; <sup>10</sup>ANSES – French Agency for Food, Environmental and Occupational Health & Safety, Fougères Laboratory, France; <sup>11</sup>University of Lyon, CNRS, Université Claude Bernard Lyon 1, Institute des Sciences Analytiques, UMR 5280, 5 rue de la Doua, F-69100 Villeurbanne, France; <sup>12</sup>Fraunhofer Institute for Cell Therapy and Immunology, Leipzig, Germany; <sup>13</sup>US EPA, Center for Computational Toxicology and Exposure (CCTE), RTP, NC27709, USA; <sup>14</sup>OECD Environment Health and Safety Division, Environment Directorate, Paris, France; <sup>15</sup>MUI Medical University Innsbruck, Regulatory Toxicology, Institute of Medical Biochemistry, Innsbruck, Austria.

# Reporting templates

## 1. Study Summary Reporting Module

Red = Recommended

Blue = Optional

1.1. Study Identifiers

1.1.1. Abstract

Recommended

**INSTRUCTIONS TO DATA PROVIDERS:** Use Column C for data entry. Text in *italics* are instructions (e.g. reporting tips) for the data submitter for individual reporting fields. Text in **bold italics** are

## 2. Toxicology Experiment Reporting Module

Red = Recommended  
Blue = Optional

REPORTING CATEGORY

## 3.2. Data Acquisition and Processing Reporting Module (DAPRM) for RNA Seq and Targeted RNA-Seq Data

**INSTRUCTIONS TO DATA PROVIDERS:** Use Column D for data entry. Text in *italics* are instructions (e.g. reporting tips) for the data submitter for individual reporting fields. Text in **bold italics** are instructions for the data submitter that are applicable to all reporting elements in a section. For additional clarification regarding a reporting field, please refer to the

**INSTRUCTIONS TO DATA PROVIDERS:** Use Column D for data entry. Text in *italics* are instructions (e.g. reporting tips) for the data submitter for individual reporting fields. Text in **bold italics** are instructions for the data submitter that are applicable to all reporting elements in a section. For additional clarification regarding a reporting field, please refer to the corresponding guidance document for this reporting template.

Toxicology Experiment Mo  
1.2. Study Rationale

## 4.1. Data Analysis Reporting Module (DARM) for Discovery of Differentially Abundant Molecules - Univariate Methods

**INSTRUCTIONS TO DATA PROVIDERS:** Use Column D for data entry. Text in *italics* are instructions (e.g. reporting tips) for the data submitter for individual reporting fields. Text in **bold italics** are instructions for the data submitter that are applicable to all reporting elements in a section. For additional clarification regarding a reporting field, please refer to the corresponding guidance document for this reporting template.

Red = Recommended  
Blue = Optional

REPORTING CATEGORY

3.2.1. Technology

REPORTING CATEGORY

REPORTING ELEMENT

RECOMMENDED /  
OPTIONAL

INPUT

4.1.1. Inputs

4.1.1.1. Data Input(s)  
4.1.1.2. Metadata Input(s)

Recommended  
Recommended

see "metadata" tab

4.1.2. Software Documentation

4.1.2.1. Software  
4.1.2.2. Operating System  
4.1.2.3. Additional Libraries used  
4.1.2.4. Software Availability

Recommended  
Recommended  
Recommended  
Optional

R version 4.0.3 (2020-10-10)  
Platform: x86\_64-pc-linux-gnu (64-bit) running under: Ubuntu 20.04.2 LTS  
<https://www.r-project.org/>, <https://www.bioconductor.org/>

4.1.3. Contrasts for Which Differentially Abundant Molecules were Identified

4.1.3.1. Contrasts

Recommended

See "contrasts" tab

4.1.4. Assay Experimental Design

4.1.4.1. Group Sizes  
4.1.4.2. Covariance  
4.1.4.3. Technical Replicates

Recommended  
Recommended  
Recommended

See "metadata" tab  
Samples were processed in plate-based batches. See "metadata" tab.  
Technical replicates were used as different wells for library building. They derive from different aliquots.

4.1.5. Statistical Analysis to Identify Differentially Abundant Molecules

4.1.5.1. Statistical Approach  
4.1.5.2. Data Transformation  
4.1.5.3. Effects Models  
4.1.5.4. Modeling Inputs  
4.1.5.5. Bayesian Approaches  
4.1.5.6. Decision Criteria  
4.1.5.7. Other

Recommended  
Recommended  
Recommended  
Recommended  
Recommended  
Recommended  
Recommended

DESeq2  
Size factor normalization  
Negative Binomial General Linear Model (GLM) fitting performed using default settings in DESeq2  
contrast. Batches were used as covariates in the formula.  
Wald statistics calculated using DESeq2 with default settings  
NA  
R-ODAF criteria were used as the basis.  
multiple testing correction performed  
- Adjusted p-value cutoff = 0.05; MTC = FDR in DESeq2; note: Cook's cutoff was disabled  
- 1.5-fold linear fold change cutoff  
- Filters applied simultaneously following DESeq results tabular output

2.2 Test and Control items

SSRN

SSRM

SSRM

1.2.1. Background Information	Recommended	Concerns over exposure to Bisphenol A (BPA) and human health, particularly during early life stages implementation of regulatory actions towards use of chemical for commercial applications. This action is in a rise in the use of BPA alternatives in recent years due to the potential for regrettable substitution in consumer products considered "BPA-free". This work uses transcriptomic data, in supporting the development of an Integrated Approaches to Testing and Assessment (IATA), to a better understanding of the mechanism of action of BPA.
1.2.2. Objectives	Recommended	Concentration response experiments using TempO (BioSpyder Technologies) were conducted in order to evaluate 16 bisphenols relative to bisphenol A in MCF7-cell line. The objective was to explore potencies across the bisphenols using benchmark concentration analysis, and similarities in altered pathways/upstream regulators, pathway analysis and transcriptomic biomarkers.
1.2.3. Test item		

SSRM

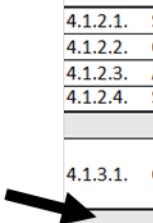


TERM



2.1.5. Model selection
a. For <i>in vivo</i> studies, describe why the selected animal species was chosen
b. Rationale for the species and strain used
interest
2.1.6. Dose / concentration level and interval selection
2.1.7. Route of administration
2.1.8. Time point selection
2.1.9. Samples and Replicates
a. Biological replicate number
b. Number of technical and analytical replicates
2.1.10. Limitations
2.2.1. Test item name

DARM



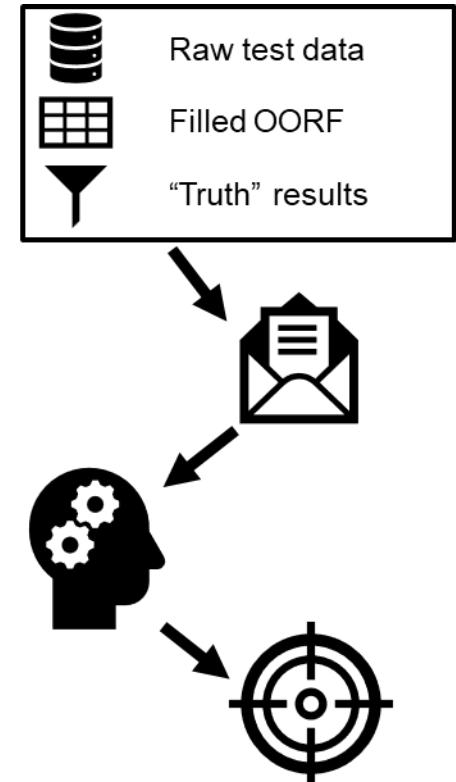
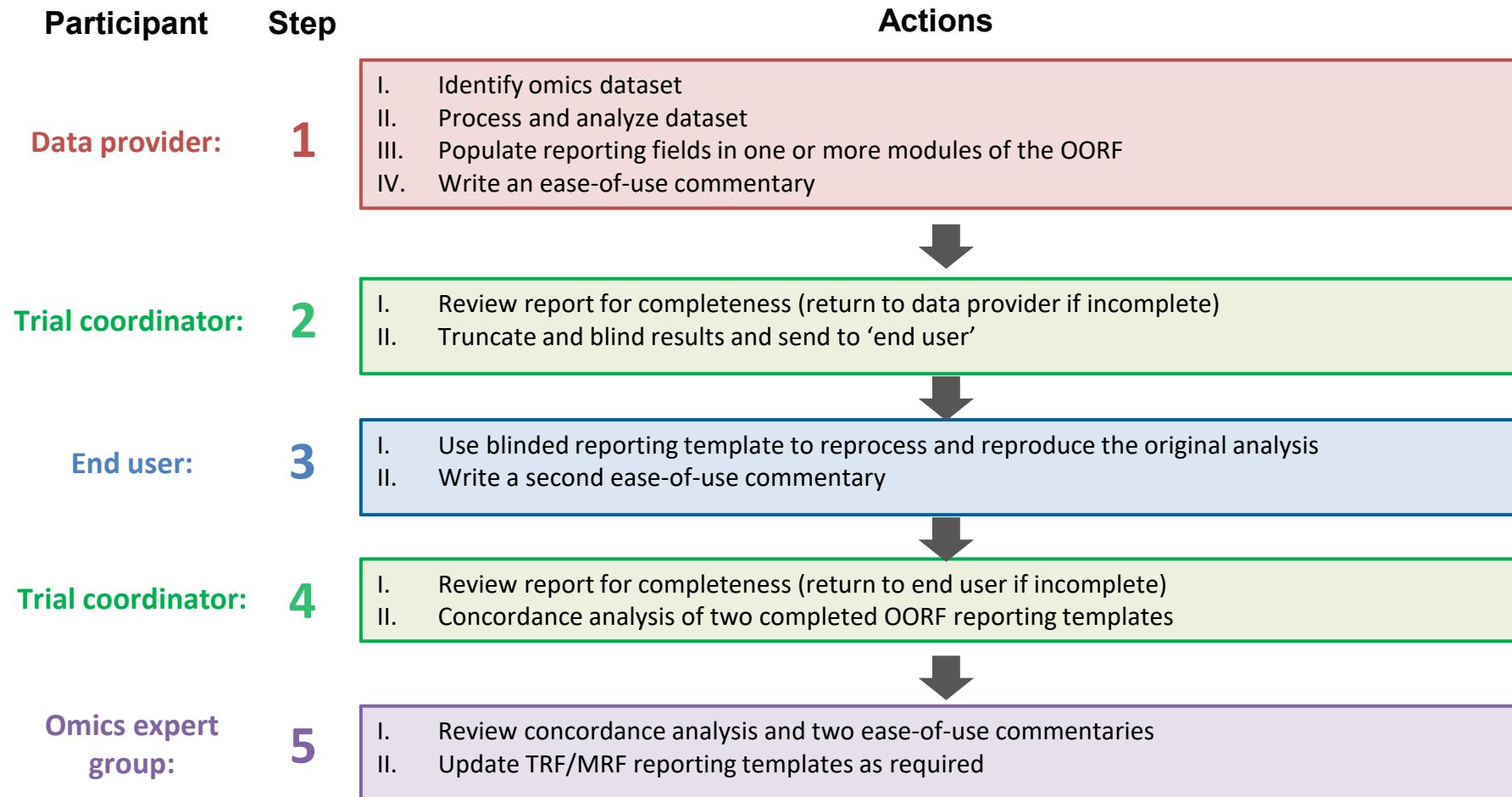
3.2.1.1. Type and version of the sequencing platform (e.g. Illumina HiSeq2500)	Recommended	III
3.2.1.2. Size and type of sequencing (e.g. 100 bp paired-end)	Recommended	50
3.2.1.3. Flow cell used (type and catalogue number)	Recommended	None
3.2.1.4. Targeting probe annotation, including list of attenuated genes (if any) (for targeted)	Recommended	Probe
3.2.1.5. Library type (e.g. mRNA libraries)	Recommended	40
3.2.1.6. Purpose (e.g. target gene expression, quality control, etc.)	Recommended	Ca
3.2.1.7. Other	Recommended	Te
<b>3.2.2.1. RNA Processing</b>		De
a. RNA Extraction		see reporting fields below
i. Type of extracted RNA (e.g. total RNA, mRNA, miRNA, etc.)	Recommended	Ce
ii. Extraction and purification techniques	Recommended	
iii. Procedures for mRNA enrichment (if applicable), or other enrichment procedures	Recommended	
iv. Storage conditions	Recommended	Ce
b. Quantification and Qualification of RNA		see reporting fields below
i. Tool for RNA assessment	Recommended	
ii. RNA quality	Recommended	
iii. RNA quantity	Recommended	
<b>3.2.2.2. Library Preparation</b>		Pr
a. Library preparation applied		see reporting fields below
i. Manual library preparation or automated systems (if yes, which automation system)	Recommended	Re
ii. Fragmentation strategy (if applicable)	Recommended	Th
iii. Probe manifest (if applicable)	Recommended	bu

DAPRM



REPORTING ELEMENT	RECOMMENDED / OPTIONAL	INPUT
4.1.1.1. Data Input(s)	Recommended	
4.1.1.2. Metadata Input(s)	Recommended	see "metadata" tab
4.1.2.1. Software	Recommended	R version 4.0.3 (2020-10-10)
4.1.2.2. Operating System	Recommended	Platform: x86_64-pc-linux-gnu (64-bit) running under: Linux
4.1.2.3. Additional Libraries used	Recommended	https://www.r-project.org/, https://www.bioconductor.org
4.1.2.4. Software Availability	Optional	
<b>4.1.3.1. Contrasts</b>	Recommended	See "contrasts" tab
4.1.4.1. Group Sizes	Recommended	See "metadata" tab
4.1.4.2. Covariance	Recommended	Samples were processed in plate-based batches. See "met
4.1.4.3. Technical Replicates	Recommended	Technical replicates were used as different wells for library exposures.
<b>4.1.5.1. Statistical Approach</b>	Recommended	DESeq2
4.1.5.2. Data Transformation	Recommended	Size factor normalization
4.1.5.3. Effects Models	Recommended	Negative Binomial Generalized Linear Model (GLM) fitting procedure. Batches were used as covariates in the formula

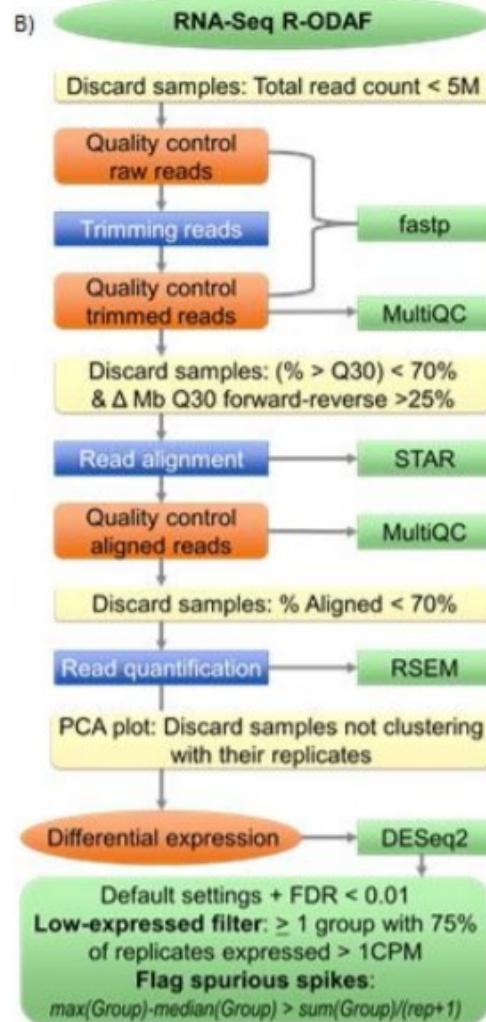
# OORF trials: how do we develop modules?



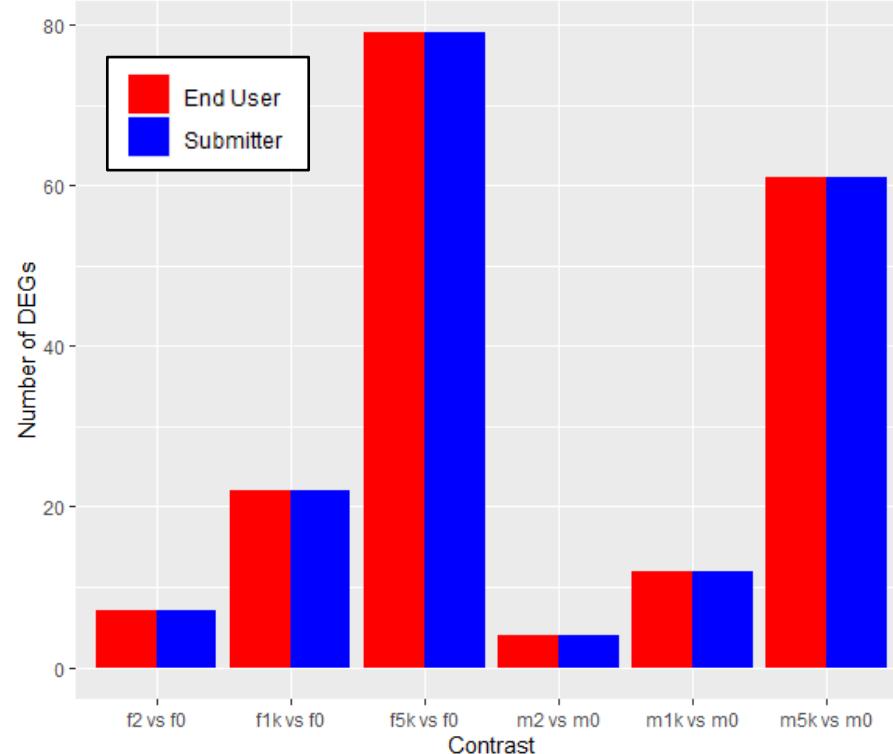
# Examples of OORF trials

Platform	Study Description	Module Tested	Computing Environment	Submitter	End User	Status
Agilent Microarray	Four-point concentration-response of furan in male and female Fisher rat liver (GEO GSE62805)	<b>DAM</b> (Custom workflow)	R	Andrew Williams (Health Canada)	Leah Wehmas (US EPA)	Complete
RNA-Seq	Three-point concentration-response of hexabromo-cyclododecane in male and female Fisher rat liver (PRJNA395549)	<b>DAM</b> (ODAF)	R	Matt Meier (Health Canada)	US EPA	Complete
		<b>BMD</b> (BMDExpress)	R	Andrew Williams (Health Canada)	Sarah Davidson (US EPA)	Complete
		<b>EARM</b> (Reactome)	R	Andrew Williams (Health Canada)	John Stead (Carleton University)	Complete
RNA-Seq	Japanese quail exposed to trenbolone (0, 1 ppm, and 10 ppm; unpublished data)	<b>DAM</b> (Eco-Omics Analyst)	Web-based tools	Krittika Mittal (McGill University)	John Martinson (US EPA)	In process
Microarray	Doxorubicin treatment in mice (3 mg/week for 8 wks; n=4)	<b>EARM</b> (Ingenuity Pathway Analysis)	Desktop software	Vikrant Vijay (US FDA)	Eunnara Cho (Health Canada)	In process
qPCR	African clawed frog exposed to benzo[a]pyrene (16.6 and 50 µg/L)	<b>qPCR</b> Module (EcoToxXplorer)	Web-based tools	Doug Crump (Environment Canada)	Jacob Collins, Dan Villeneuve (US EPA)	In process

# Trial results: Differentially Abundant Molecules DARM

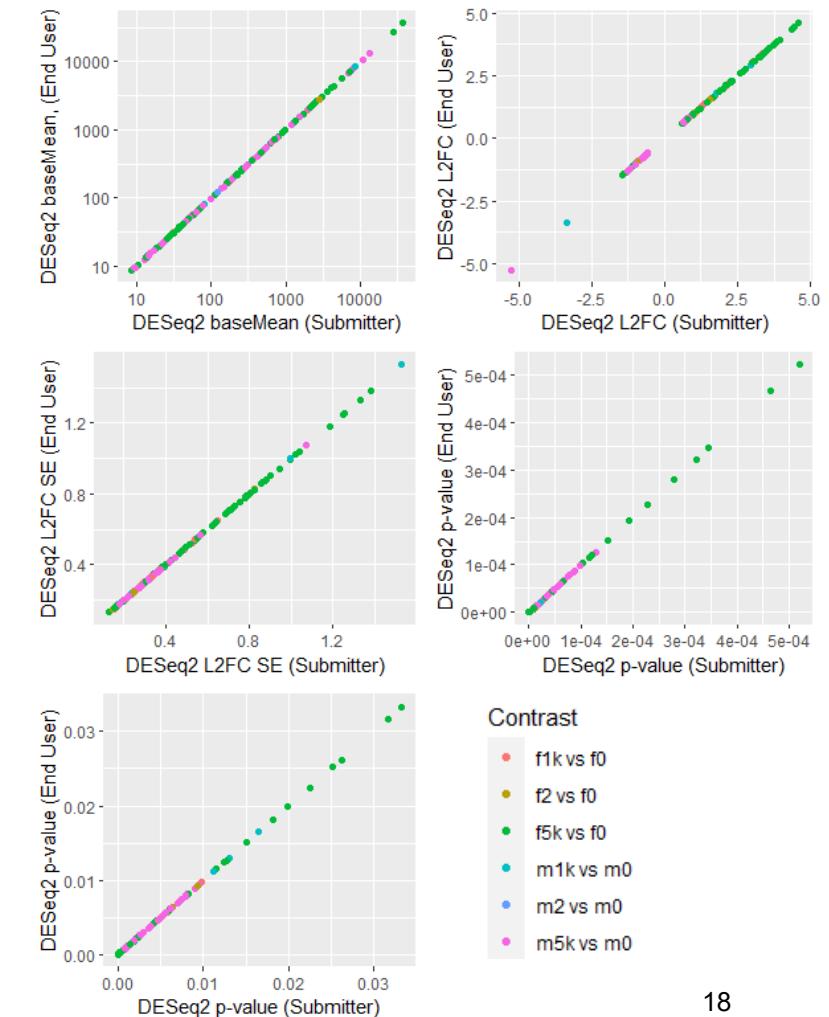


## Number of Differentially Expressed Genes



- Highly comparable results from Submitter and End User

## Correlation of DESeq2 Outputs



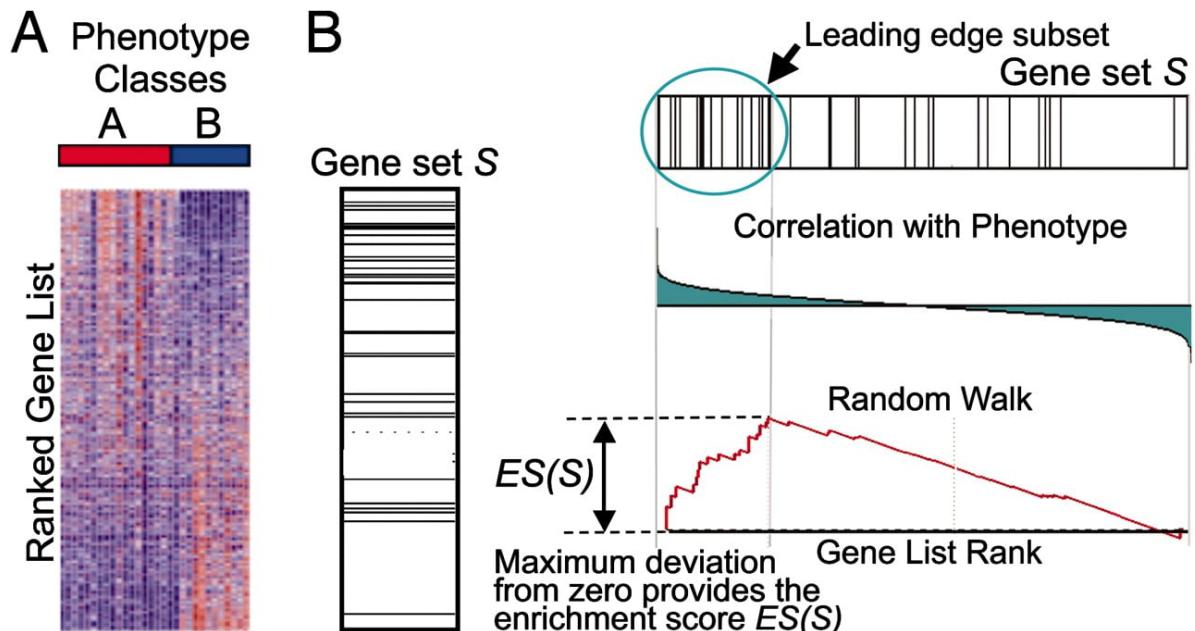
# Reporting templates – DARMs – enrichment analysis, in development

- Expert group met to reach consensus on parameter inclusion
  - Participants: Richard Beger, Carole Yauk, Timothy Ebbels, Joshua Harrill, Pim Leonards, Mark Viant, Oliver Schmitz, Magda Sachana, Vikrant Vijay, Andrew Williams
- Module & guidance drafted to capture elements across different enrichment analysis experiments
- Reviewed by experts in metabolomics, proteomics, and transcriptomics
- Conducted trials to test module
- Comments from Omics Expert Group now being addressed

8. Data Analysis Reporting Module (DARM) for Detection of Enriched Biological Pathways				INSTRUCTIONS TO DATA PROVIDERS: Use Column D for data entry. Text in <i>italics</i> are instructions (e.g. reporting tips) for the data submitter for individual reporting fields. Text in <b>bold</b> italics are instructions for the data submitter that are applicable to all reporting elements in a section. For additional clarification regarding a reporting field, please refer to the corresponding guidance document for this reporting template.
REPORTING CATEGORY	REPORTING ELEMENT	REQUIRED / OPTIONAL	INPUT	NOTES AND INSTRUCTIONS FOR DATA PROVIDERS
8.1. Software Documentation	Software and Documentation			
	Software	Recommended		Specify version(s) or repository hash of software
	Operating System	Recommended		Specify if it is a web-based application and provide the URL
	Additional Libraries used	Recommended		Specify version(s) or repository hash of additional libraries
8.2. Description of Data Used as Input for Enrichment Analysis	Software Availability	Optional		
	Data Description			
	Data used as input	Recommended		Provide a description and a <i>data object</i> that contains the data used as input to the analysis by counts matrix, list of fold-changes.
	Methods used to produce input	Recommended		Describe how the data used as input was produced. For example, differentially abundant genes by counts matrix, list of fold-changes.
	Pre-filtering of input data	Recommended		Report the methods used to pre-filter features in the data (e.g., log-fold change cut off).
8.3. Contrasts for Which Enrichment Analysis was Performed	Pre-processing and/or normalization of input data	Recommended		Report the methods used to pre-process elements in the data (e.g., transformation mean, DESeq2 normalization, etc.). If no additional processing was done, report "N/A".
	Background set(s) used	Recommended		Report the background set of features, and how it was established.
8.4. Database of Pathways or Gene Sets Used to Detect Enrichment	Contrasts			
	Contrasts	Recommended		Provide a <i>data object</i> describing the factors and levels within each factor being compared; in cases where enrichment is done on features derived from the features.
	Biological Entity or Biological Set Annotation			
	Biological Entity or Biological Set Annotation Used for the Analysis	Recommended		Report the annotation source (e.g., GO, KEGG, WikiPathways, MSigDB, IPA, custom).
Species Name				Report the species name.
Version, Date of Biological Set Annotations				Report the version used of the feature annotations; or, if not applicable, provide additional information about the annotations were obtained or downloaded. In the event that custom annotations for non-model organisms, the annotations should be provided as a <i>data object</i> , an

# Development of Enrichment Analysis DARM (transcriptomics point of view)

- **Features** to be tested for enrichment could be derived from many sources:
  - » DEGs
  - » Fold changes in expression
  - » Gene clusters (e.g., WGCNA modules)
  - » Dose-responsive genes
- **Common Statistical Methods**
  - » GSEA (Gene Set Enrichment Analysis)
  - » ORA (Over-Representation Analysis)
- **Common Databases**
  - » Reactome
  - » WikiPathways
  - » MSigDB
  - » CLUE (CMap and LINCS Unified Environment)
  - » GO
  - » KEGG
  - » IPA (Ingenuity pathway analysis: proprietary database and software, Qiagen)



# OORF trial case study

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**Trial module:** enrichment analysis DARM

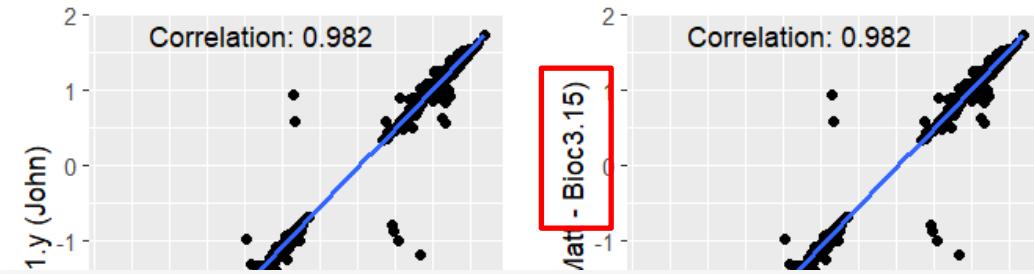
**Topic:** hexabromocyclododecane (HBCD) exposure dataset with RNA-Seq results (male and female rat liver, PRJNA395549)

**Data submitter:** Andrew Williams produced a GSEA analysis using the *fgsea* R package

**End user:** John Stead (Carleton University) reproduced the analysis

# Challenges in trials

- GSEA: Andrew provided code alongside completed EA DARM and raw data
  - John successfully ran the code
  - Result: tables of normalized enrichment scores (NES) for all Reactome pathways



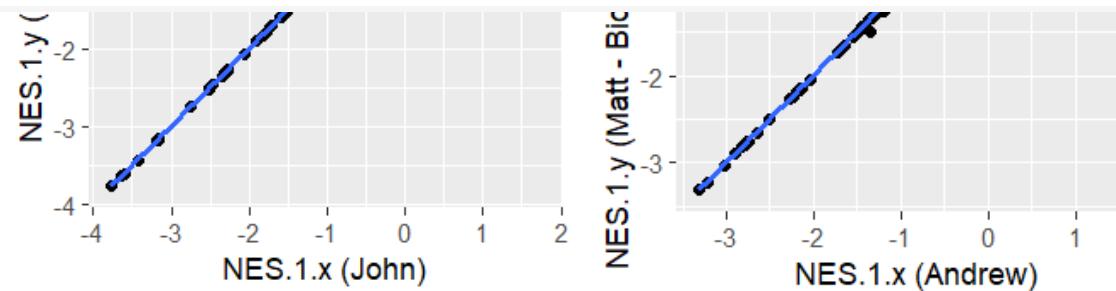
I work in an analysis core facility, and we often need to re-run old analyses. So we have R installs going back to R-3.5.0 or so, along with the associated library dirs. And I am old and stuff, so I use emacs/ESS with various scripts so M-x R will start the correct R version. So it's easy for me to do small additional things to existing analyses without having to update entirely. Which is good, primarily for annotations, because people don't like it when their results change. Genes come into and out of existence with some regularity, and losing the top gene just because NCBI no longer thinks it's a thing is hard to explain to some people.

[ADD REPLY](#)

• [link](#)

2.4 years ago James W. MacDonald ⚡ 64k

- The main determinant in this example turned out to be the reactome.db version: **however – big caveat – the version used by Andrew (1.79, correctly reported in the reporting module) was no longer available! Anywhere!!**



# Lessons learned from OORF trials: analysis reproducibility

---

Analyses in **open-source computing environments** (R, Python, etc.)

- Reproducibility depends less on reporting fields and more on code/scripts (versions still important)
- Issue: users may not have sufficient expertise with open-source computing environments (easier for end users with coding skills to reproduce)
- No financial or licensing barriers for accessing tools

Analyses using **freeware analysis software or web applications** (BMDExpress)

- Reproducibility depends on clear and precise reporting in the OORF documentation and/or a configuration file the end user could follow
- More user friendly and require less technical/statistical expertise compared to open-source computing
- No “pay wall”

Analyses using **proprietary software** (Partek, Ingenuity, etc.):

- Reproducibility depends on clear and precise reporting in the OORF documentation and/or a configuration file the end user could follow
- End user needs access to the same software (and version)
- “Pay wall” issues

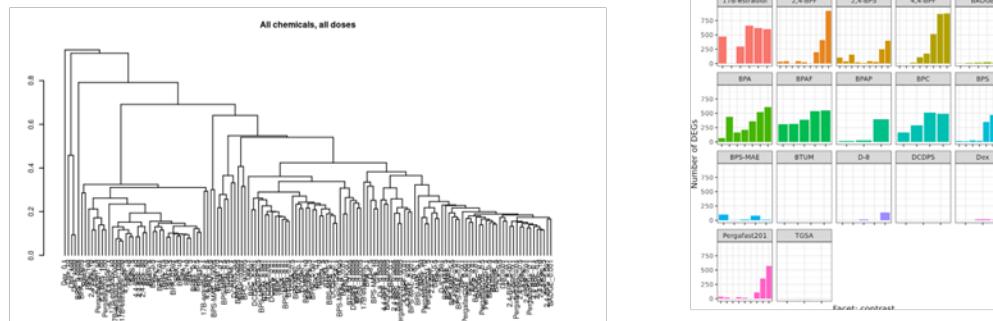
# Lessons learned from OORF trials: big picture

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- Trials have demonstrated where clarifications/revisions in the OORF were needed
- High degree of concordance observed in the trial results (i.e., the framework works)
  - **Minor differences** in pipelines that implement permutation analyses (expected but not major differences), and different database versions (also expected)
  - **Differences would not change study conclusions**
- Challenges include finding submitters and end-users that have access to, or are willing to apply, the same software or pipeline
  - Paywall issues with some software
  - Complexity of using other people's pipelines
    - » GitHub repositories and Omics Data Analysis Framework for Regulatory Application (R-ODAF) or EPA's httrpl are solutions
- Experience with regulatory partners demonstrates utility for increasing transparency and reproducibility of the omics analyses

# Application Reporting Module case study: chemical grouping using gene expression data for a common group of chemicals (BPA and alternatives)

- Qualitative Analysis:
  - *Bioactivity profile-based grouping* using the complete set of features (top figures)
  - Established thresholds to focus on chemicals with demonstrated bioactivity after exposure
- Grouping Analysis:
  - *Bioactivity profile-based grouping* using a subset of pre-filtered differentially expressed genes
  - *Omics signature-based grouping* using subset of genes from a published estrogen receptor biomarker
- The methods, approaches, and results are being used to develop and trial the reporting module



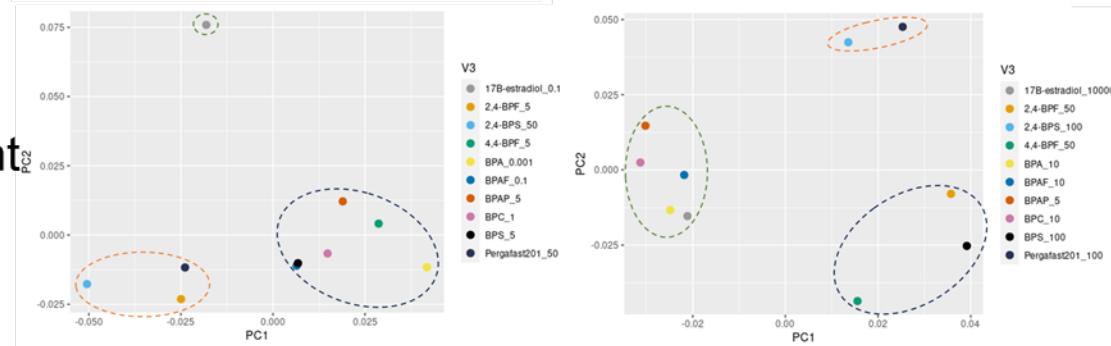
Applying thresholds for bioactivity (% DEGs)



## Hierarchical Cluster Analysis (HCA)



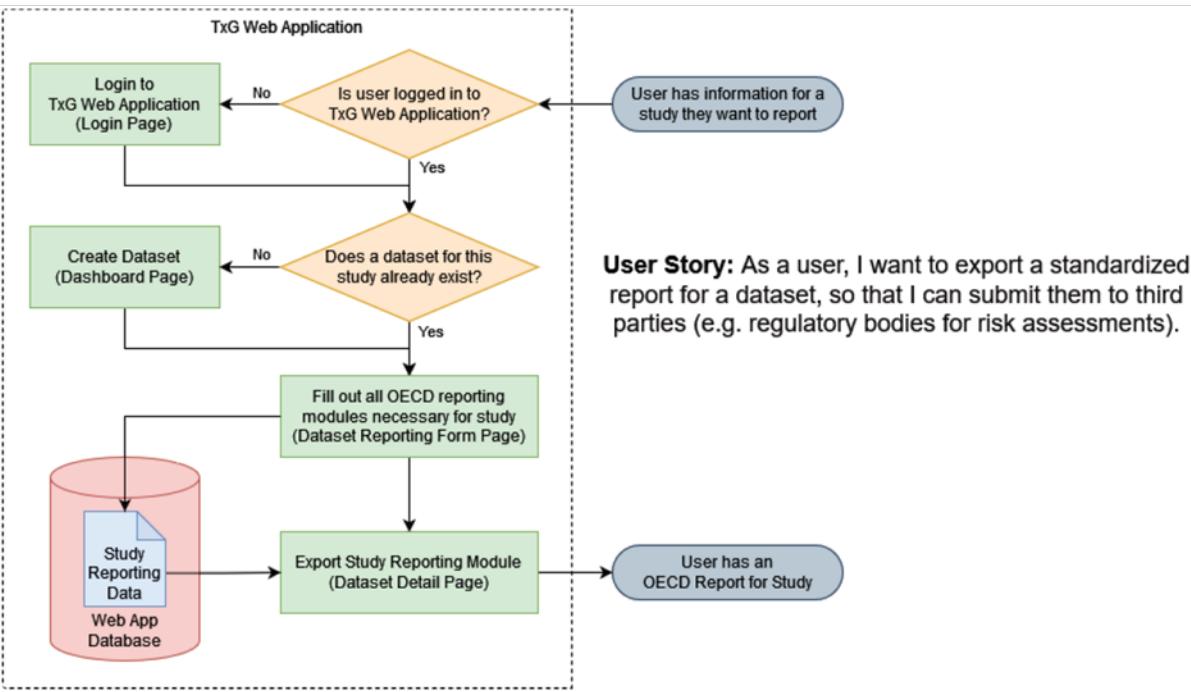
## Principal Component Analysis (PCA)



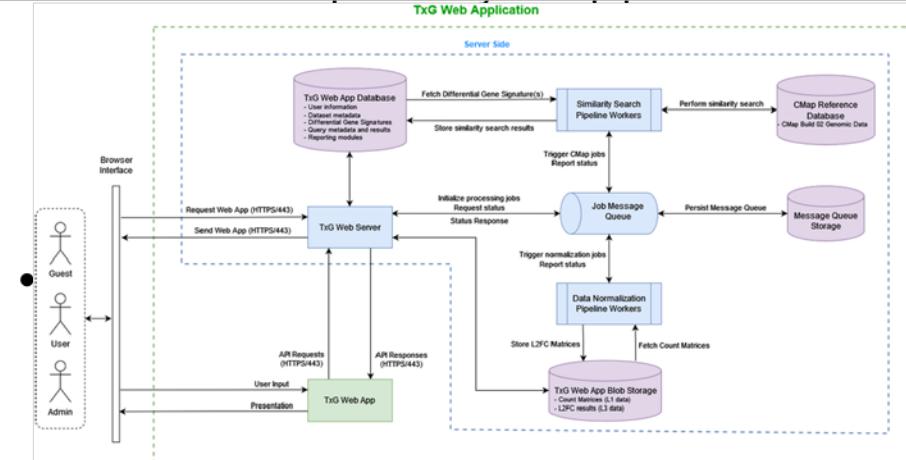
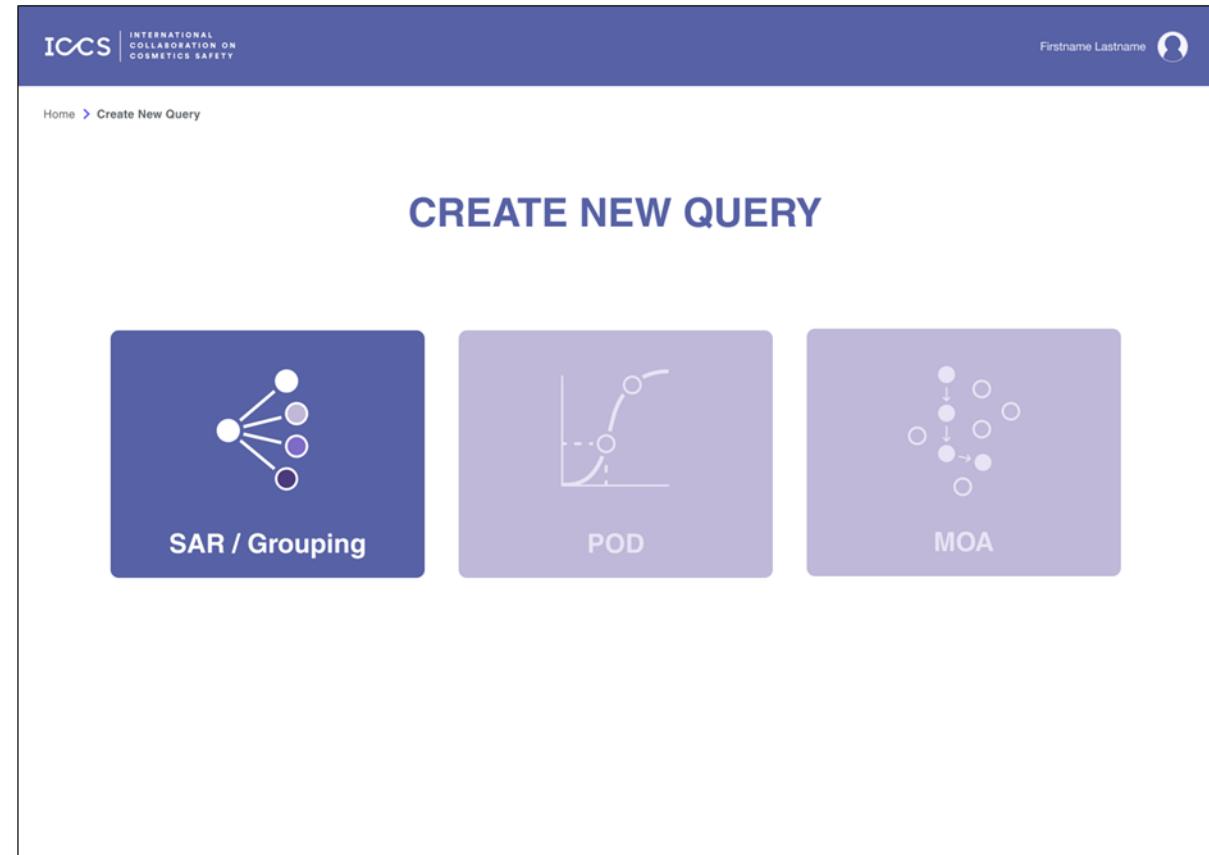
Lowest Conc.

Highest Conc.

# ICCS Toxicogenomics Web Application



data and



# Future of the OORF: the “last mile” of NAMs?

- Reporting module for ‘omics biomarkers/pattern matching
  - Outcome from discussion at International Workshop on Genotoxicity Testing (IWGT) in Ottawa (2022), Working group on “TGx-Biomarkers that Predict Genotoxicity”
- OECD Omics website provides a resource for sharing and disseminating the OORF
  - <https://www.oecd.org/en/topics/sub-issues/testing-of-chemicals/omics-technologies-chemical-testing.html>
- Application Reporting Modules (ARMs)
  - Development and trialling of ‘Chemical Grouping’ ARM adopted into WPHA work plan in March 2022
- Revise and update as required
  - It would be valuable to harmonize the format of the OORF into fully machine-readable schema
- Health Canada is aiming for better interoperability with international agencies for consistency in analytical approaches used for transcriptomics
- The OORF contributes guidance and a framework, key components for getting NAMs implemented in practice
  - Case studies, training, and implementation are the other key components for omics use in decision-making
  - A remaining challenge is encouraging regulators to request a filled OORF where omics experiments may have been done (solution: make it easier for people to fill out, e.g., ICCS web application; education on use – that is, teach about modular nature and re-useability of components)



# Collaborators and Acknowledgements



## University of Ottawa

Dr. Carole Yauk  
The Genomics in Regulatory and Applied Toxicology Laboratory



## Health Canada, PSD

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## Health Canada, MSD

Dr. Lauren Bradford  
Dr. Tanvi Sharma  
Annette Dodge  
Andrea Rowan-Carroll  
Julie Buick

## Health Canada, ESRAB

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Dr. Anthony Reardon  
Dr. Alexandra Long

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Nadira De Abrew



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Dr. Russell Thomas  
Dr. Joshua Harrill  
Dr. Logan Everett

## Maastricht University

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## Kinetic Vision

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Kelly Semrau

## ICCS TxG WG

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Genomics Research & Development Initiative  
Chemicals Management Plan  
Regulatory partners - Health Canada  
NSERC  
Canada Research Chairs Program



Health  
Canada

Santé  
Canada

GENOMICS  
R&D INITIATIVE

# Recent Developments in High-Throughput Toxicokinetics

John Wambaugh, Barbara Wetmore, Caroline Ring, Sarah Davidson-Fritz,  
Gilberto Padilla Mercado, Meredith Scherer, Sahar Tabatabaei Sadeghi,  
Colin Thompson, Rachael Cogbill, Risa Sayre, Rogelio Tornero-Velez,  
Elaina Kenyon, Marina Evans, Kimberly Truong, Taylor Wall,  
Katie Paul Friedman, Michael Devito, and Russell Thomas

*Center for Computational Toxicology and Exposure  
U.S. Environmental Protection Agency*

November 6, 2024

The views expressed in this presentation are those of the author  
and do not necessarily reflect the views or policies of the U.S. EPA

# Estimating Points of Departure (PODs) using New Approach Methods (NAMs)

- *In vitro-in vivo* extrapolation (IVIVE) allows estimation of chemical-specific Points of Departure (PODs) based on new approach methods (NAMs)

*In Vitro* Measured Bioactive  
Point of Departure ( $POD_{in\ vitro}$ )

$\mu M$

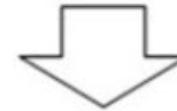
Paul Friedman et al. (2020)

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Apply high-  
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mg/kg/day

*in vitro-in vivo* extrapolation  
(IVIVE) converts  $\mu M$  to  
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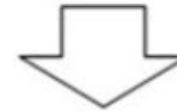
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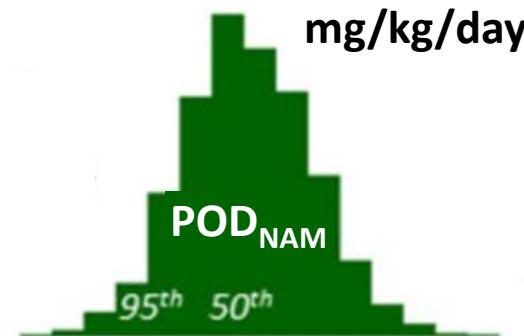
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Paul Friedman et al. (2020)



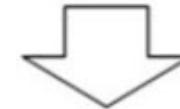
Distribution of  $POD_{NAM}$  for a single chemical

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Paul Friedman et al. (2020)

mg/kg/day

Exposure

95<sup>th</sup>

mg/kg/day

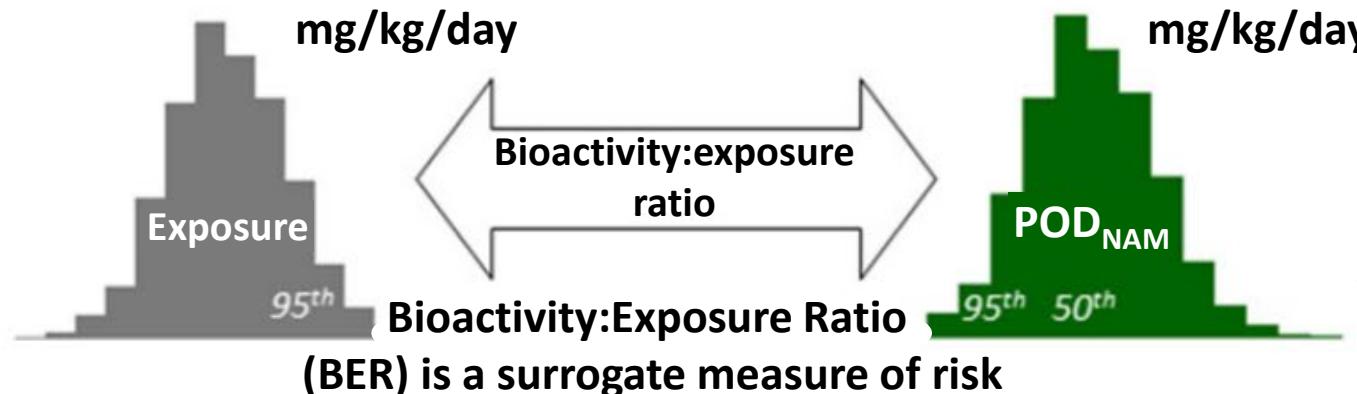
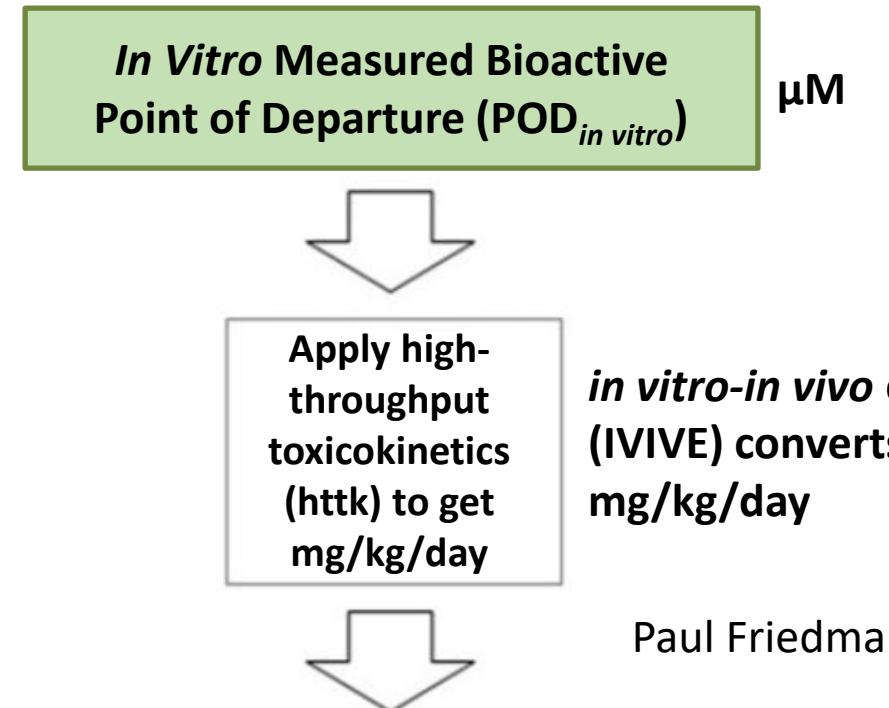
$POD_{NAM}$

95<sup>th</sup> 50<sup>th</sup>

Uncertainty in intake rate for single chemical

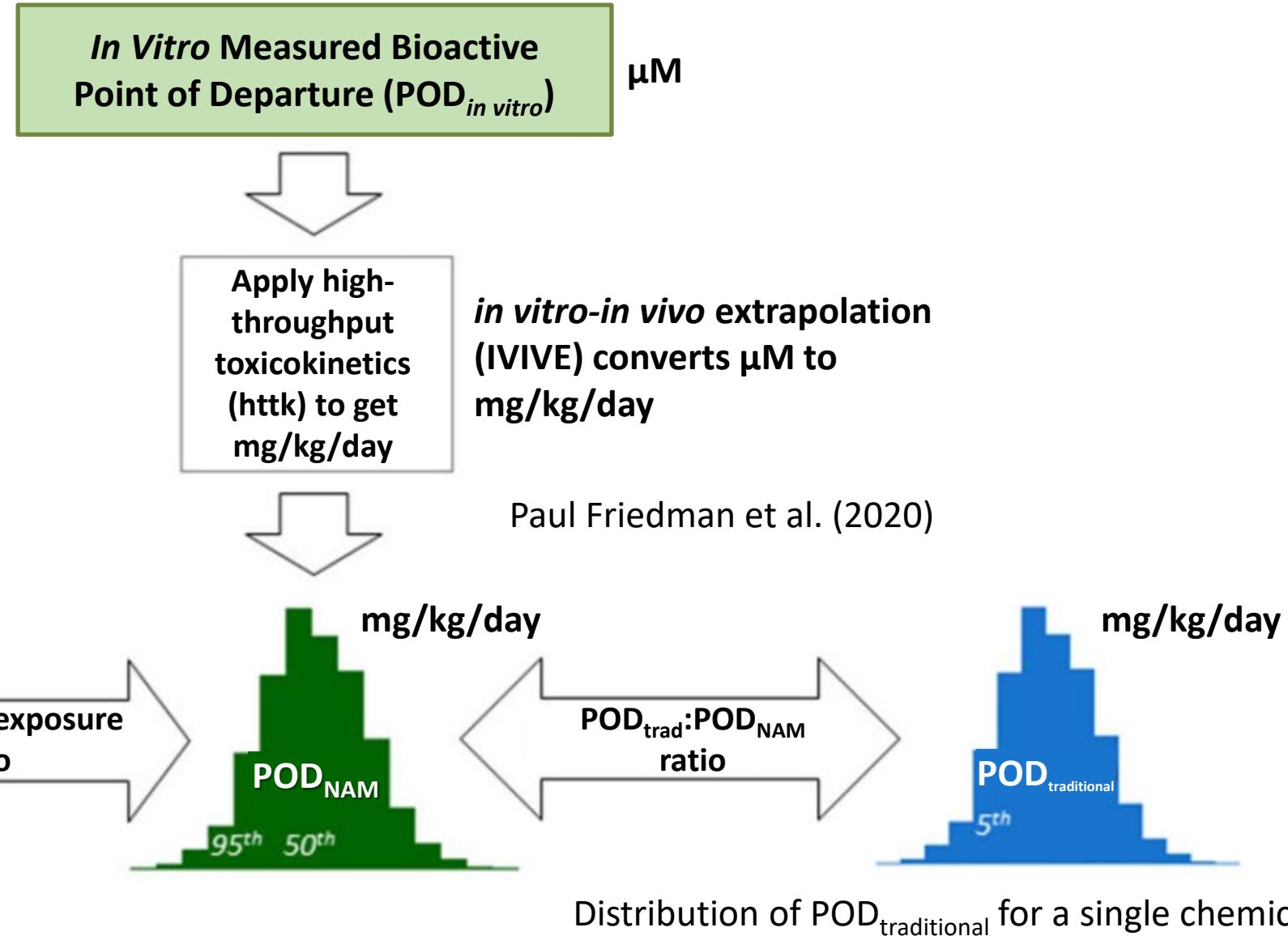
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- *In vitro-in vivo* extrapolation (IVIVE) allows estimation of chemical-specific Points of Departure (PODs) based on new approach methods (NAMs)
- Conservative assumptions allow calculation of a protective  $POD_{NAM}$  that is less than  $POD_{traditional}$



# Estimating Points of Departure (PODs) using New Approach Methods (NAMs)

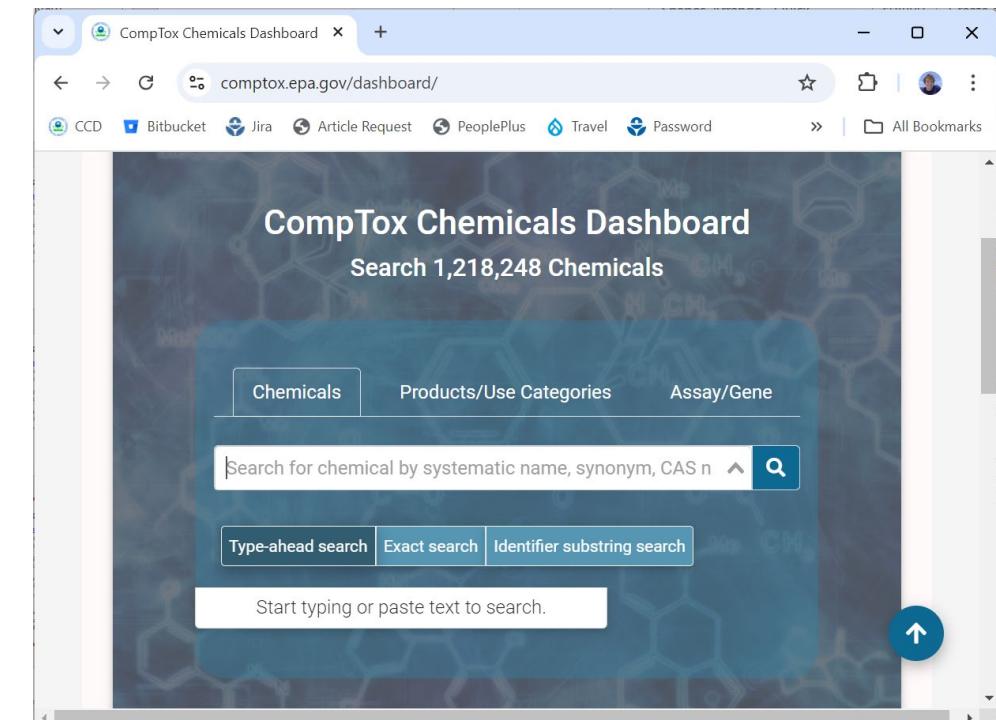
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# Standardized NAM Data and Tools

<https://comptox.epa.gov/dashboard>

- **Hazard (ToxCast/Tox21):** There are nearly 10,000 chemicals with *in vitro* bioactivity data
- **Exposure:** There are more than 400,000 chemicals with “exposure forecasts” (ExpoCast)
- **Dose-Response:** There are currently 7,569 chemicals with high throughput toxicokinetics (HTTK) data/predictions (including  $C_{ss}$ ,  $V_d$ ,  $t_{half}$ )

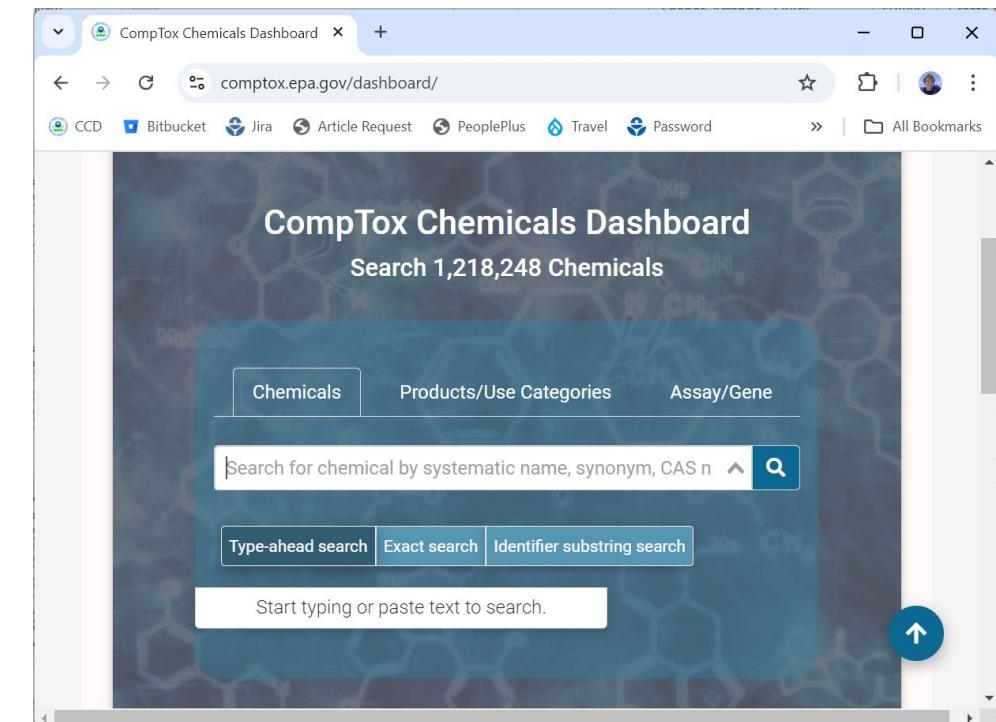


# Standardized NAM Data and Tools

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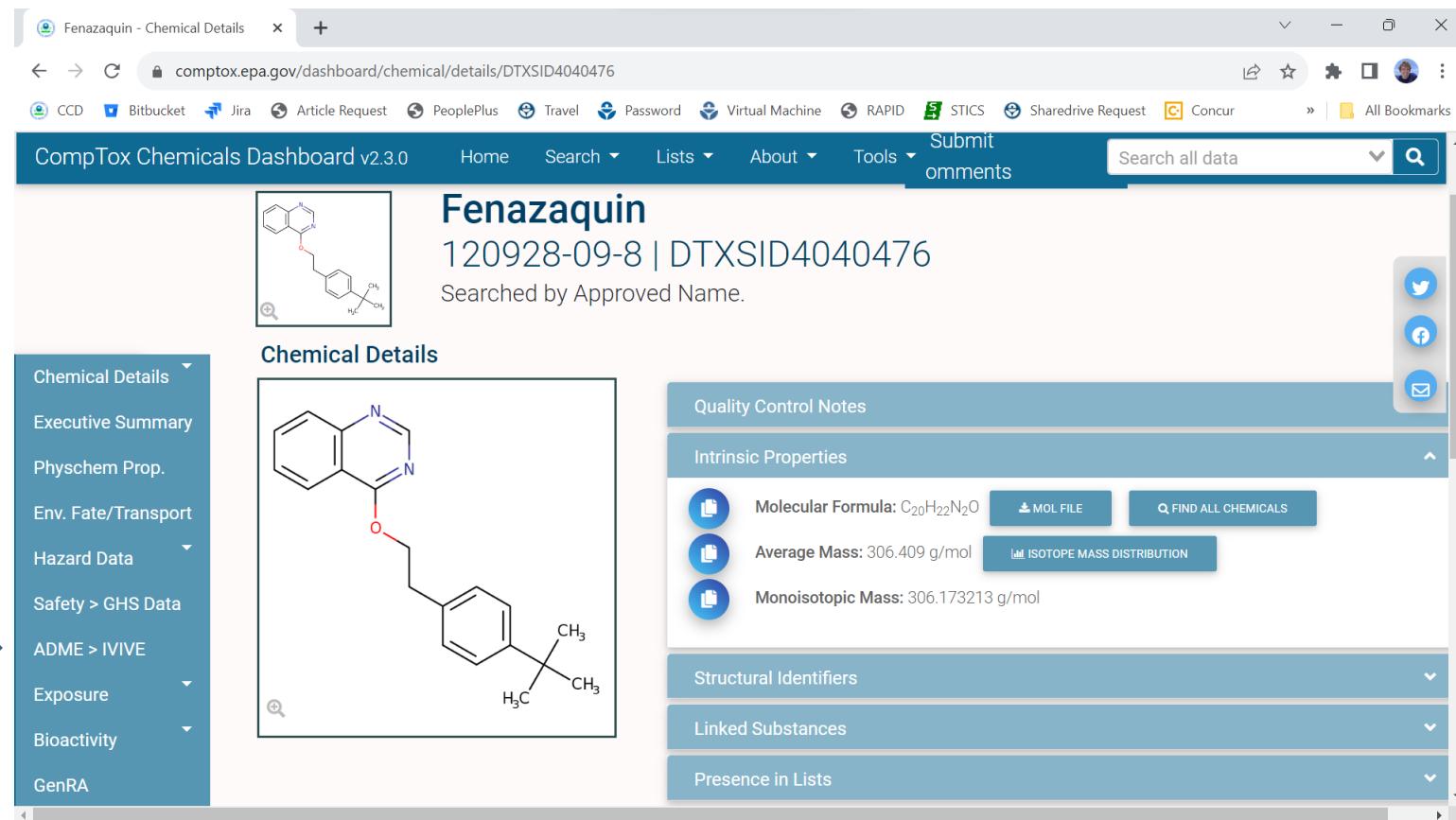
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**HTTK is the combination of  
chemical-specific *in vitro* TK data and  
generic physiologically-based TK models  
(Breen et al., 2021)**



# Openly Available TK Information

- EPA's data and tools for HTTK are made available through R package "httk"
- The "httk" tool has been used to calculate key TK information that is available on the CompTox Chemicals Dashboard and elsewhere



Fenazaquin - Chemical Details

comptox.epa.gov/dashboard/chemical/details/DTXSID4040476

CompTox Chemicals Dashboard v2.3.0

Home Search Lists About Tools Submit comments

Search all data

**Fenazaquin**  
120928-09-8 | DTXSID4040476  
Searched by Approved Name.

**Chemical Details**

**Quality Control Notes**

**Intrinsic Properties**

- Molecular Formula: C<sub>20</sub>H<sub>22</sub>N<sub>2</sub>O
- Average Mass: 306.409 g/mol
- Monoisotopic Mass: 306.173213 g/mol

**Structural Identifiers**

**Linked Substances**

**Presence in Lists**

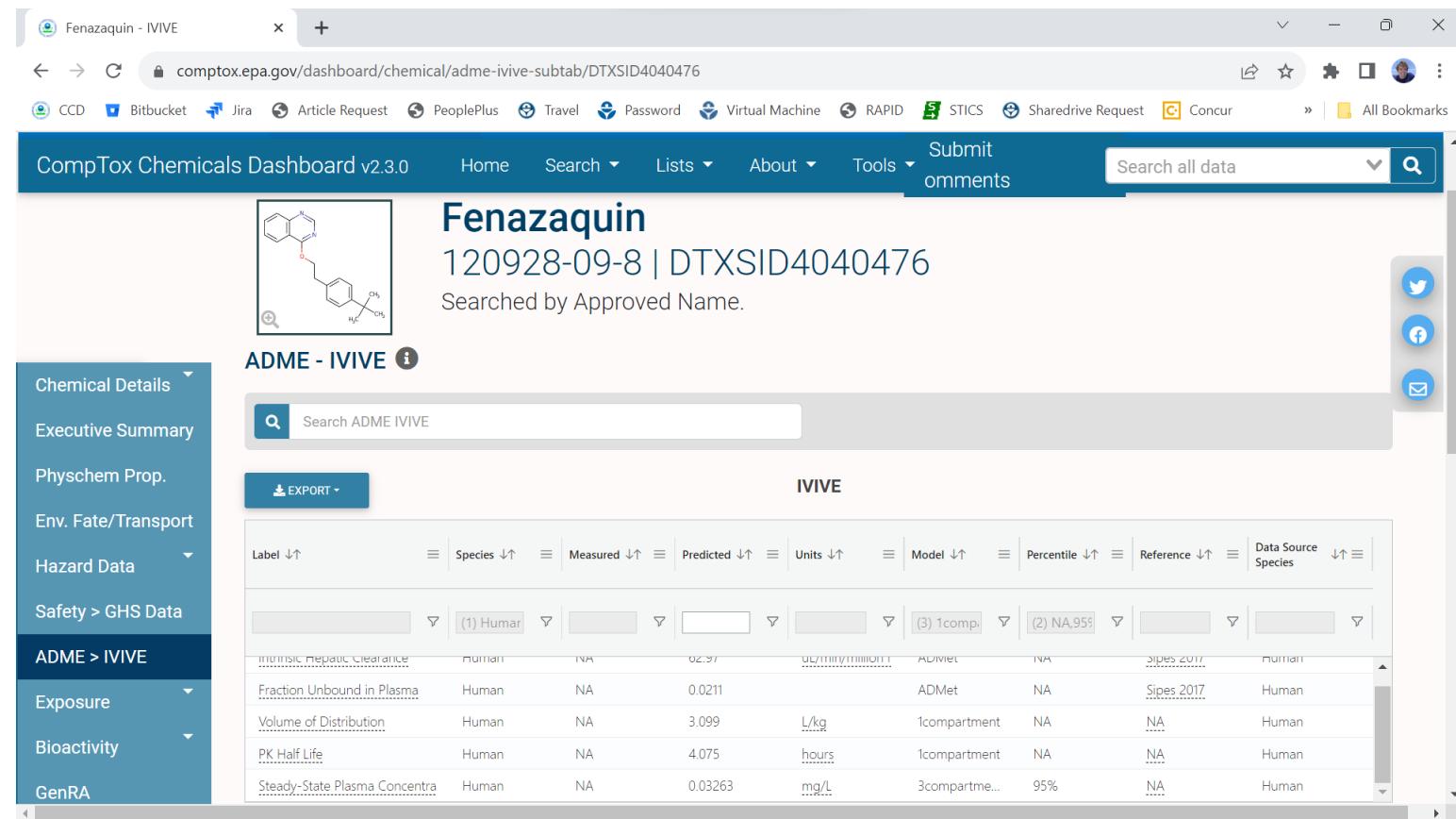
Chemical structure of Fenazaquin: CN1C=NC2=C1C=C(C=C2)OCC3=CC=C(C=C3)C(C)(C)C

<https://comptox.epa.gov/dashboard>

The current HTTK data in CCD is HTTK v2.2.1. Please see the Data Sources table in the [Release Notes](#) for more information

# Openly Available TK Information

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The screenshot shows the CompTox Chemicals Dashboard v2.3.0. The main page displays information for Fenazaquin, with its chemical structure and CAS number (120928-09-8 | DTXSID4040476). The search bar shows "Fenazaquin". The left sidebar has a "Chemical Details" dropdown and links to "Executive Summary", "Physchem Prop.", "Env. Fate/Transport", "Hazard Data", "Safety > GHS Data", "ADME > IVIVE" (which is selected), "Exposure", "Bioactivity", and "GenRA". The "ADME > IVIVE" section shows a table of pharmacokinetic parameters for Fenazaquin, including:

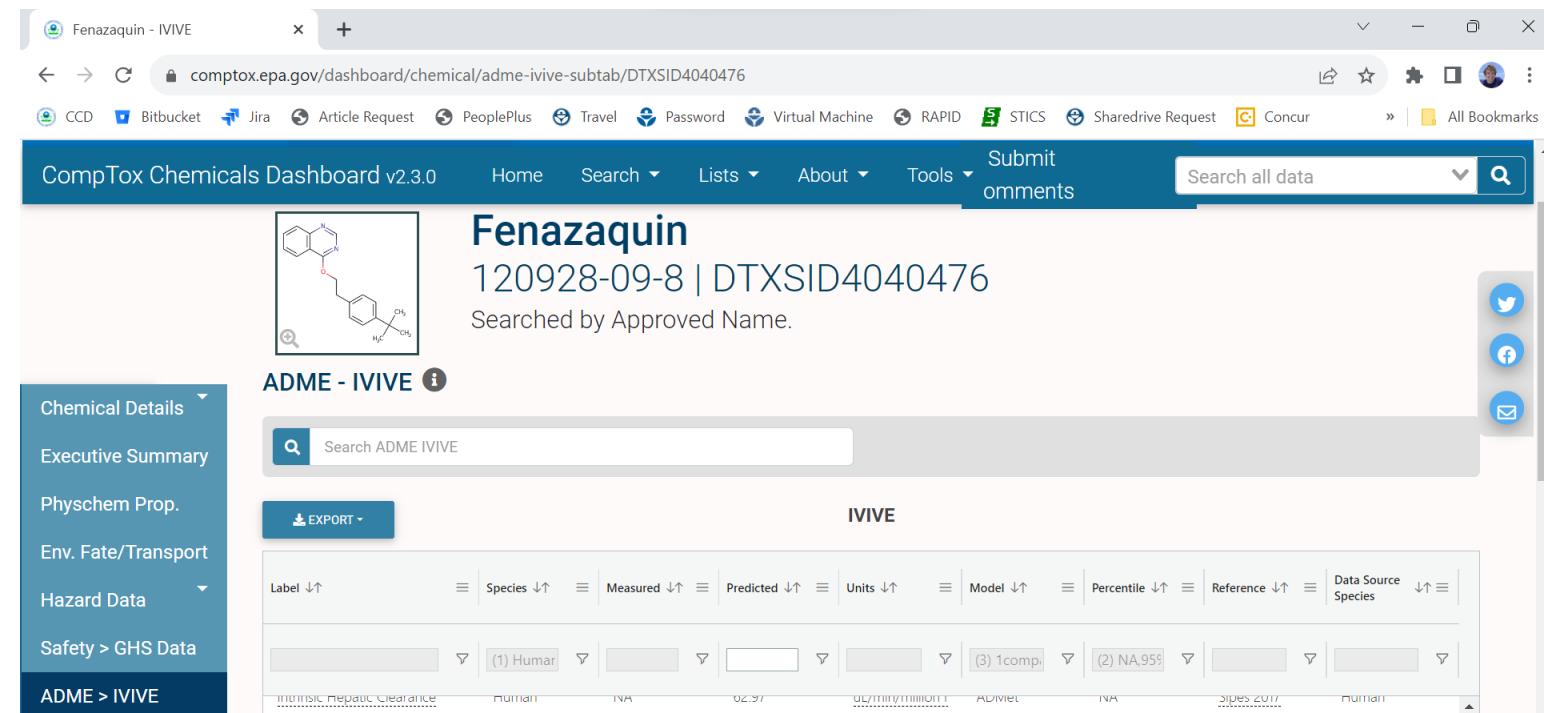
Parameter	Species	Value	Unit	Model	Reference	Data Source	
Intrinsic Hepatic Clearance	Human	NA	02.97	0.0297	ADMet	NA	30/05/2017
Fraction Unbound in Plasma	Human	NA	0.0211	ADMet	NA	Sipes, 2017	Human
Volume of Distribution	Human	NA	3.099	1compartment	NA	NA	Human
PK Half Life	Human	NA	4.075	hours	1compartment	NA	NA
Steady-State Plasma Concentra	Human	NA	0.03263	mg/L	3compartment	95%	NA

<https://comptox.epa.gov/dashboard>

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Fenazaquin - IVIVE

comptox.epa.gov/dashboard/chemical/adme-ivive-subtab/DTXSID4040476

CompTox Chemicals Dashboard v2.3.0

Home Search Lists About Tools Submit comments

Search all data

Fenazaquin  
120928-09-8 | DTXSID4040476  
Searched by Approved Name.

ADME - IVIVE

Search ADME IVIVE

EXPORT

IVIVE

Label	Species	Measured	Predicted	Units	Model	Percentile	Reference	Data Source
Intrinsic Hepatic Clearance	(1) Human	NA	02:97	U/min/mmol	ADmet	NA	30/5 2017	Human

Chemical Details

Executive Summary

Physchem Prop.

Env. Fate/Transport

Hazard Data

Safety > GHS Data

ADME > IVIVE

$V_d$	Volume of Distribution	Human	NA	3.099	L/kg	1compartment	NA	NA	Human
$t_{half}$	PK Half Life	Human	NA	4.075	hours	1compartment	NA	NA	Human
$C_{ss}$	Steady-State Plasma Concentra	Human	NA	0.03263	mg/L	3compartme...	95%	NA	Human

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# IVIVE by Scaling Factor

- There are many approaches to IVIVE, but we choose a relatively simple one:
- We make various assumptions that allow conversion of an *in vitro* concentration  $[X]$  ( $\mu\text{M}$ ) into an **administered equivalent dose** (AED) with units of mg/kg body weight/day:

$$\text{AED} = F_{\text{IVIVE}} \times [X]$$

- AED is the **external dose rate** that would be needed to **produce a given steady-state plasma concentration**
- $F_{\text{IVIVE}}$  is a scaling factor that varies by chemical

**HTTK can predict  $F_{\text{IVIVE}}$**

# IVIVE by Scaling Factor

**“httk” predicts IVIVE scaling factors using probabilistic methods that account for human variability and measurement uncertainty**

- For a given chemical,  $F_{IVIVE} = 1 / C_{ss,95}$
- $C_{ss,95}$  is the steady-state plasma concentration resulting from a 1 mg/kg/day exposure
- HTTK can predict  $C_{ss,95}$  using “reverse dosimetry” IVIVE (Tan et al., 2007), leading to an oral equivalent dose (OED):

$$OED_{95} = \frac{[X]}{C_{ss,95}}$$

- The “95” refers to the upper 95<sup>th</sup> percentile – due to human variability and measurement uncertainty there are a range of possible  $C_{ss}$  values

# Means of Obtaining HTTK Scaling Factors

- EPA's R package "httk" (targeting bioinformatics community) (Pearce et al., 2017)  
<https://CRAN.R-project.org/package=httk>
- CompTox Chemicals Dashboard (in use by US EPA) (Williams et al., 2017)  
<https://comptox.epa.gov/dashboard/>
- SimCYP SimRFlow Tool (in use by EU-ToxRisk) (Khalidi et al., 2022)  
<https://www.certara.com/software/simcyp-pbpk/>
- NICEATM Integrated Chemistry Environment (in use by US NTP) (Bell et al., 2020)  
<https://ntp.niehs.nih.gov/whatwestudy/niceatm/comptox/ct-ivive/ivive>
- TKPlate (in use by EFSA) (Dorne et al., 2023)  
<https://zenodo.org/record/2548850>

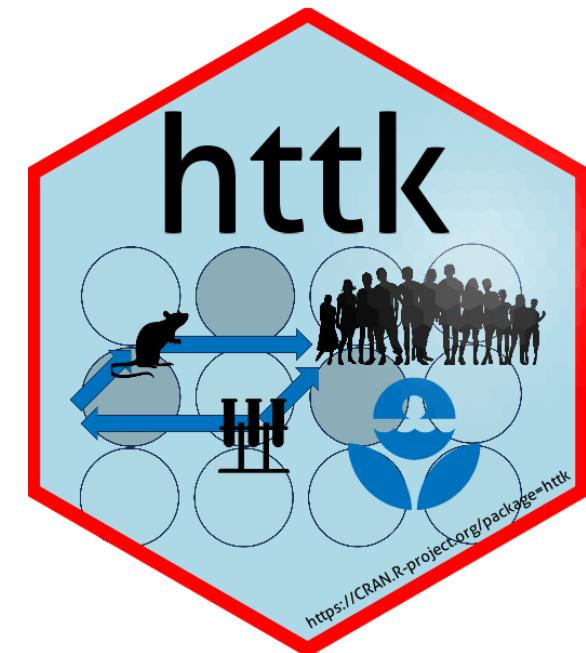
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- SimCYP SimRFlow Tool (in use by EU-ToxRisk) (Khalidi et al., 2022)  
<https://www.certara.com/software/simcyp-pbpk/>
- NICEATM Integrated Chemistry Environment (in use by US NTP) (Bell et al., 2020)  
<https://ntp.niehs.nih.gov/whatwestudy/niceatm/comptox/ct-ivive/ivive>
- TKPlate (in use by EFSA) (Dorne et al., 2023)  
<https://zenodo.org/record/2548850>

**All these tools make use of data/models from EPA's open-source "httk" package**

# Recent Updates to EPA “httk” Package

- Updated “httk-pop” human variability simulator to reflect most recent NHANES cohorts (Breen et al., 2022)
- Developed human gestational exposure model (Kapraun et al., 2022)
- Measured *in vitro* gut permeability data using Caco-2 cell-line for non-pharmaceuticals
  - Values allow prediction of chemicals that are poorly absorbed orally (Honda et al., 2024)
- EPA now analyzing *in vitro* TK data with upcoming tool “invitroTKstats”
  - “invitroTKstats” is a standardized workflow that allows auditable and reproducible analysis
  - Analysis estimates chemical-specific measurement uncertainty



<https://CRAN.R-project.org/package=httk>

# Adjusting for Oral Absorption

- *In vitro* Caco-2 measurements (or QPSRs) characterize absorption from the gut
- Administered equivalent dose (AED) depends on predicted steady-state plasma concentration:

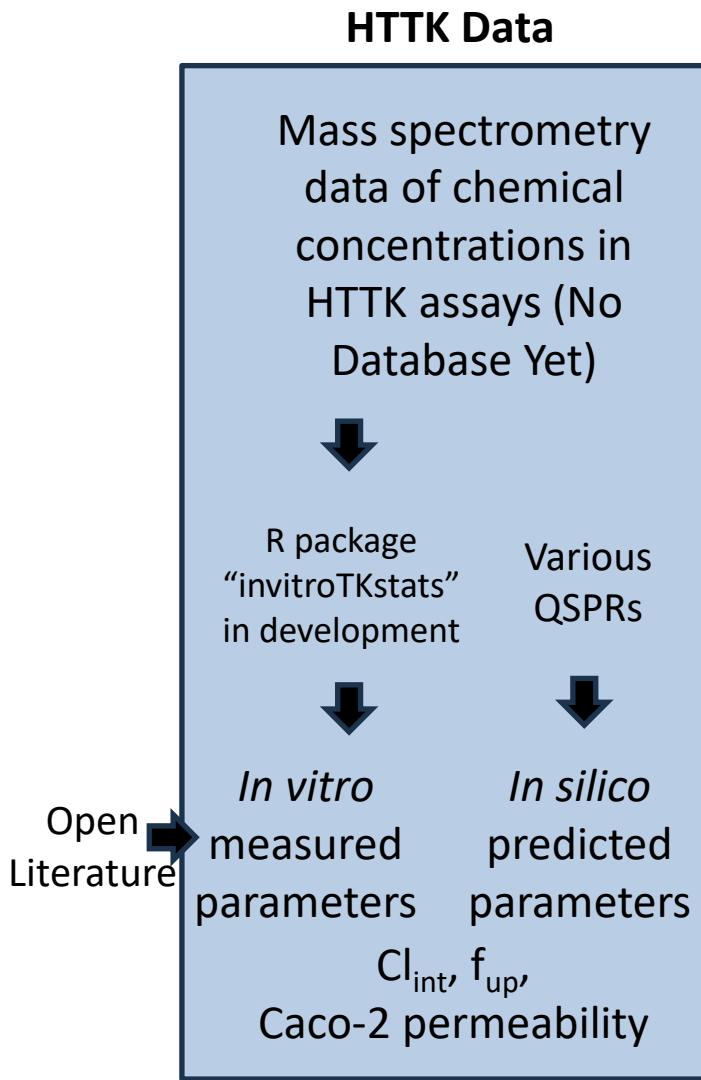
$$\text{OED}_{95} = \frac{[X]}{C_{ss,95}}$$

- Steady-state plasma concentration is proportional to dose and fraction absorbed ( $F_{abs}$ ):

$$C_{ss,95} \sim \text{dose} * F_{abs}$$

- If  $F_{abs} < 100\%$ , then  $C_{ss,95}$  decreases and therefore the necessary  $\text{AED}_{95}$  increases

# New *In Vitro* TK Measurements

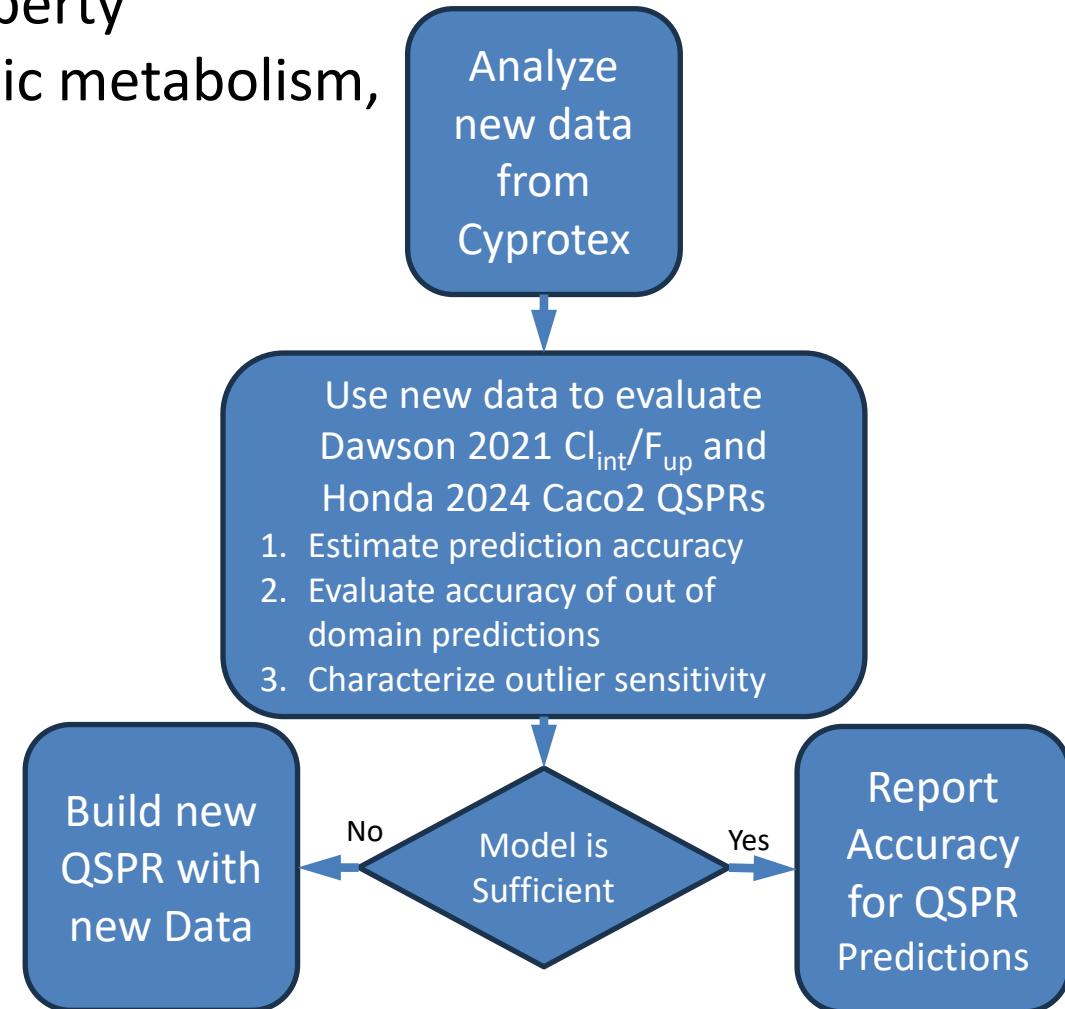


R package "httk"

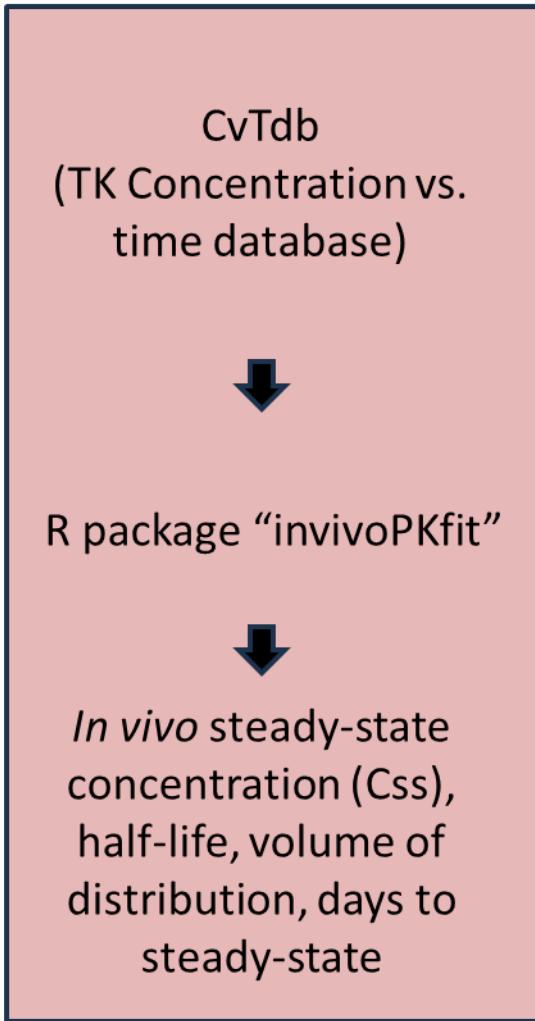
- EPA's laboratories (Wetmore), contractors (Cyprotex), and collaborators (National Toxicology Program, EC Joint Research Centre, Health Canada) continue to generate new, chemical-specific *in vitro* TK measurements
- These data are analyzed with unreleased tool "invitroTKstats" to produce chemical-specific estimates of measurement uncertainty
- Literature *in vitro* TK data curated by ICF

# HTTK QSPRs

- Machine-learning based quantitative structure-property relationship (QSPR) models now available for hepatic metabolism, plasma protein binding, and Caco-2 (Dawson et al., 2021, Honda et al., 2024)
- Model domains of applicability indicate chemical properties that are consistent with the training set
- Nearly 10,000 predictions based on HTTK QSPRs available on the CompTox Chemicals Dashboard
- New *in vitro* TK measured data being used to establish accuracy of QSPRs and build new models (Tabatabaei Sadeghi, in preparation)

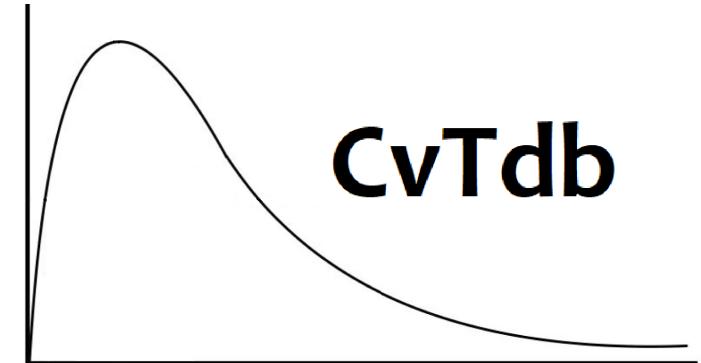


## Characterizing Confidence



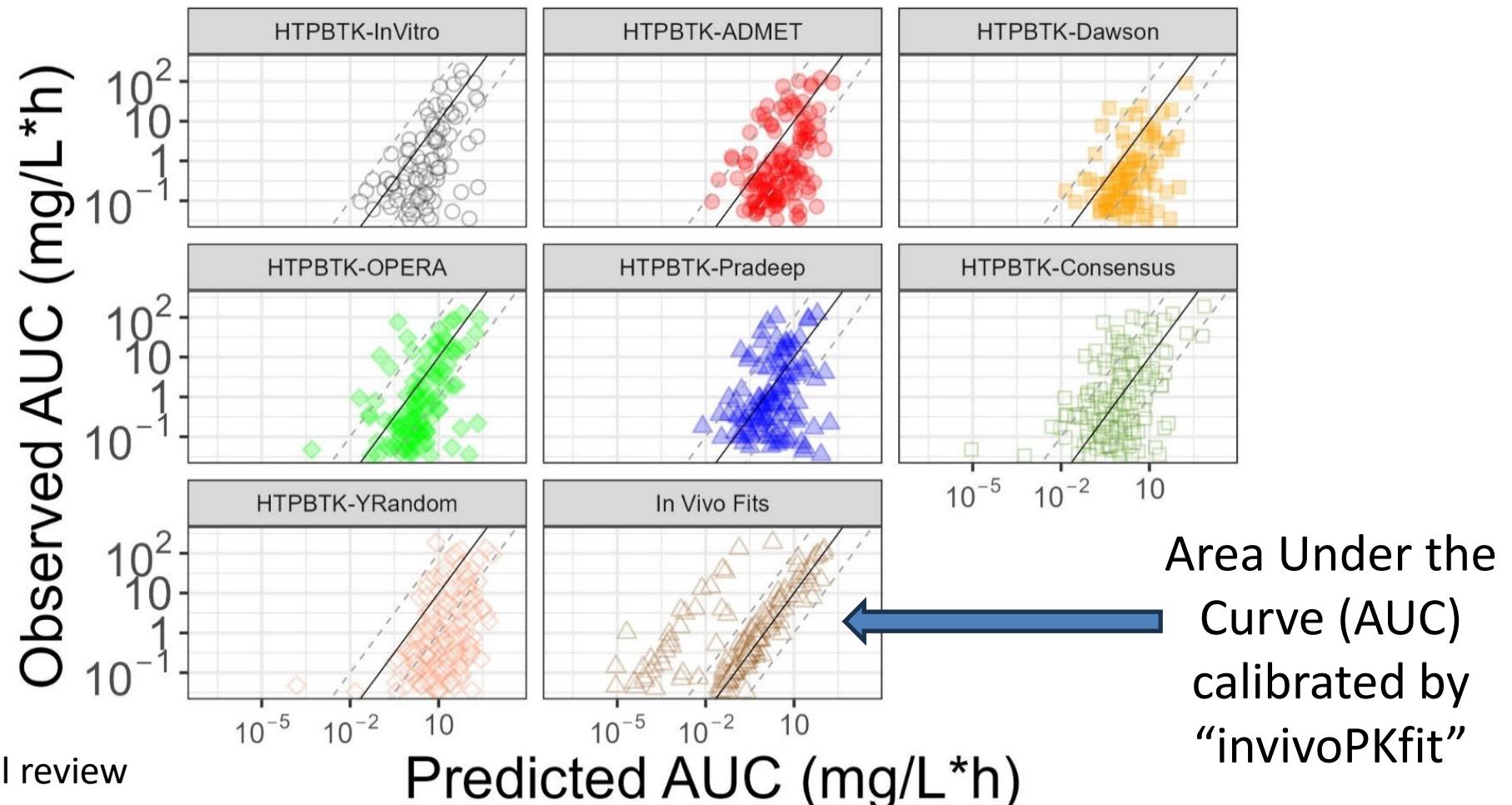
# HTTK Confidence Assessment

- How well does HTTK predict chemical absorption, distribution, metabolism, and excretion?
- Toxicokinetic Concentration vs. Time database (CvTdb) contains structured TK data from the peer-reviewed literature (Sayre et al., 2020)
  - EPA actively curating data from publications
  - Collaborators including Showa Pharmaceutical also providing data
- CvTdb data can be analyzed with open-source tool “invivoPKfit” to estimate TK statistics such as half-life and volume of distribution (Padilla Mercado et al., in preparation)



# HTTK Confidence Assessment

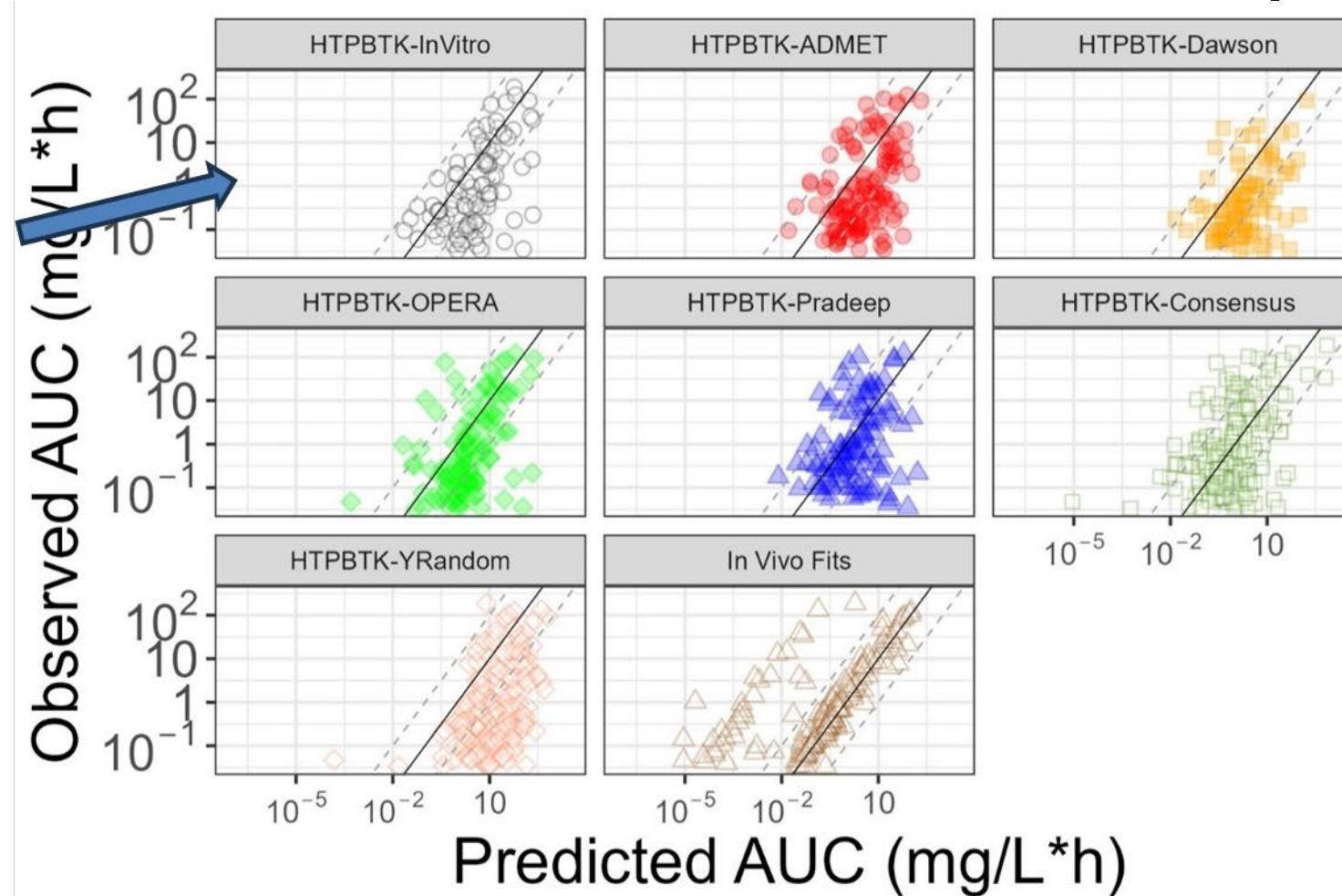
**CvTdb combined with “invivoPKfit” allows evaluation of HTTK predictions**



# HTTK Confidence Assessment

## CvTdb combined with “invivoPKfit” allows evaluation of HTTK predictions

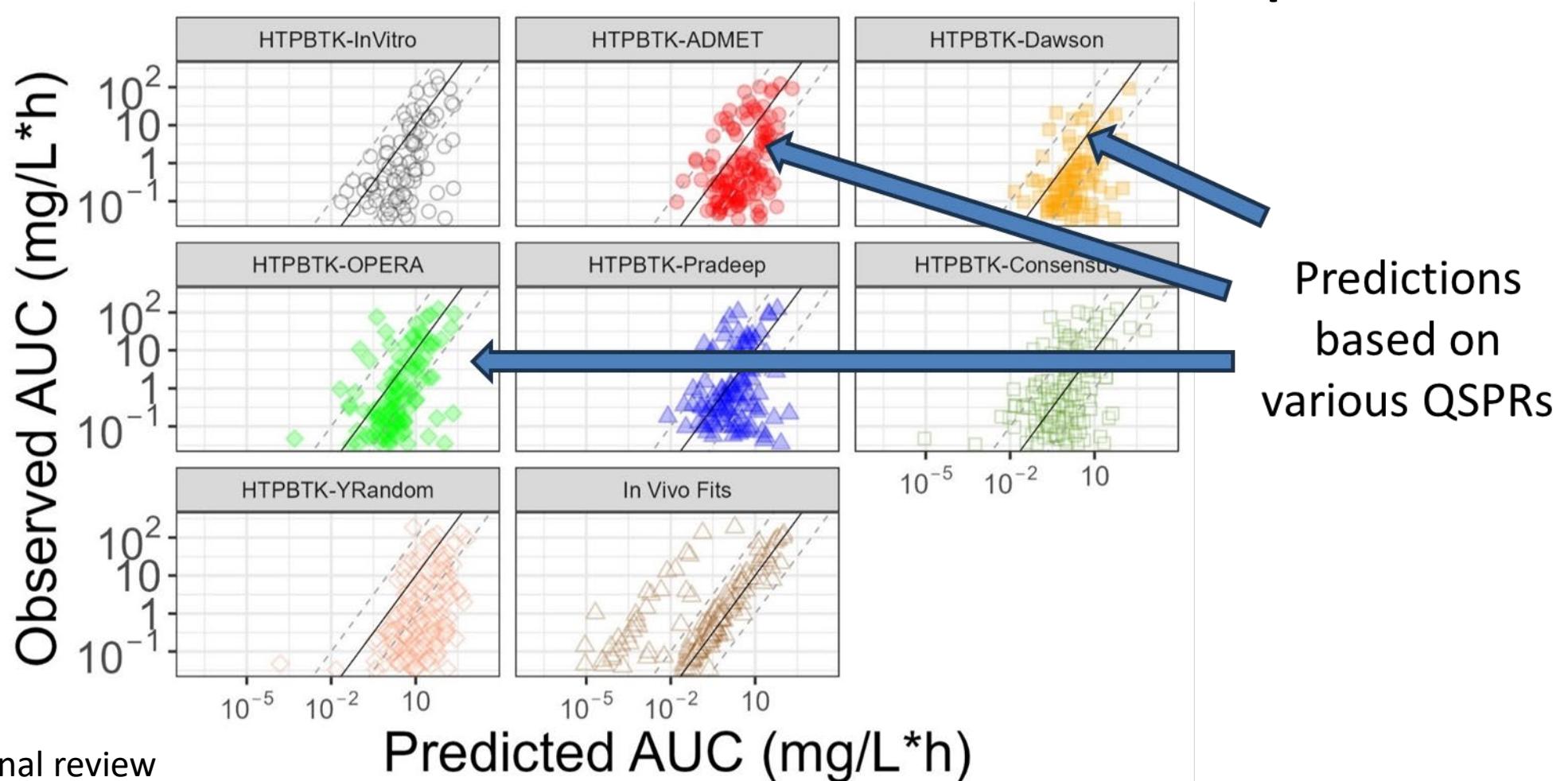
## Predictions based on *in vitro* TK measurements



Manuscript under internal review

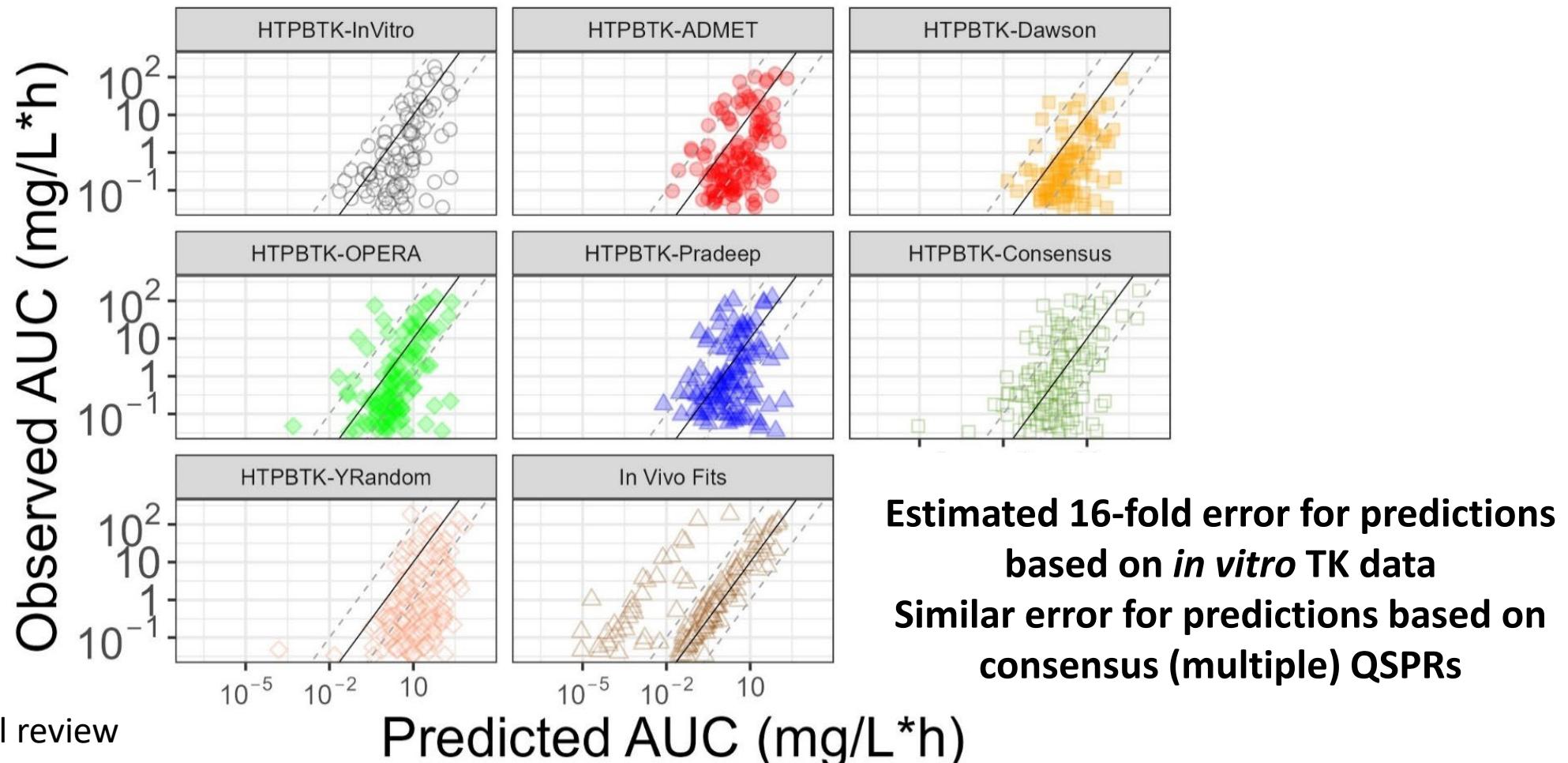
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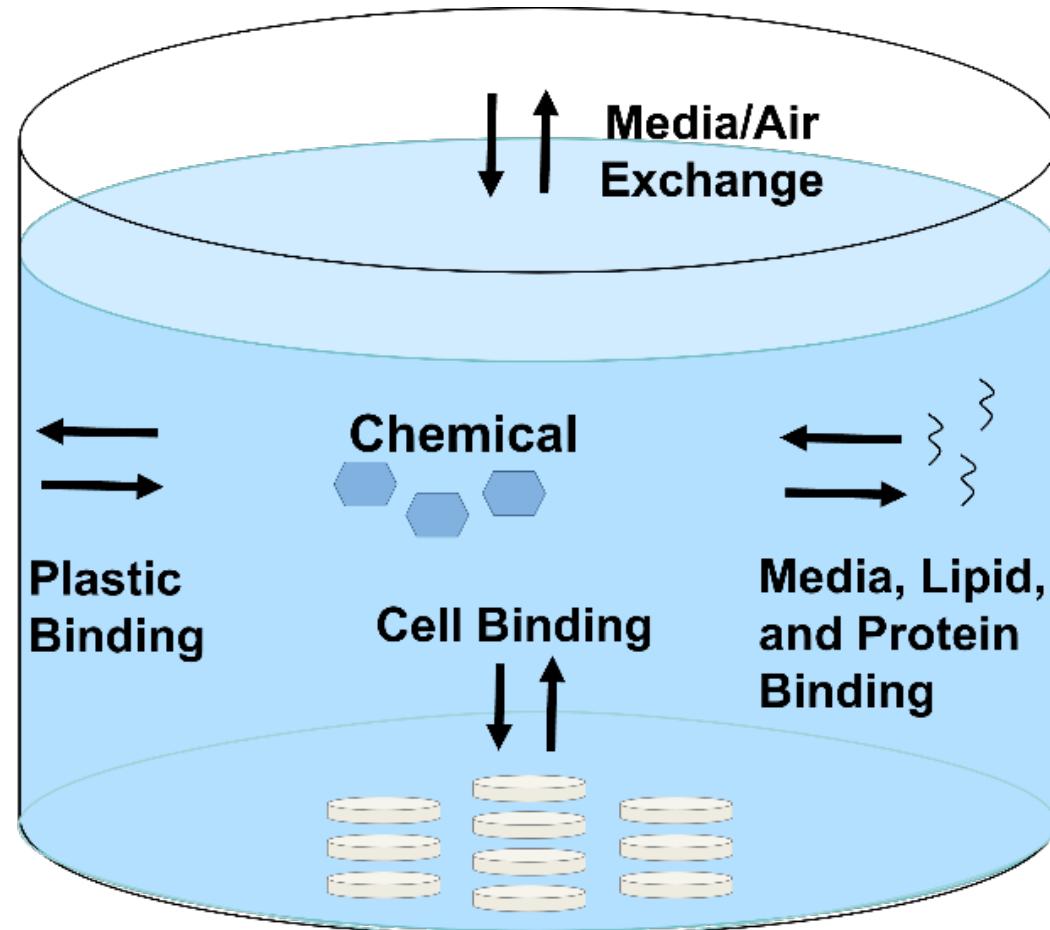
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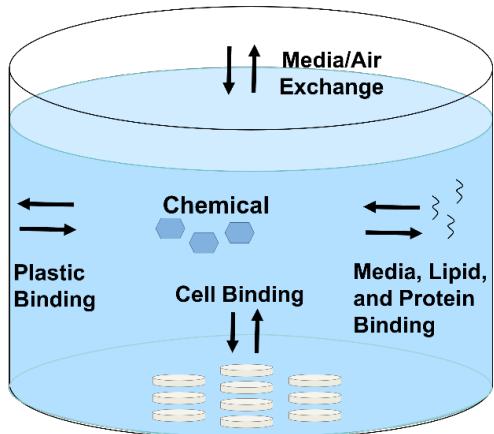
# In Vitro Distribution

- “httk” tool includes modified\* Armitage et al. (2014) model for estimating *in vitro* distribution
- Nominal tested concentration does not equal concentration in the cells

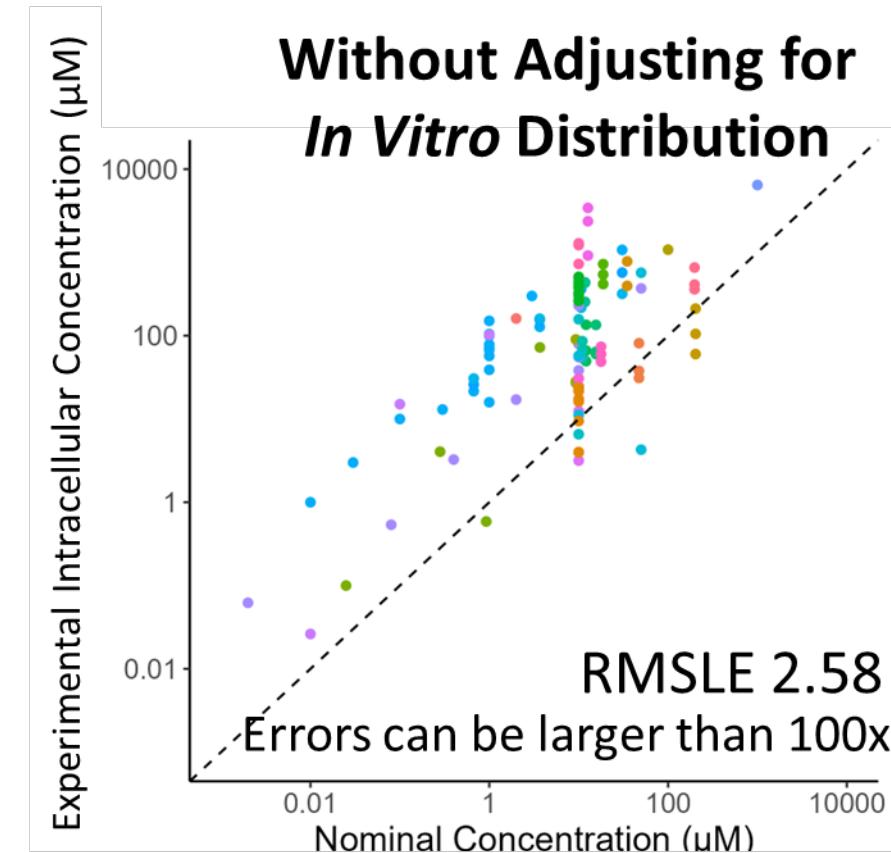


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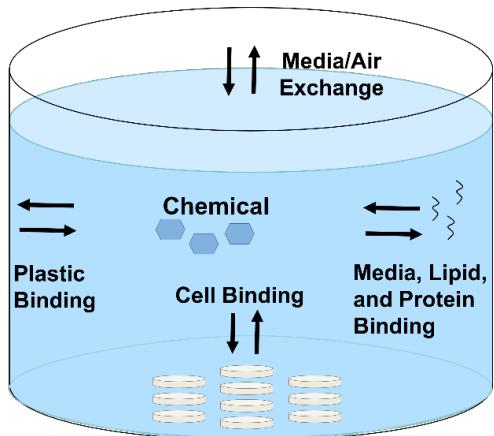


Chemical

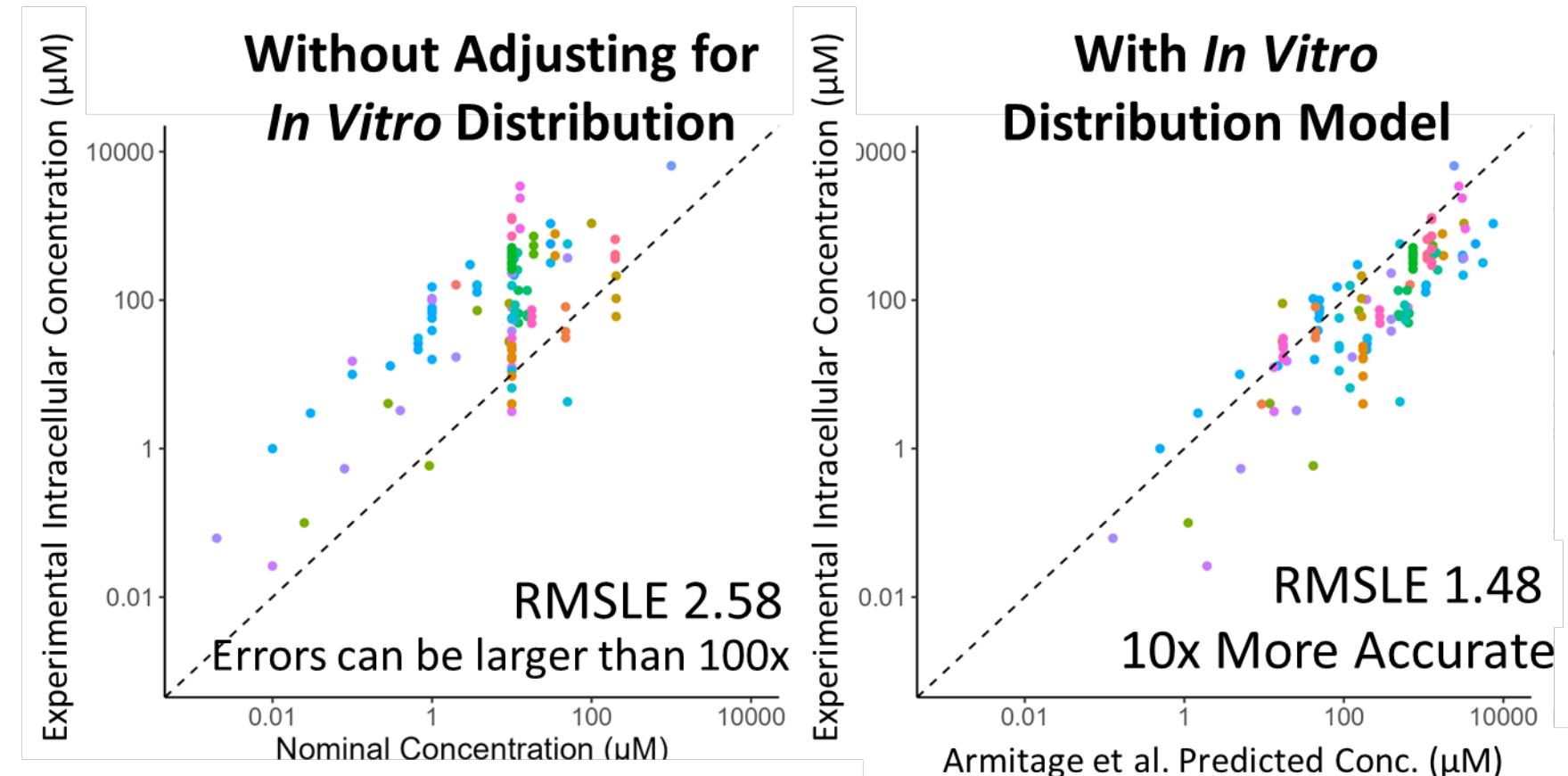
• 3-Methylcholanthrene	• Colchicine	• Ketoconazole	• PBDE 99	• Thiacloprid
• Acetaminophen	• Cyclosporine A	• Methyltestosterone	• Phenobarbital	• Trenbolone
• Atrazine	• Fenarimol	• N-Phenyl-1,4-benzenediamine	• Rifampicin	• Triphenyl phosphate
• BPA	• Flusilazole	• Omeprazole	• Ritonavir	• Warfarin
• Caffeine	• Flutamide	• PBDE 153	• Rosiglitazone	
• Chenodeoxycholic Acid	• Genistein	• PBDE 47	• Tamoxifen	

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Meredith Scherer, Katie Paul Friedman



Chemical	3-Methylcholanthrene	Acetaminophen	Atrazine	BPA	Caffeine	Chenodeoxycholic Acid	Colchicine	Cyclosporine A	Fenarimol	Flusilazole	Flutamide	Genistein	Ketoconazole	Methyltestosterone	N-Phenyl-1,4-benzenediamine	Omeprazole	PBDE 153	PBDE 47	PBDE 99	Phenobarbital	Rifampicin	Ritonavir	Rosiglitazone	Tamoxifen	Thiacloprid	Trenbolone	Triphenyl phosphate	Warfarin
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# New Models for HTTK

- HTTK is the combination of *in vitro* TK data and high throughput physiologically-based toxicokinetic (HT-PBTK) models

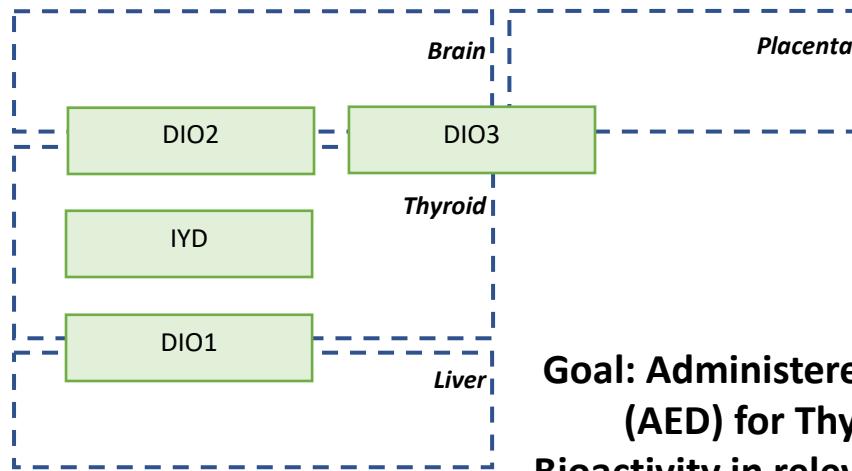
**A recent manuscript provides a guide to developing new HT-PBTK models using existing HTTK data and methods (Davidson-Fritz et al., pre-print)**

Multiple new models are in development including:

- Full human gestational (Truong and Paul Friedman)
- Chemical mixtures (Schacht and Evans)
- PFAS (Wetmore and Tornero-Velez)
- Dermal Exposure (Meade and Evans)
- Inhalation Steady-State (Ring and Schacht)
- Blood-Brain Barrier (Unnikrishnan, Chang, Sluka, Kreutz, Li...)

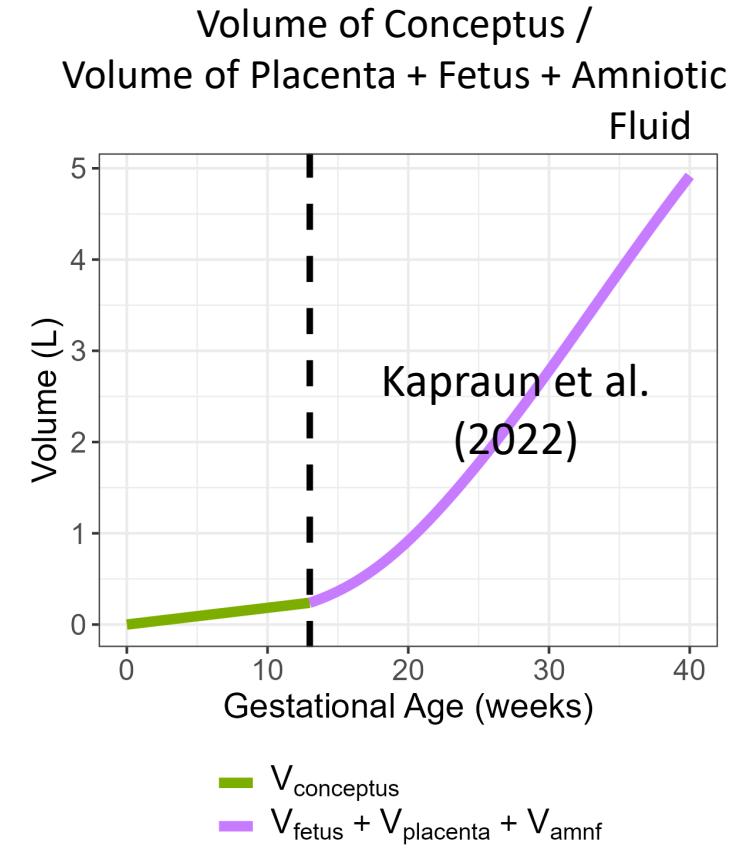
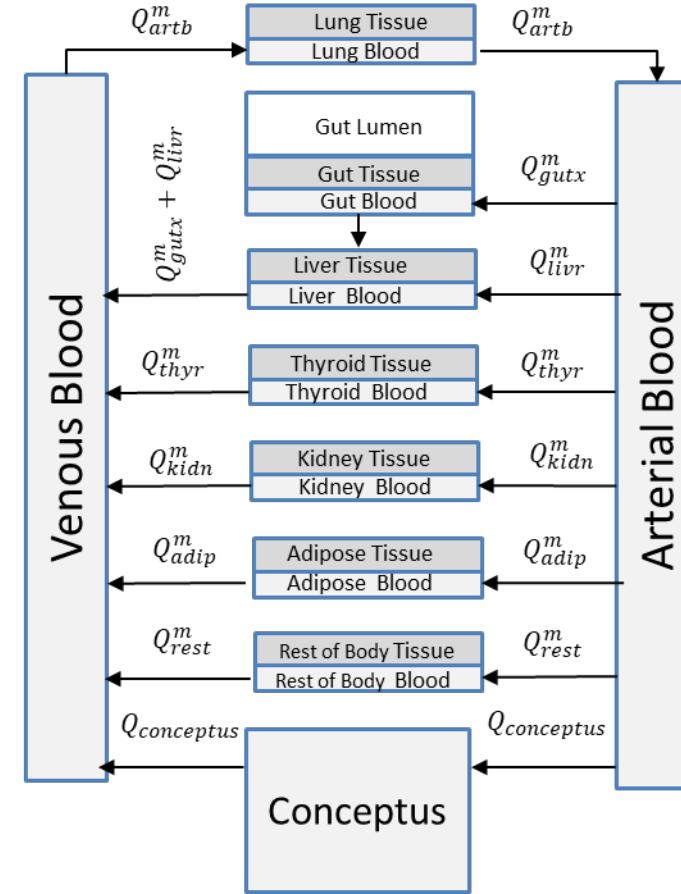
# Full Human Gestational IVIVE

- Kapraun et al. (2022) model describes human gestation in second and third trimesters
- Truong et al. have developed a new HT-PBTK model for full gestation
- Allows IVIVE for thyroid bioactivity during the human perinatal period



**Goal: Administered Equivalent Dose (AED) for Thyroid-Relevant Bioactivity in relevant compartments and lifestages of concern**

Maternal



Kimberly Truong, Dustin Kapraun, Katie Paul Friedman  
 Manuscript under internal review

# HTTK for PFAS

- Typical *in vitro* TK measurements do not capture the role of transporters that may be important for understanding per- and polyfluorinated alkyl substances (PFAS)
- Dawson et al. (2023) machine learning model approximates the impact of transporters on toxicokinetics
- New PFAS-specific *in vitro* TK measurements from Wetmore lab and NTP were recently published (Smeltz, et al. 2023, Kreutz, et al. 2023, Crizer et al. 2024)
- New HT-PBTK models specifically for PFAS allow prediction of TK and IVIVE, including interspecies differences

PFAS	Wallis et al. (2023) Observed Half-Life (Days)	Dawson et al. (2023) Predicted Half-Life (Days)
DTXSID30892354	397-1980	>33
DTXSID90723993	72.9-340	>33
DTXSID50723994	287-1220	>33
DTXSID80515849	8.80-111	>33
DTXSID20892348	13-133	<7

# Inhalation IVIVE

New IVIVE model including inhalation and exhalation allows estimation of inhalation equivalent doses in ppm

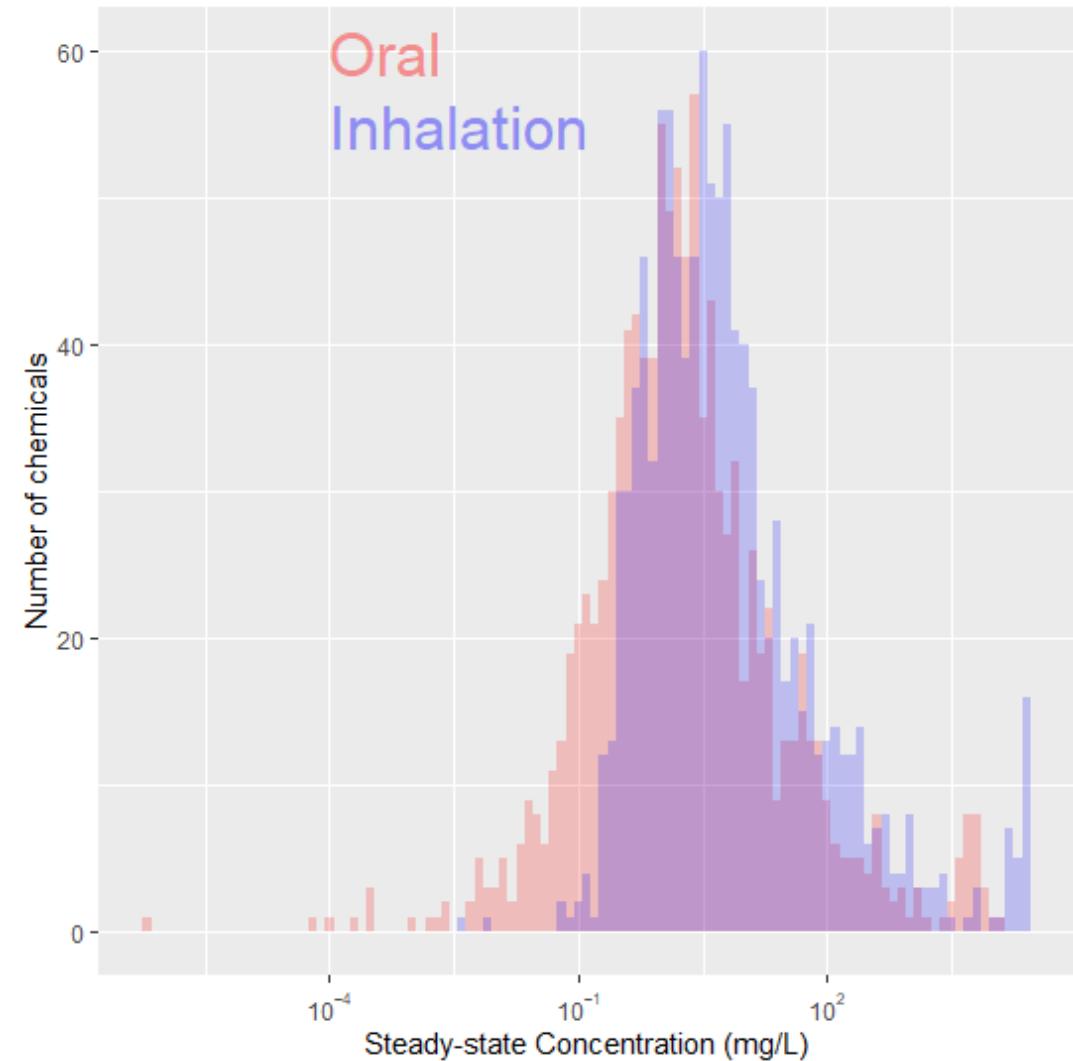
*In Vitro*  
 Point of Departure

Administered Equivalent Dose

$$\mathbf{AED}_{95} = \frac{[X]}{C_{ss,95,unit}}$$

Inhalation Equivalent Dose

$$\mathbf{IED}_{95} = \frac{[X]}{C_{ss,95,1ppm}}$$

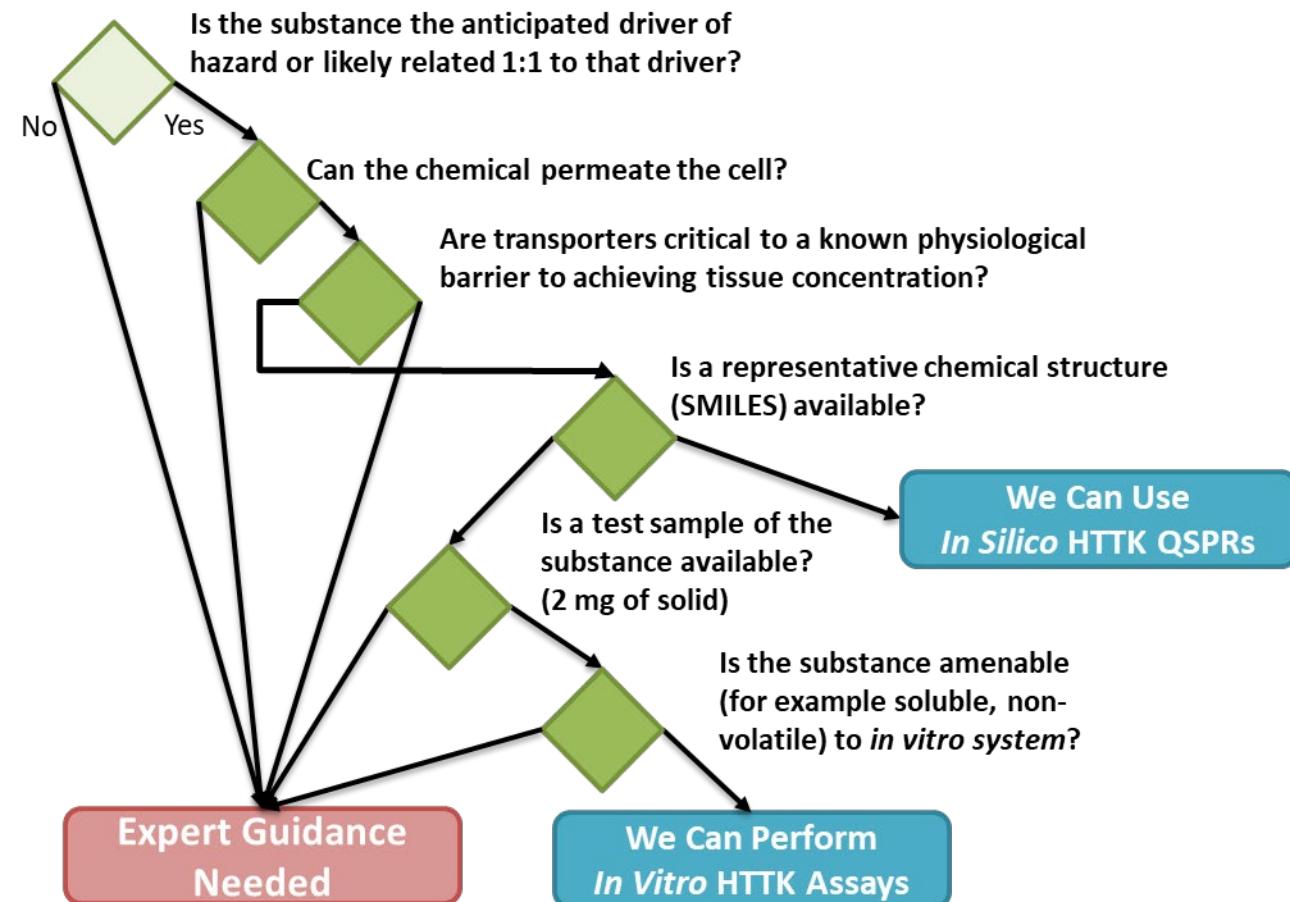


Celia Schacht, Caroline Ring  
 Manuscript under internal review

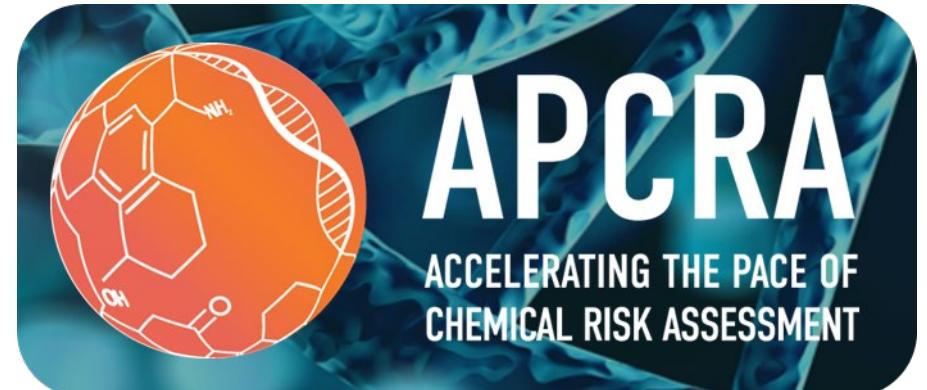
# Open-Source Tools

- R package “httk” allows for probabilistic *in vitro-in vivo* extrapolation (IVIVE) and toxicokinetics (TK)  
*Pearce et al. (2017)*
  - Simulates human variability and propagates measurement uncertainty
  - *In vitro* TK data for >1,000 chemicals and QSPR predictions for ~9,000 more
  - <https://CRAN.R-project.org/package=httk>
- Toxicokinetic Concentration vs. Time database (CvTdb) provides public, curated data with study annotation  
*Sayre et al. (2020)*
  - >250 analytes from hundreds of studies
  - <https://github.com/USEPA/CompTox-PK-CvTdb>
- R package “invivoPKfit” allows for consistent, reproducible TK parameter estimation from CvT data  
*Padilla Mercado et al. (2024)*
  - <https://github.com/USEPA/CompTox-ExpoCast-invivoPKfit>
- Forthcoming R package “invitroTKstats” allows for transparent estimation of *in vitro* TK parameters  
*Wambaugh et al. (2019)*
  - Calculates chemical-specific measurement uncertainty

# Using HTTK in Decision Making



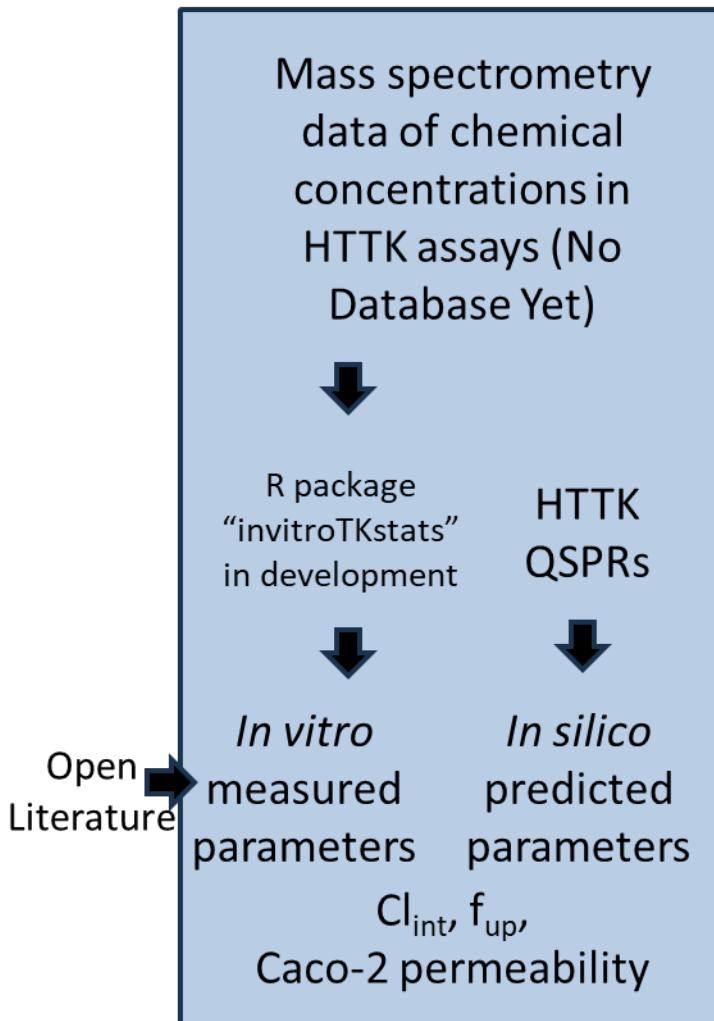
- Accelerating the Pace of Chemical Risk Assessment (APCRA) international government collaboration is developing decision trees to guide consideration of using HTTK in decision making



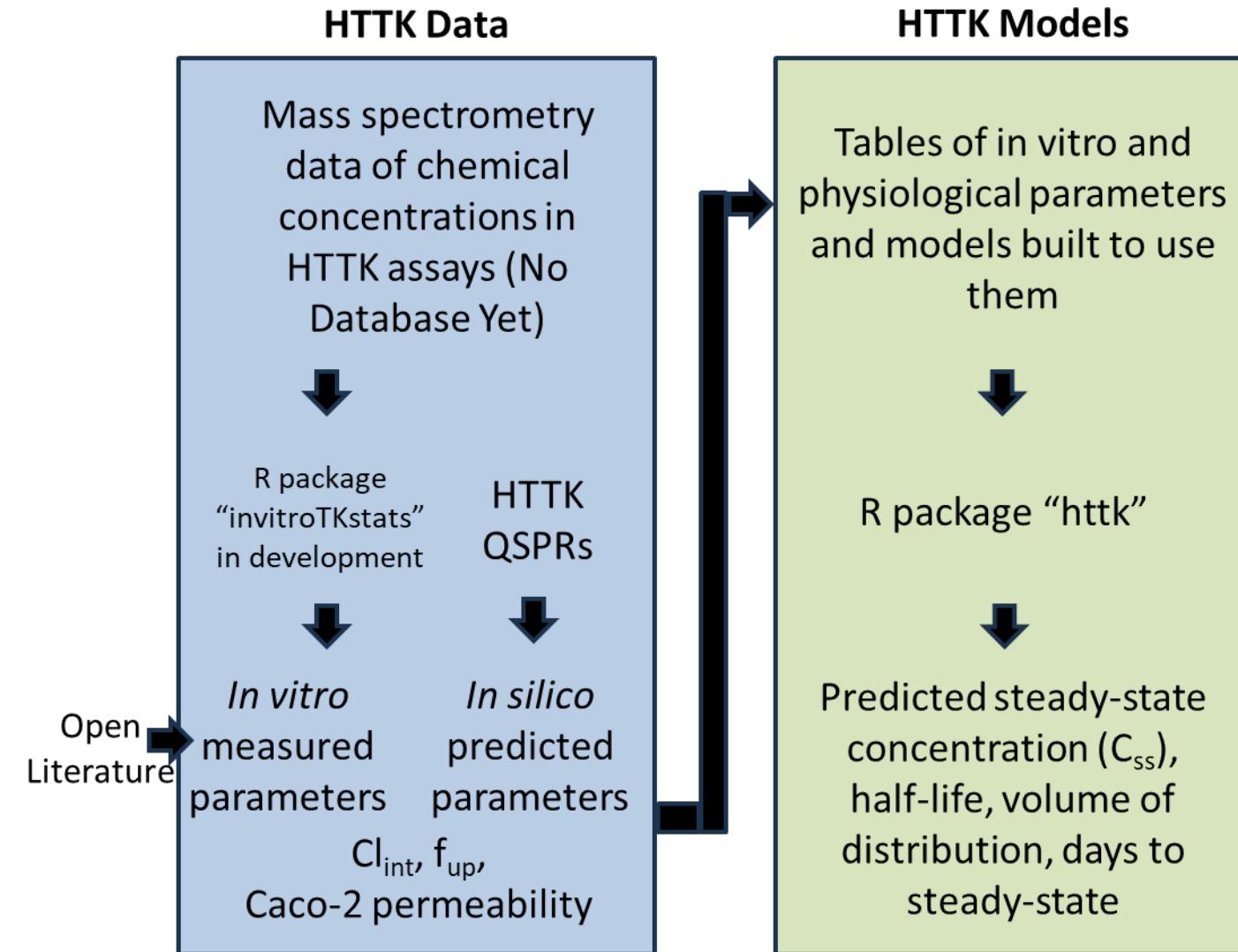
# EPA HTTK Research

APCRA workgroup  
HTTK in  
decision making

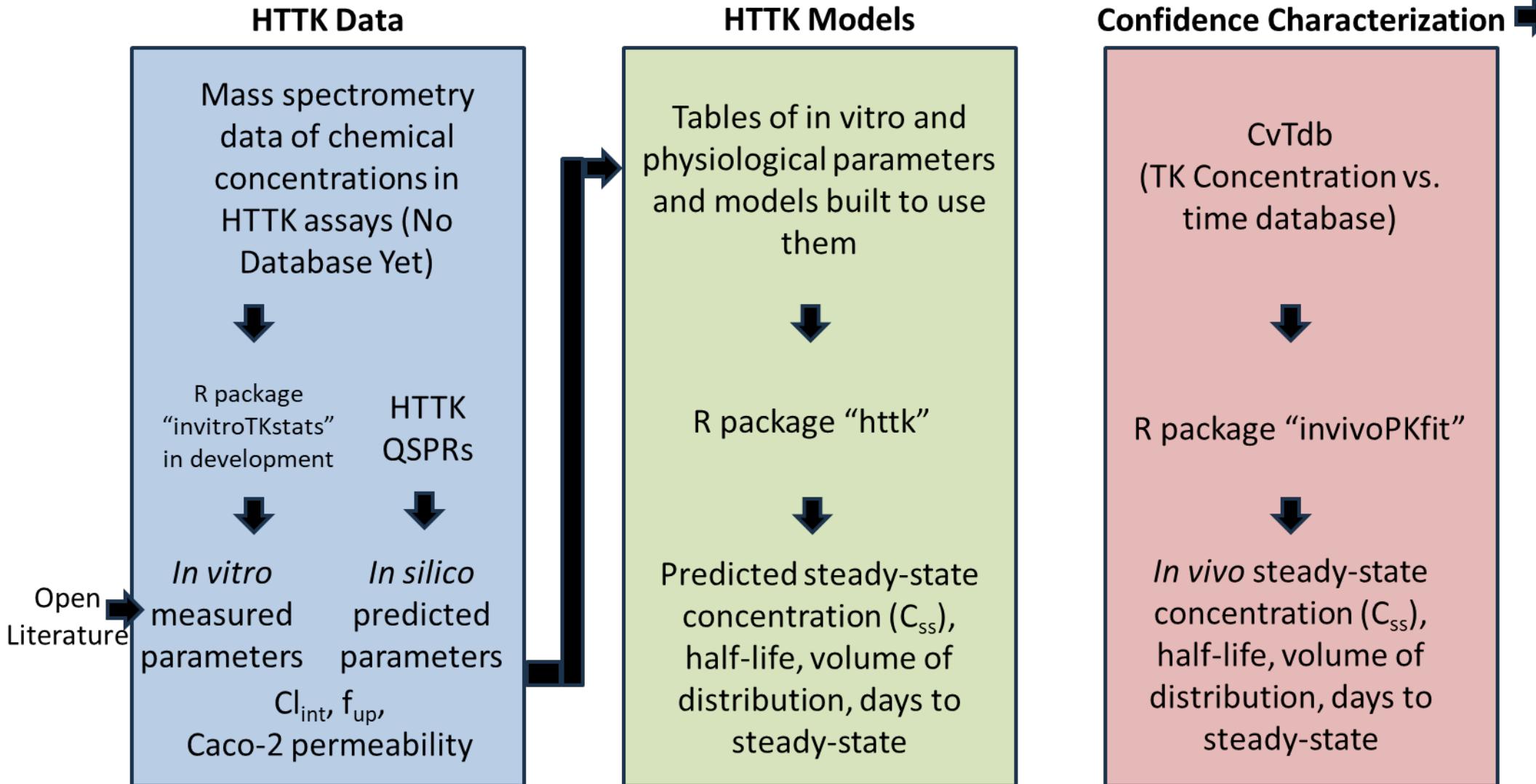
## HTTK Data



# EPA HTTK Research

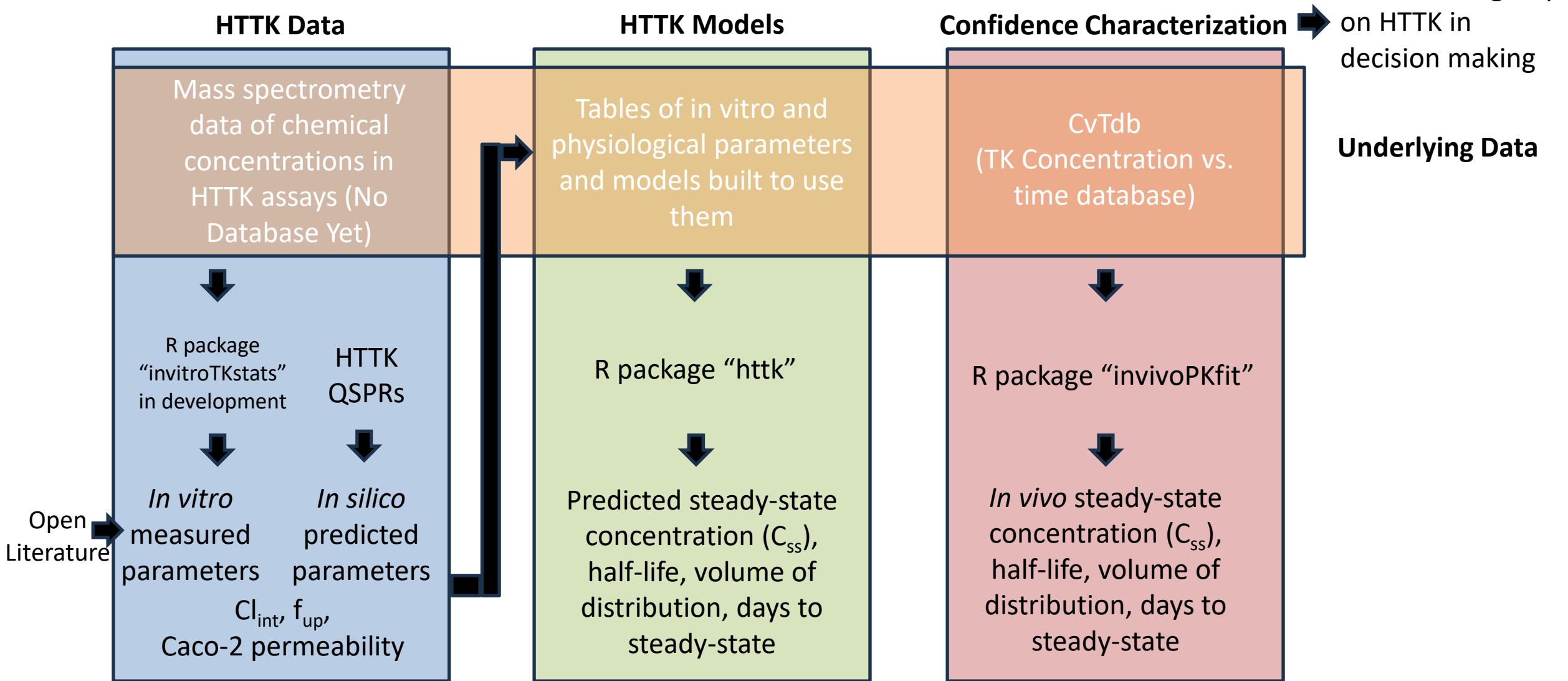


# EPA HTTK Research

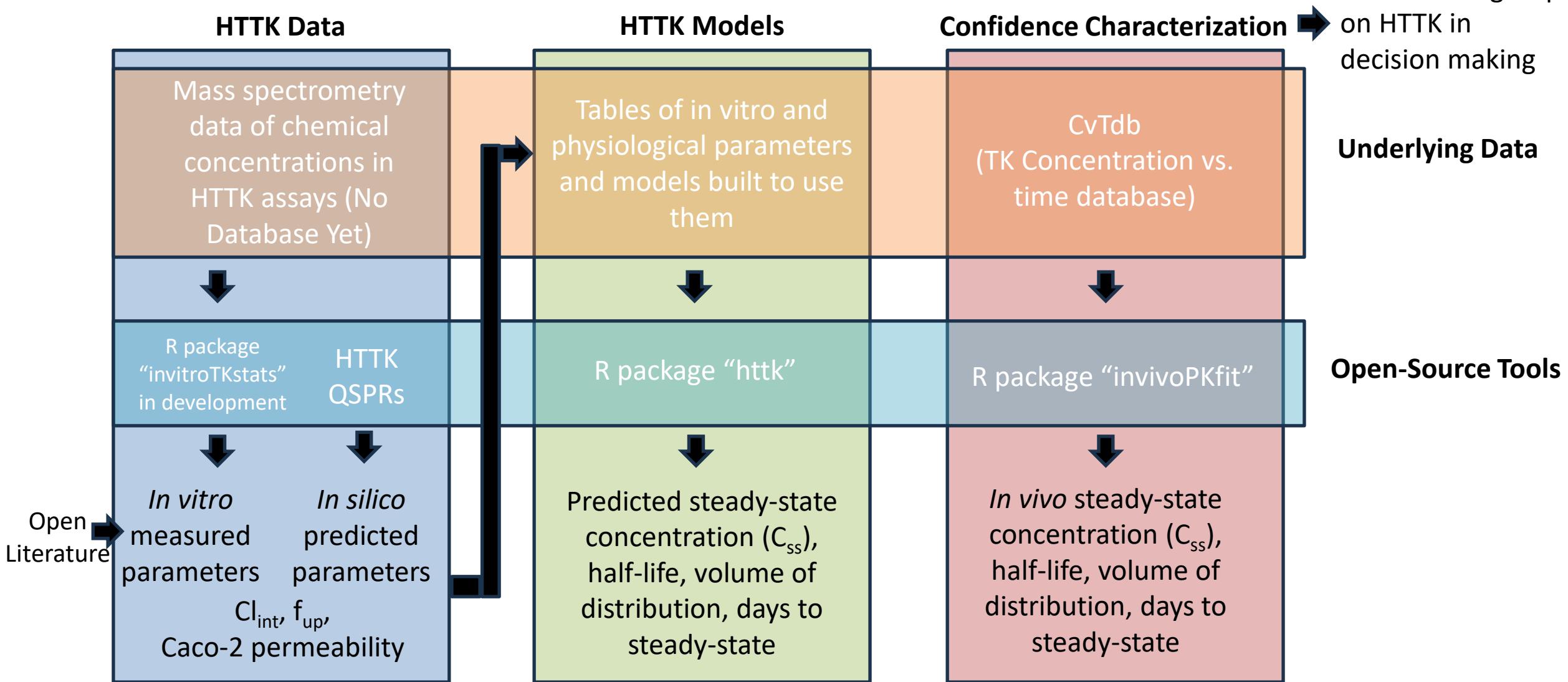


APCRA workgroup  
on HTTK in  
decision making

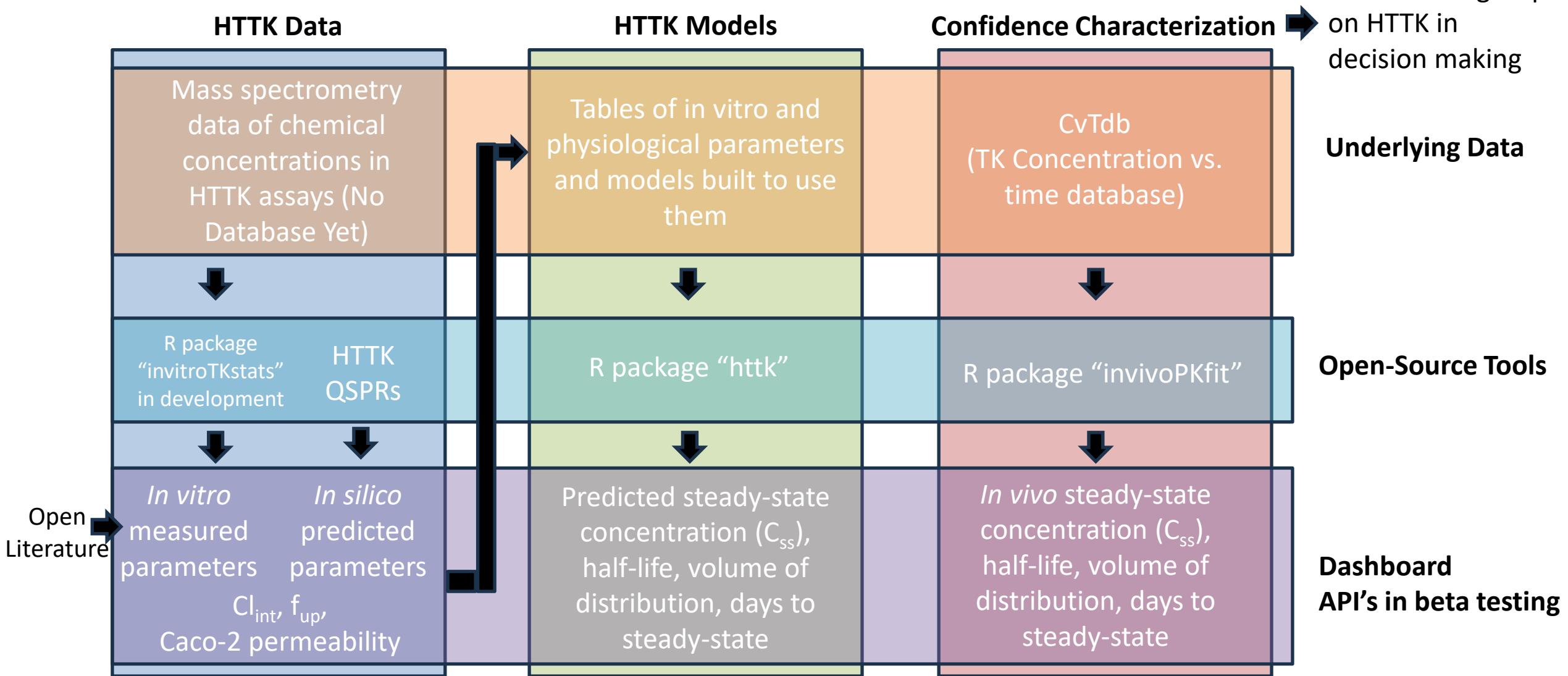
# EPA HTTK Research



# EPA HTTK Research

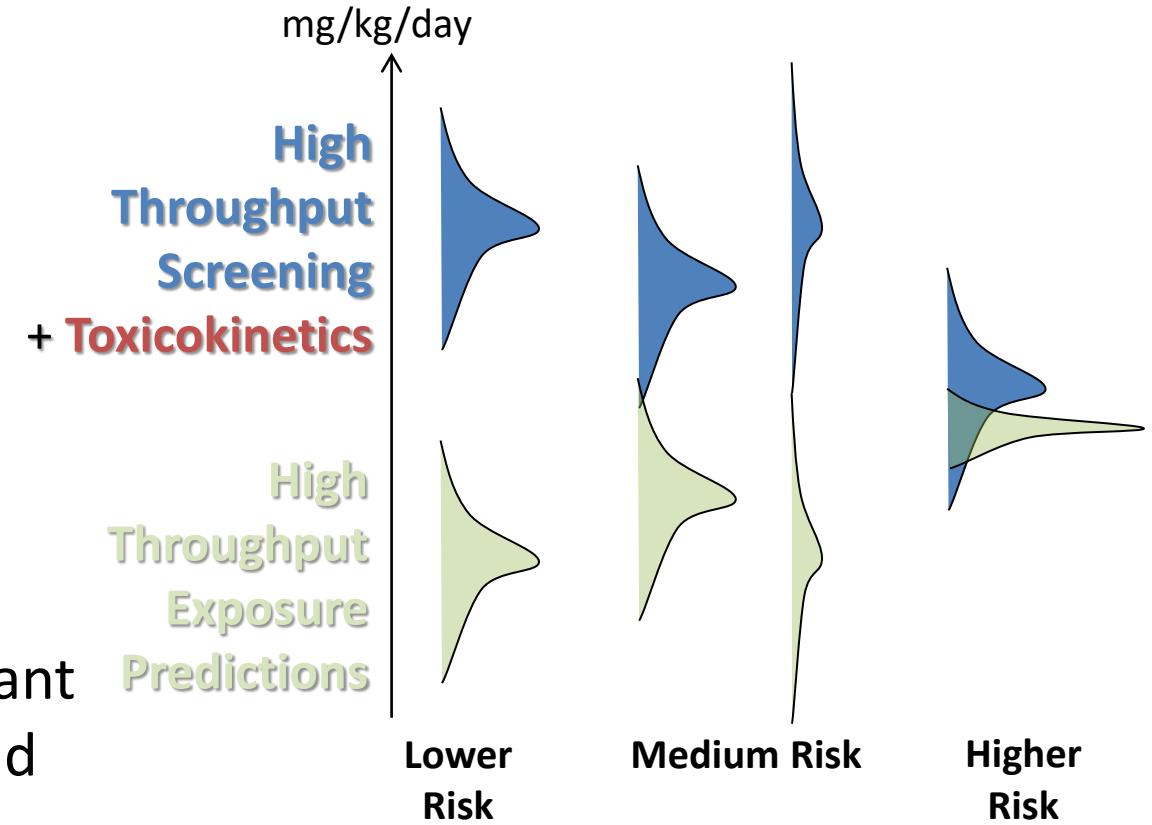


# EPA HTTK Research

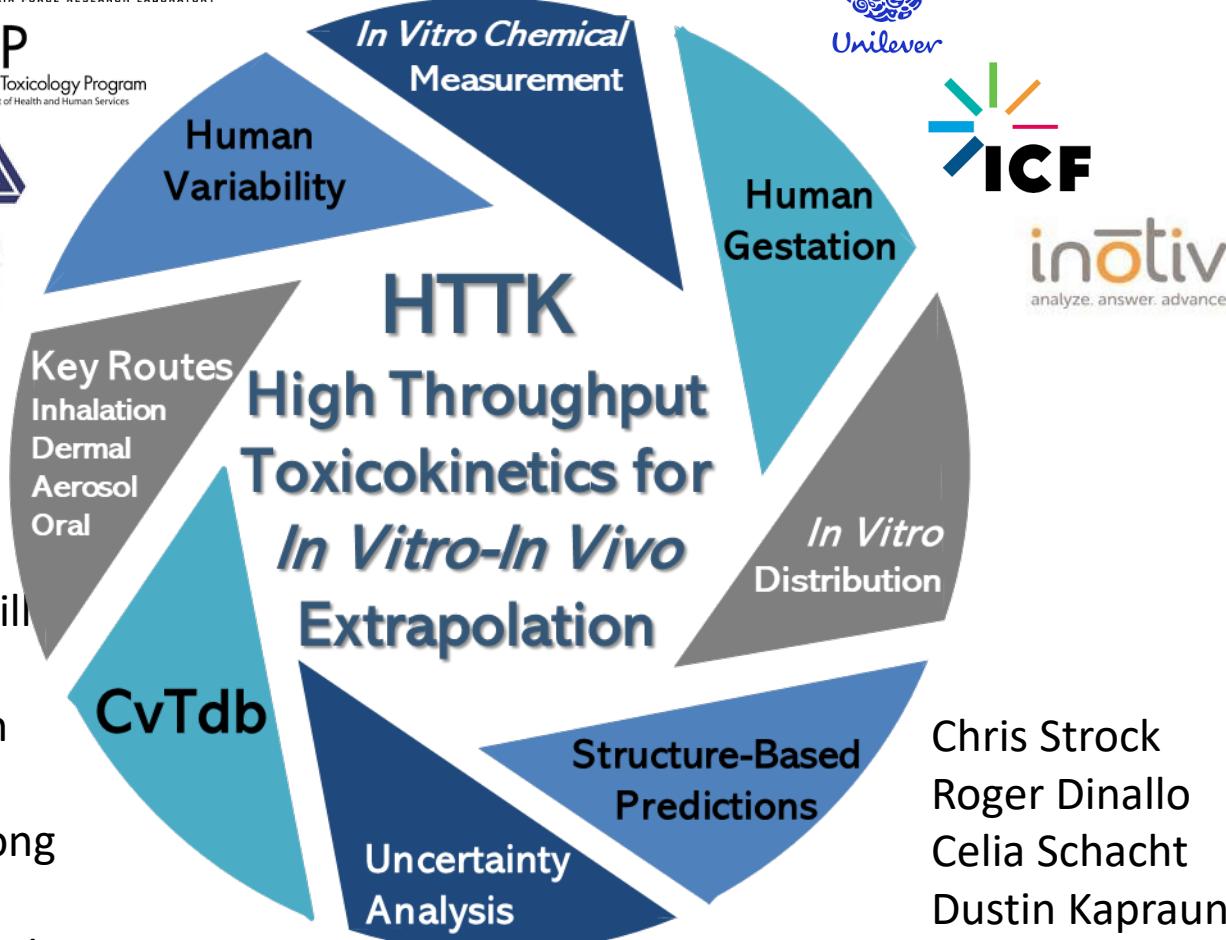


# Conclusions

- HTTK is needed to convert from bioactive *in vitro* concentrations to putative dose rates needed to produce those concentrations in the body
- HTTK allows rapid calculations for a variety of scenarios
- HTTK is being expanded to better cover relevant chemicals (volatiles, PFAS) and susceptible and highly exposed populations (pregnancy, occupational)
- HTTK resources are widely available on-line



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# References

Please send questions and slide requests to [wambaugh.john@epa.gov](mailto:wambaugh.john@epa.gov)

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**The views expressed in this presentation are those of the author and do not necessarily reflect the views or policies of the U.S. EPA**

# Translation of *In Vitro* Effect Concentrations: Equilibrium or Kinetic Distribution Models?

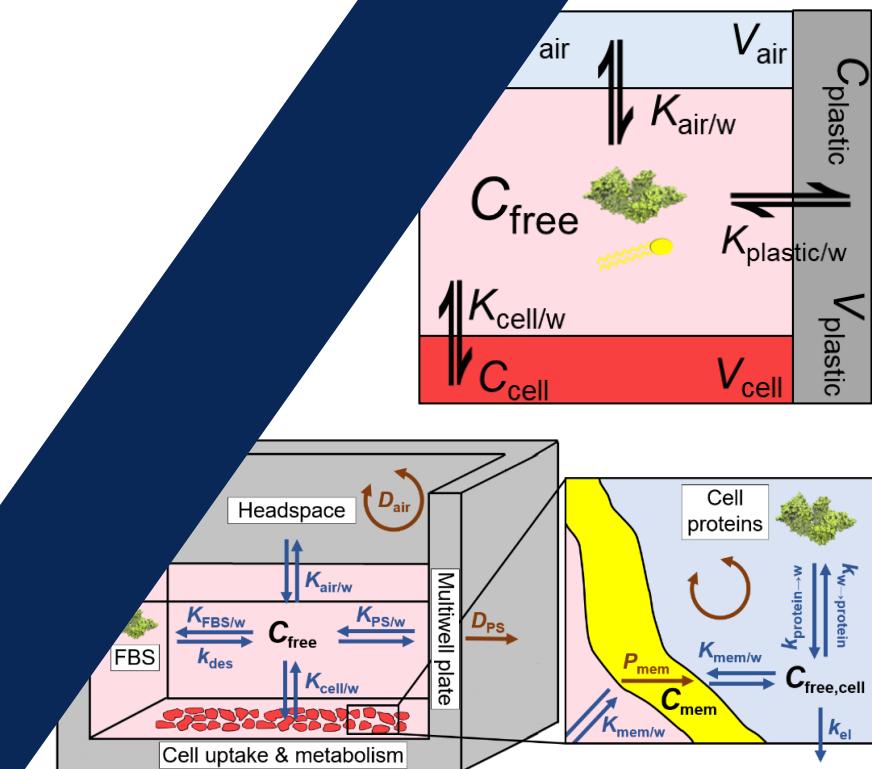
Fabian C. Fischer

Cedric Abele, Luise Henneberger, Rita Schlichting, Maria König, Beate Escher, **and many more...**

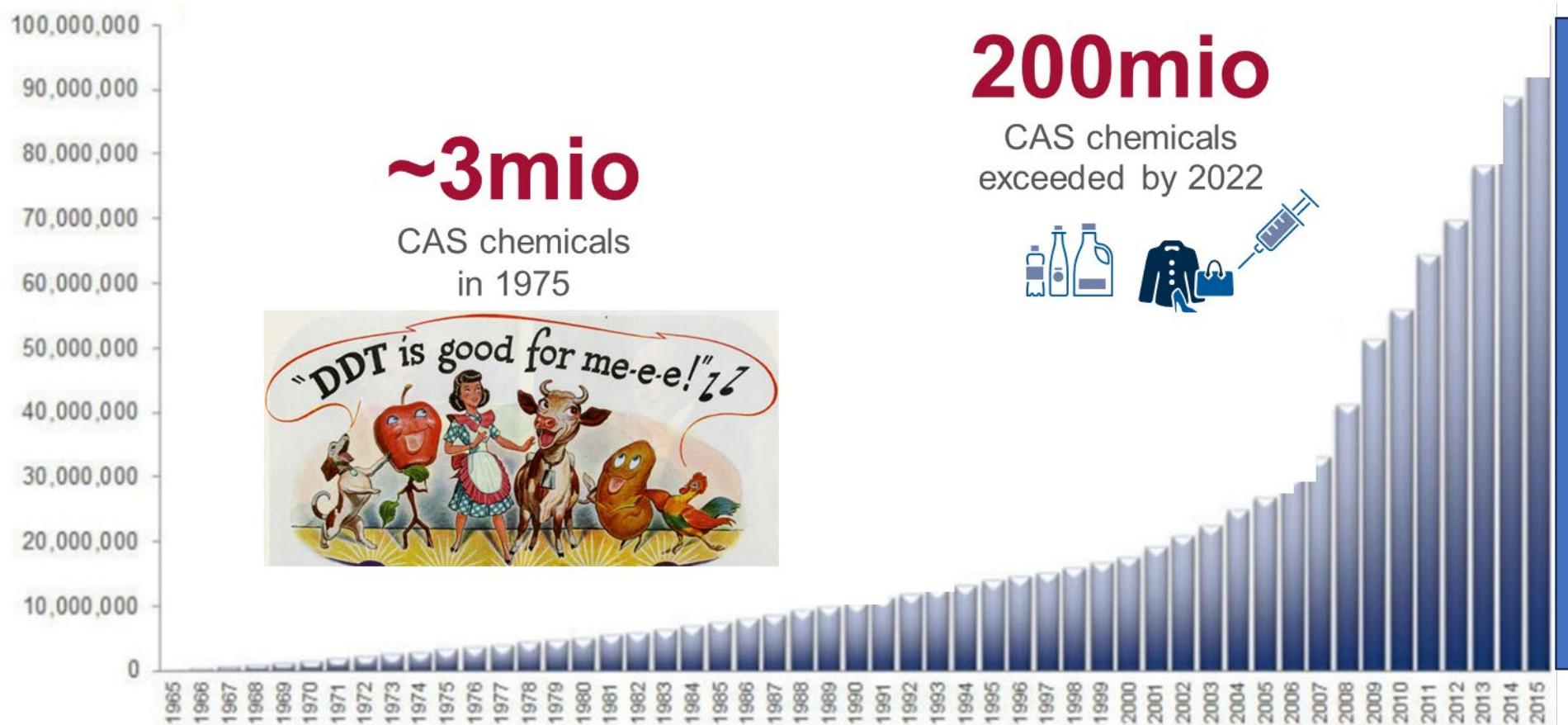


EPA 4th NAMs Conference  
November 6<sup>th</sup>, 2024

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# The use of anthropogenic organic chemicals has changed over the last decades

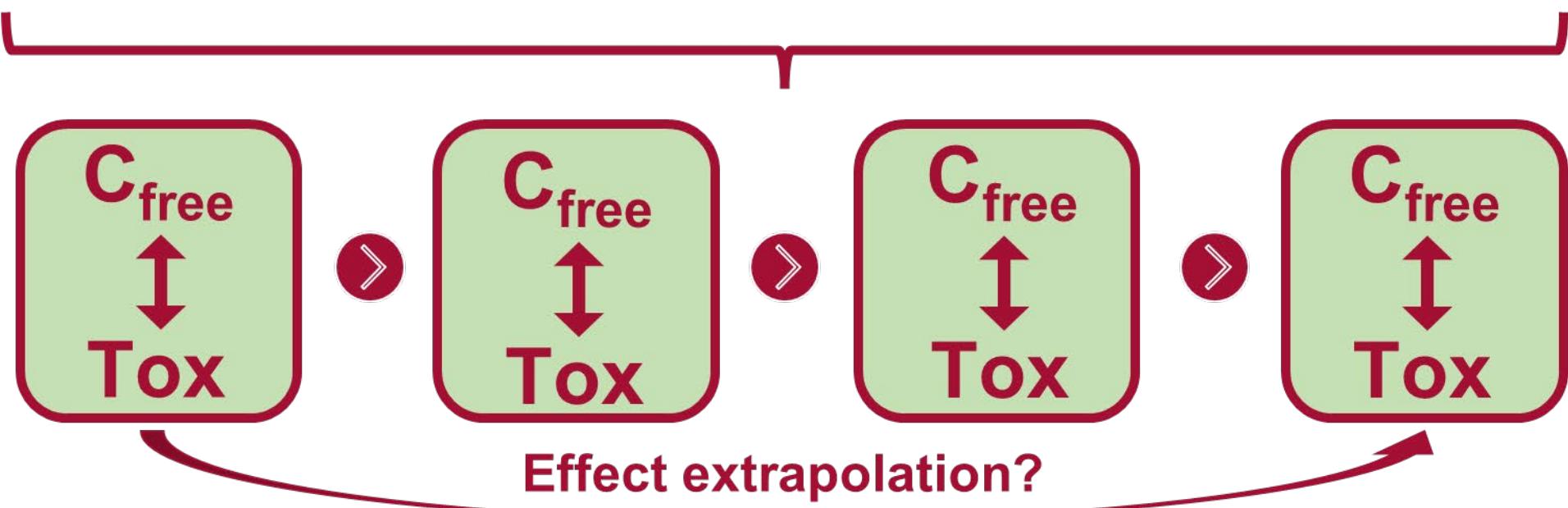
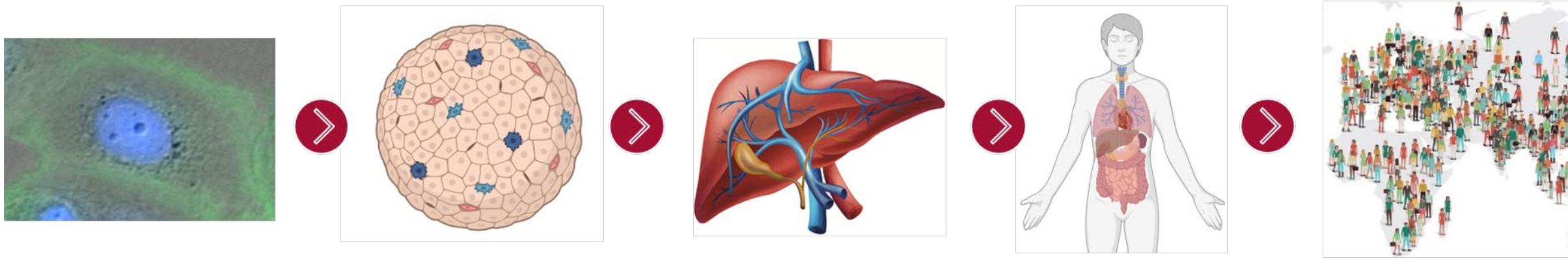


- Not only the number increased, so did the **diversity in chemical structures!**
- Global release into environment: **310 kg** of toxic chemicals **per second!**<sup>[1]</sup>
- **One in every six** children suffers from neurodevelopmental abnormality suspected to be triggered by exposure to environmental chemicals!<sup>[2]</sup>

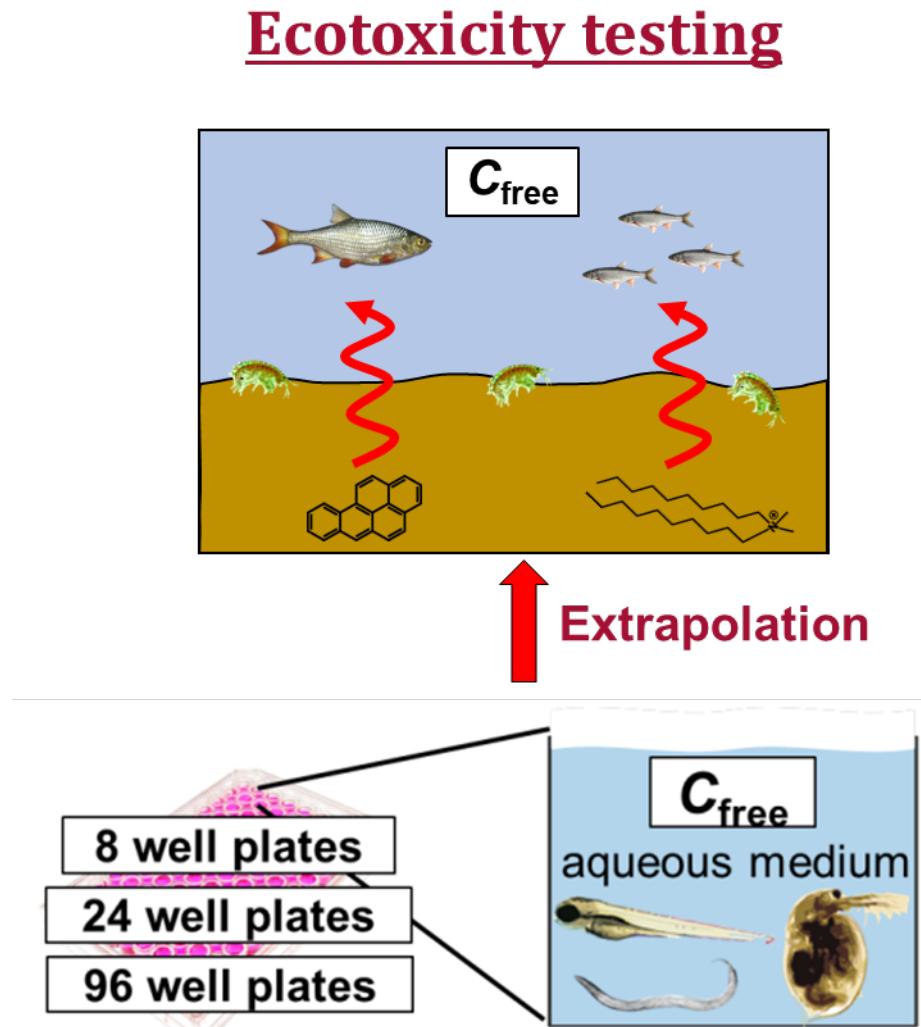
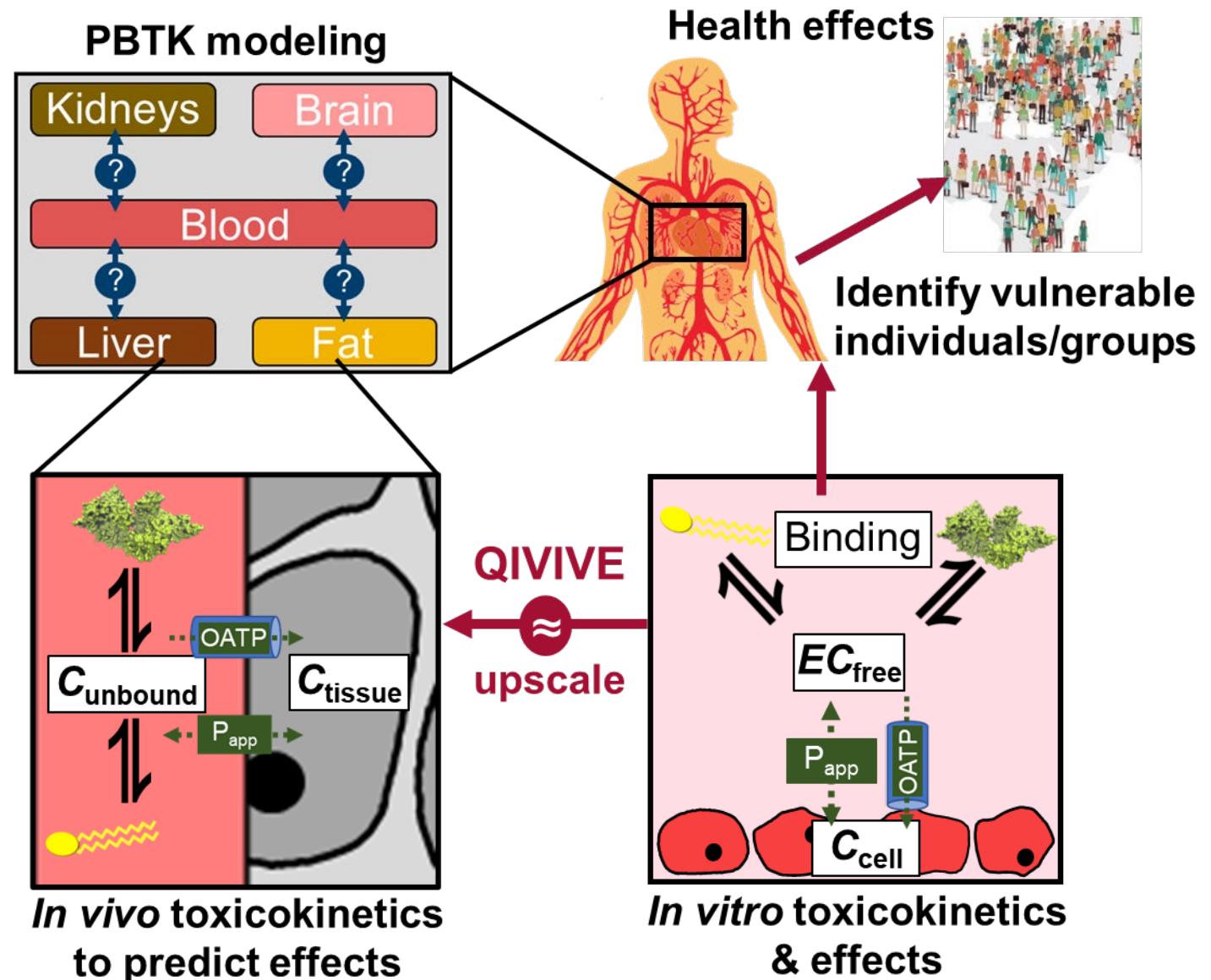
[1] <https://www.worldometers.info/>  
[2] <https://braindrain.dk/>

# Explore chemical exposures and toxicity based on freely dissolved concentrations

$C_{\text{free}} = C_{\text{unbound}}$  in pharmaceutical sciences



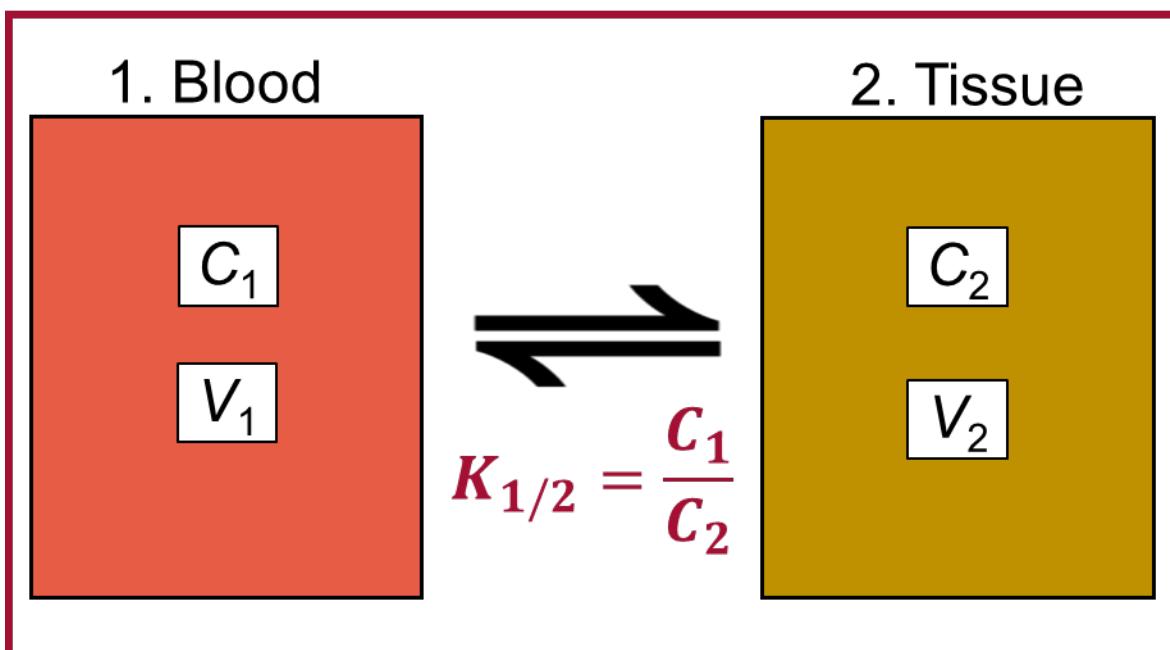
# Quantitative *in vitro*-*in vivo* extrapolation (QIVIVE)



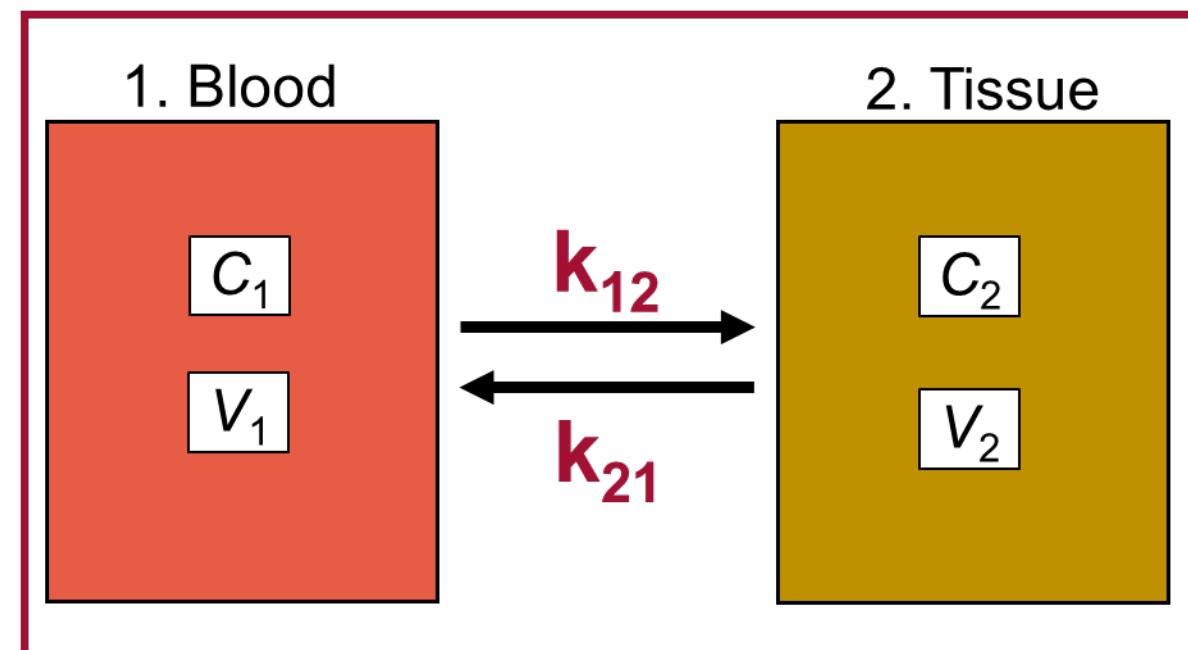
# Chemical transport models to investigate the distribution of chemicals in *in vitro* cell assays

- ? Can the system be approximated with an equilibrium or kinetic model?
- ? How much complexity is necessary to predict/explain exposure?

Equilibrium mass balance model



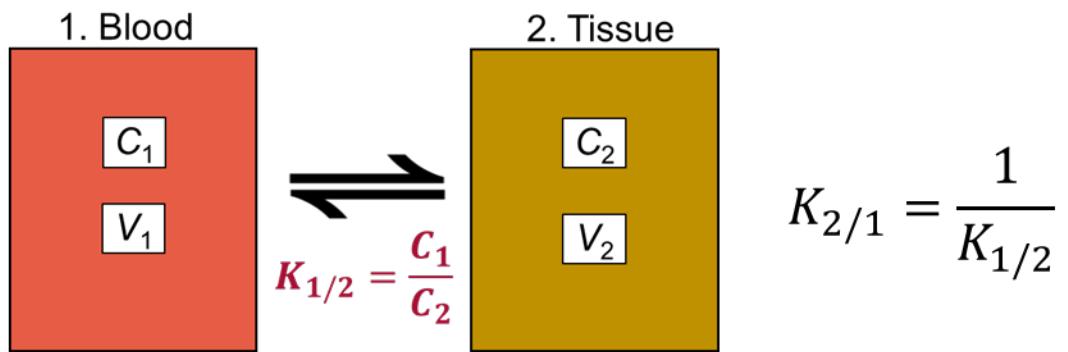
Kinetic model (rate-limited)



Can we simplify?

# Computing chemical transport models: The benefit of simplicity

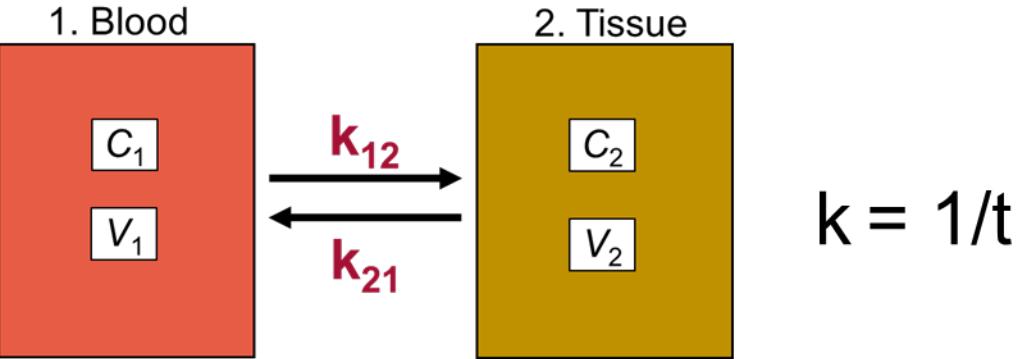
## Equilibrium mass balance model



$$f_1 = \frac{1}{\left(1 + K_{2/1} \cdot \frac{V_2}{V_1}\right)} \quad > \quad C_1 = \frac{f_1 \cdot n_{\text{total}}}{V_1}$$

$$\text{Multi-box: } f_1 = \frac{1}{\left(1 + K_{2/1} \cdot \frac{V_2}{V_1} + K_{3/1} \cdot \frac{V_3}{V_1} + \dots\right)}$$

## Kinetic model (rate-limited)



$$\frac{dC_1}{dt} = -k_{12} \cdot C_1 + k_{21} \cdot C_2$$

$$> C_1(t) = C_1(t_{-1}) - k_{12} \cdot C_1(t_{-1}) + k_{21} \cdot C_2(t_{-1})$$

Multi-box becomes a bit more complex...

# Computing chemical transport models: Implementation of numerical models in *R*



Solve the equations in **discrete time steps** – increase the number of time steps ( $\Delta t$ ) to increase the model accuracy and to avoid numerical errors

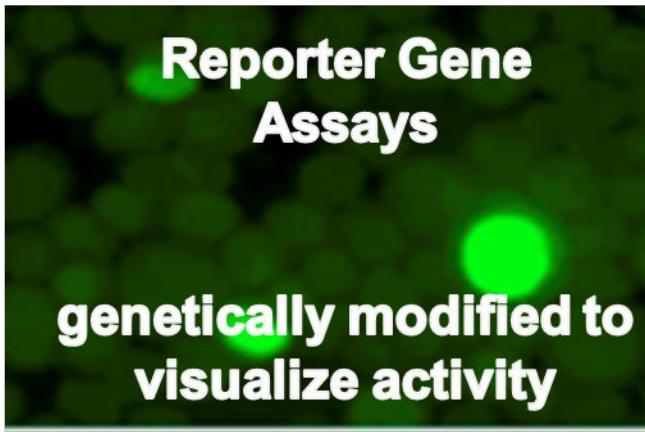
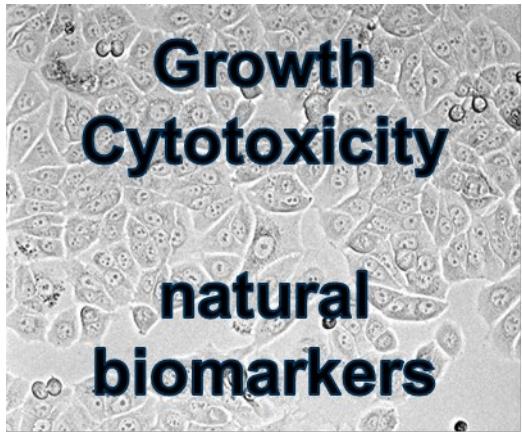
$$C_1(t) = C_1(t_{-1}) - k_{12} \cdot C_1(t_{-1}) \cdot \Delta t + k_{21} \cdot C_2(t_{-1}) \cdot \Delta t$$

$$C_2(t) = C_2(t_{-1}) + k_{12} \cdot C_1(t_{-1}) \cdot \Delta t - k_{21} \cdot C_2(t_{-1}) \cdot \Delta t$$

```
1 # Duration of simulation and delta t
2 days      = 0.01
3 t_end     = days * 86400      # duration of simulation in seconds
4 dt        = 0.1                # time step of iteration
5
6 # rate constants (1/s)
7 k_1_2     = 0.005
8 k_2_1     = 0.002
9
10 # Compartment volumes (cm³)
11 v_1       = 2
12 v_2       = 2
13
14 # Start concentrations (mol/cm³)
15 c_1       = 100
16 c_2       = 0
17 c_ratio   = c_1/c_2
```

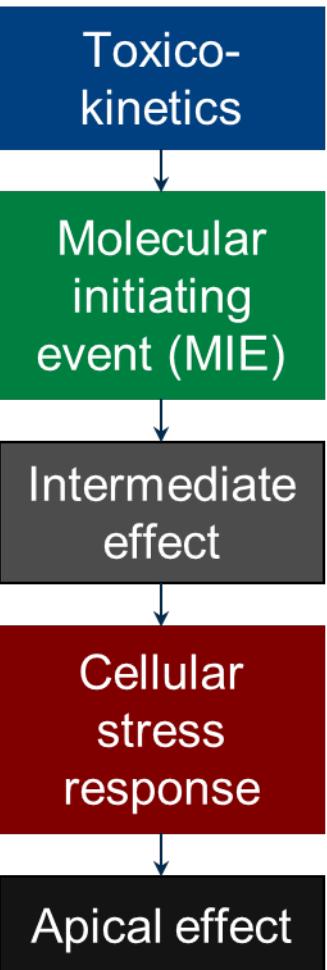
```
29 ##### Loop calculation #####
30 for (t in times) {
31
32   c_1      = c_1
33   c_2      = c_2
34   c_ratio  = c_ratio
35
36   # Change in concentrations per time point (dt)
37   c_1 = c_1 - k_1_2 * c_1 * dt + k_2_1 * c_2 * dt
38   c_2 = c_2 + k_1_2 * c_1 * dt - k_2_1 * c_2 * dt
39
40   # Concentration ratio between compartments
41   c_ratio = c_2/c_1
42
43   if (t %% 60 < dt) {
44     c_1_report[i]      = c_1
45     c_2_report[i]      = c_2
46     c_ratio_report[i] = c_ratio
47     t_report[i]        = t/60
48     i=i+1
49 }
```

# *In vitro* cell assays to measure toxicokinetics and various toxic endpoints – Translation of effect concentrations



adapted from Escher and Leusch (2012)

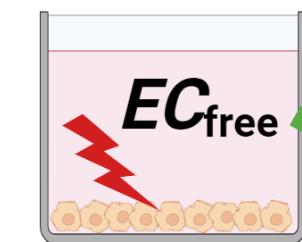
- ! Cost-efficient, reproducible in high-throughput format, classified as non-animal test<sup>1</sup>
- ! Potential to reveal mode of action (MoA)
- ! Large databases already exist, e.g., “Toxicology in the 21st Century” (Tox21) program<sup>2</sup>  
→ Databases neglect chemical bioavailability



! Quantitative predictive power depends on controlling & assessing **effective concentrations!**

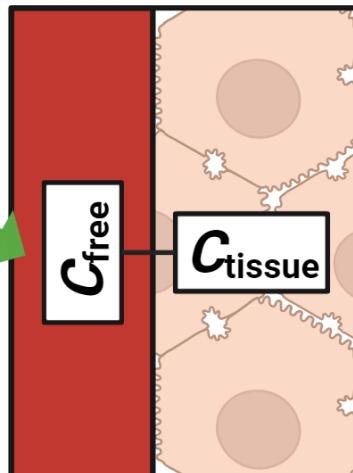
!  $C_{\text{free}}$  and  $C_{\text{cell}}$

*in vitro*



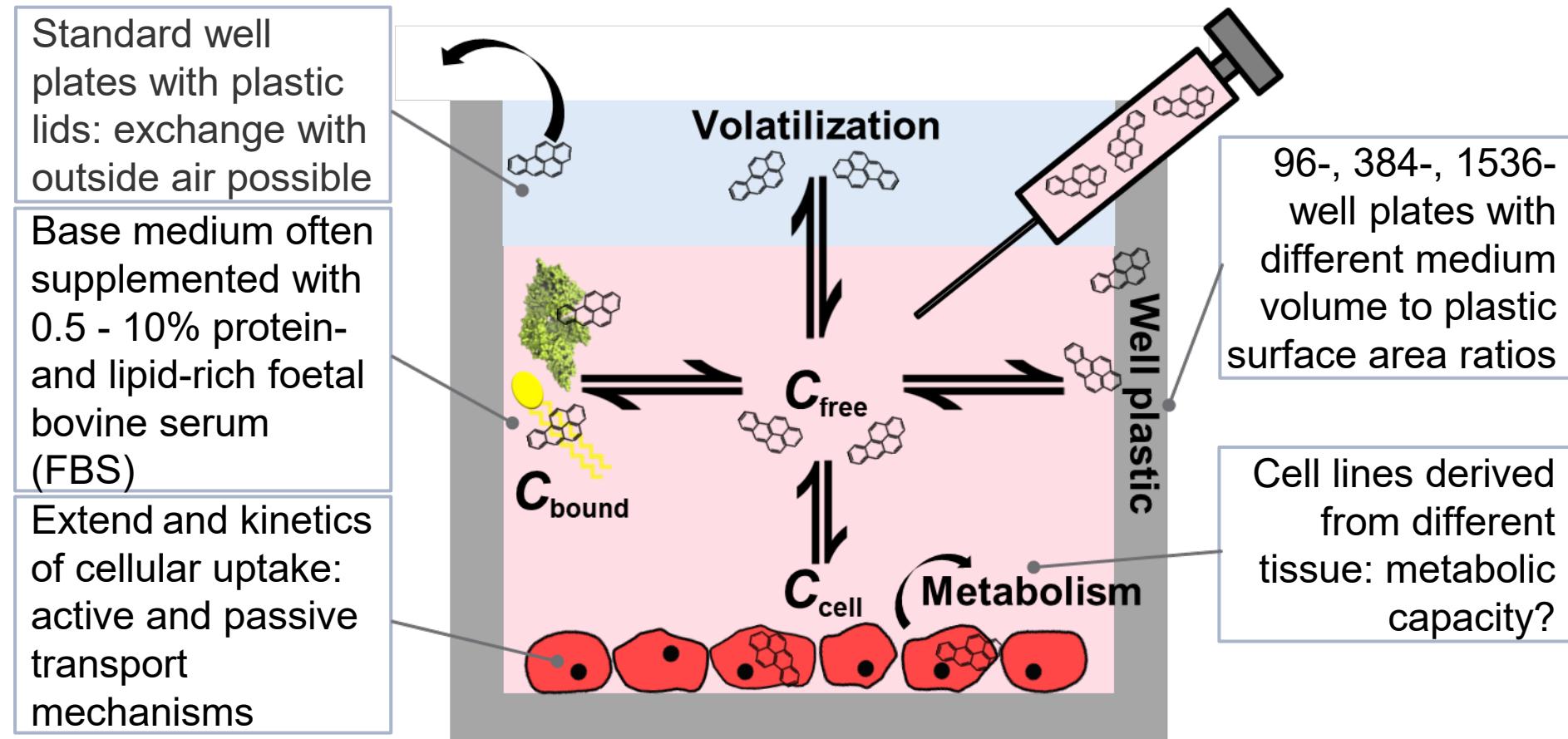
QIVIVE

*in vivo*



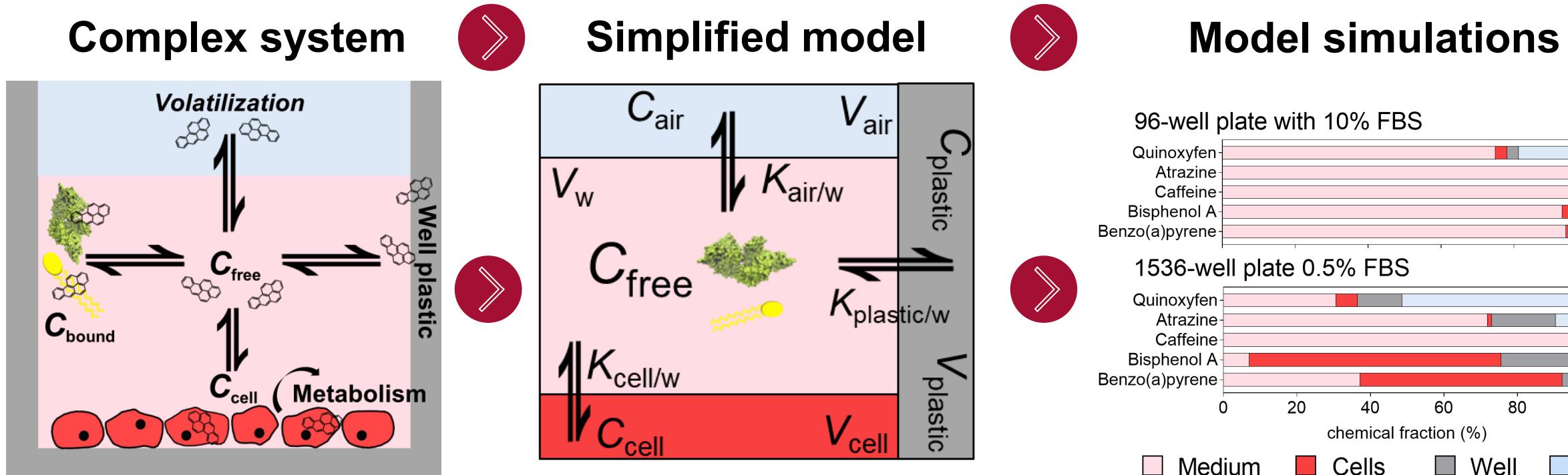
$$\frac{EC_{\text{free}} \text{ (in vitro)}}{C_{\text{free}} \text{ (in vivo)}} = \text{risk}$$

# Chemical fate in *in vitro* cell-based assays results from various processes



What level of model complexity do we need to predict exposure ( $C_{free}$ ,  $C_{cell}$ ) reasonably accurate?

# Equilibrium mass balance model to quantify the effective dose in HTS *in vitro* bioassays



$$f_{free} = \left( 1 + K_{cell/w} \cdot \frac{V_{cell}}{V_w} + K_{air/w} \cdot \frac{V_{air}}{V_w} + K_{plastic/w} \cdot \frac{V_{plastic}}{V_w} \right)^{-1}$$

$$C_{free} = C_{nom} \cdot f_{free} \cdot \frac{V_{total}}{V_w}$$

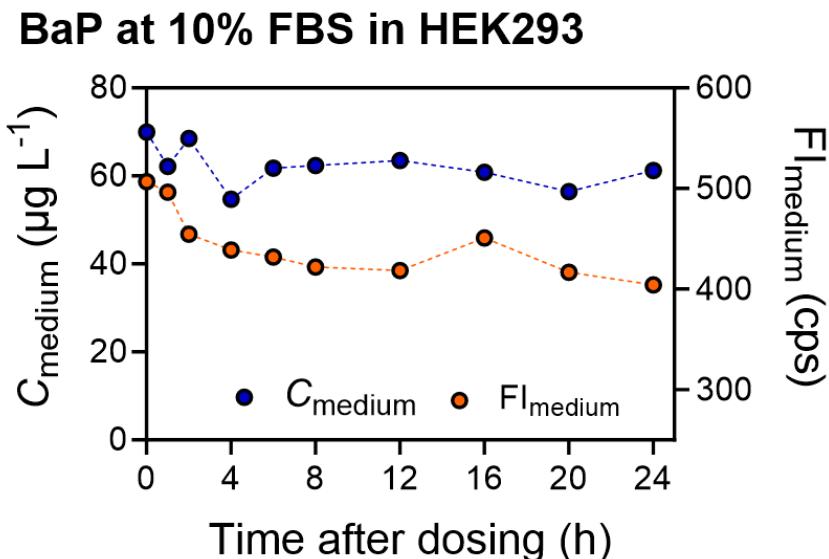
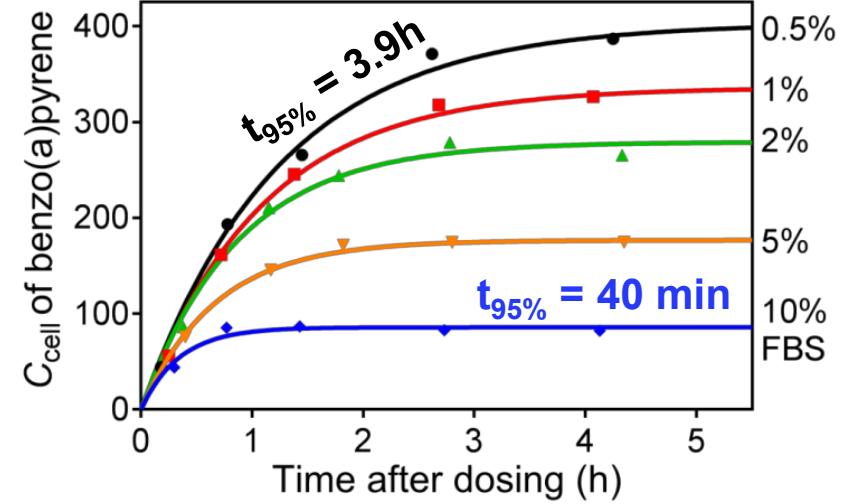
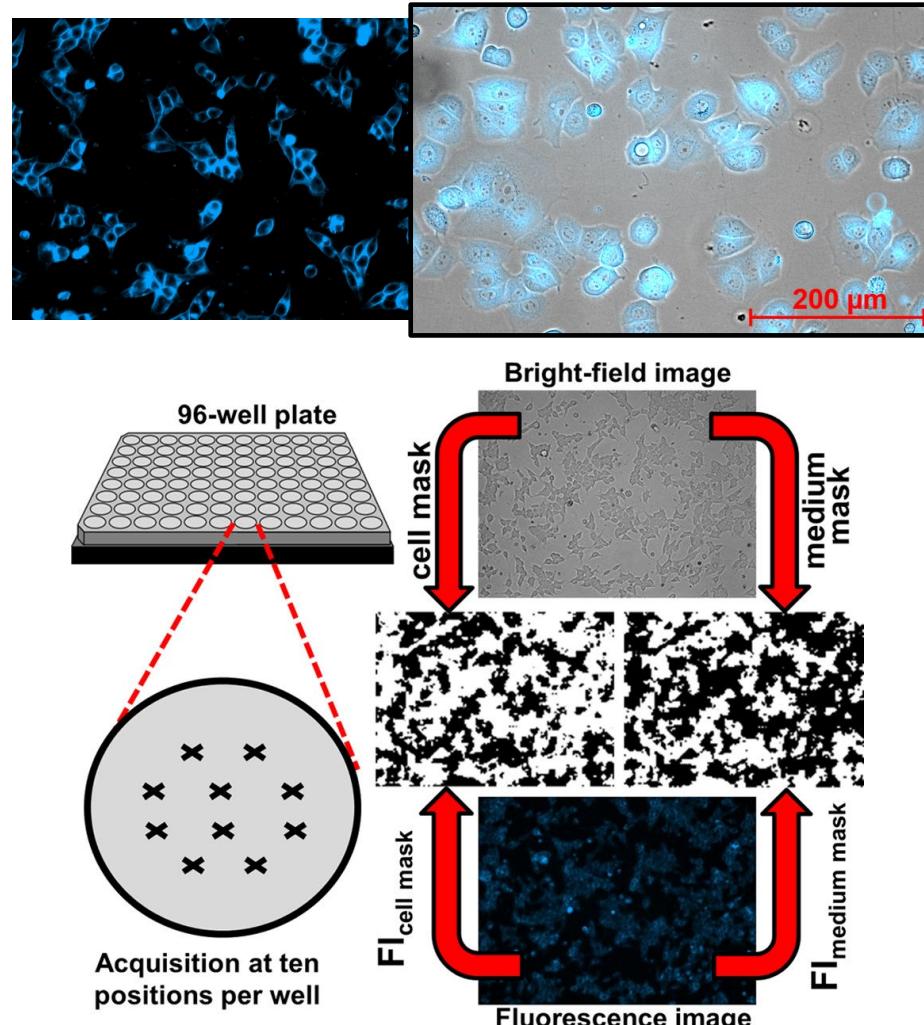
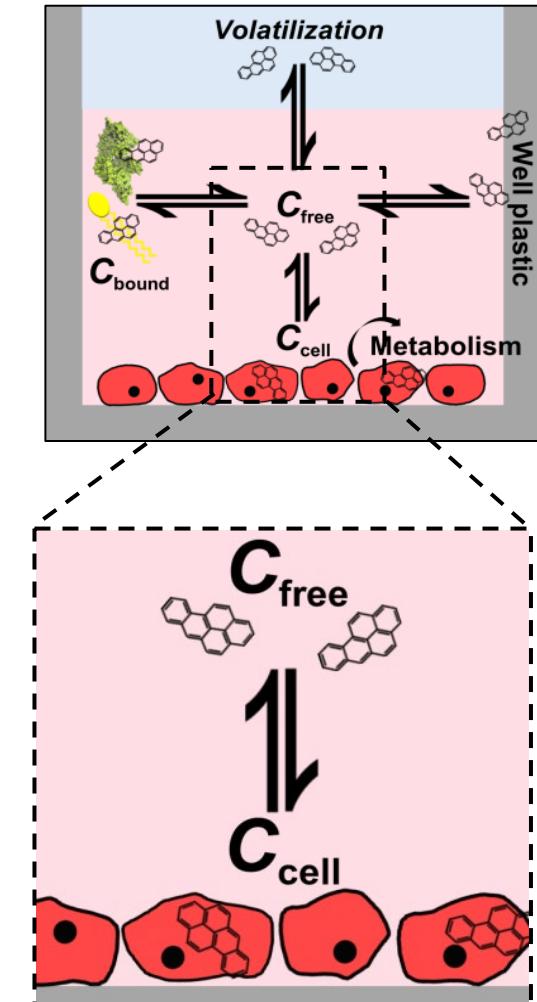
## Modeling Exposure in the Tox21 *in Vitro* Bioassays

Fabian C. Fischer,<sup>\*†</sup> Luise Henneberger,<sup>†</sup> Maria König,<sup>†</sup> Kai Bittermann,<sup>‡</sup> Lukas Linden,<sup>‡</sup> Kai-Uwe Goss,<sup>‡</sup> and Beate I. Escher<sup>†</sup>

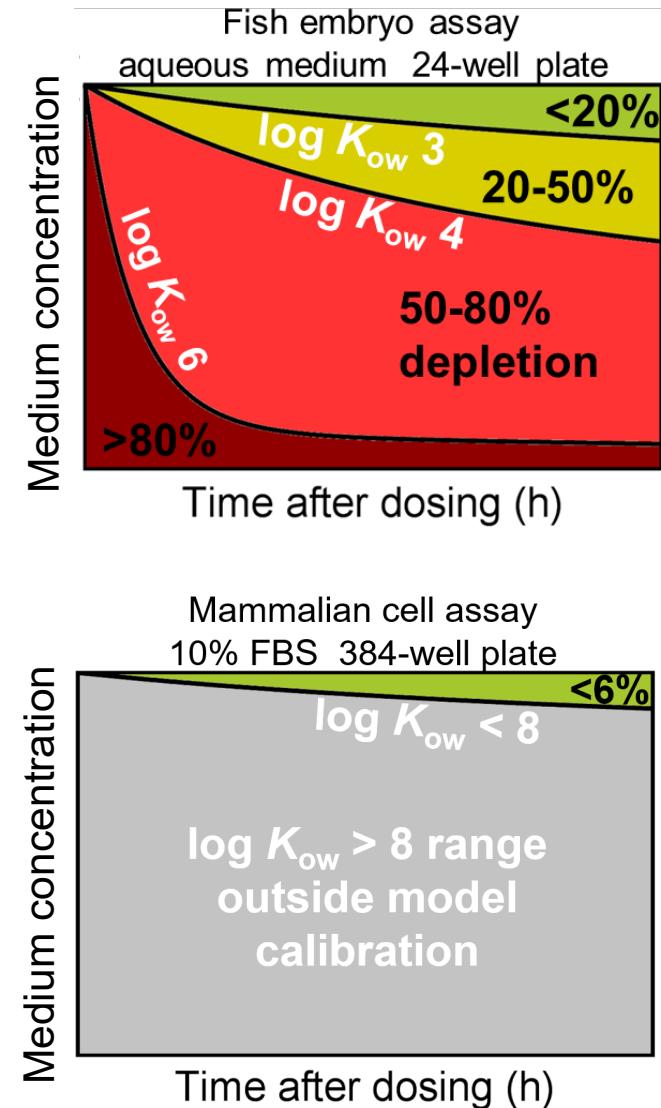
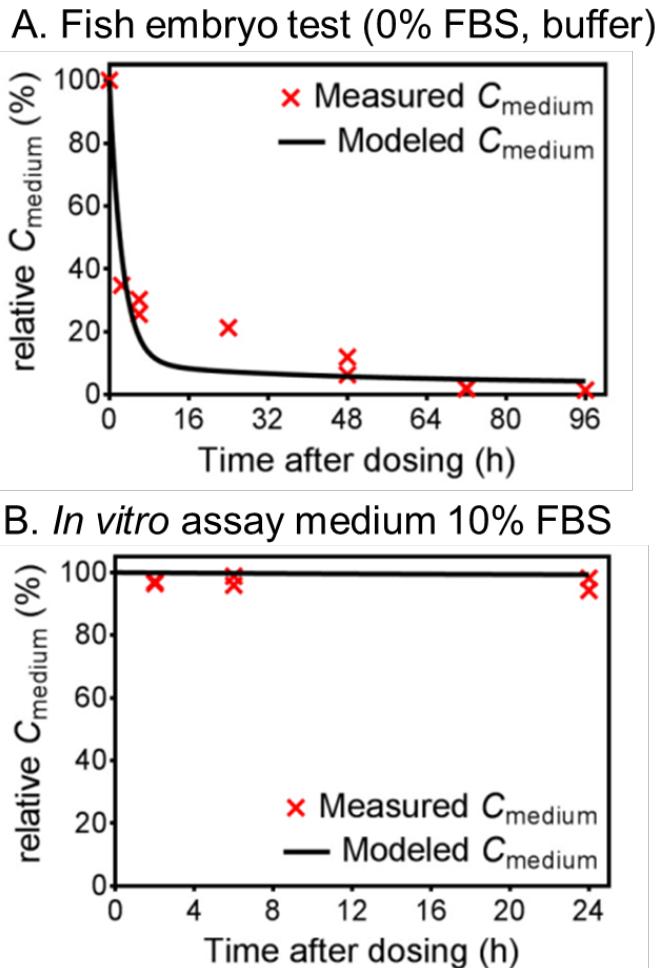
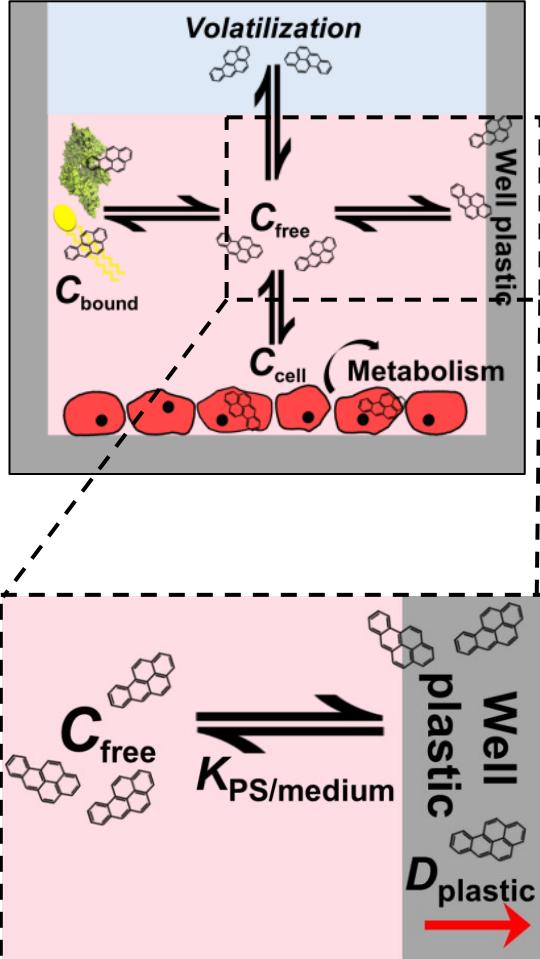
**FBS proteins**  
and **lipids**



# Measuring chemical uptake kinetics with fluorescence microscopy

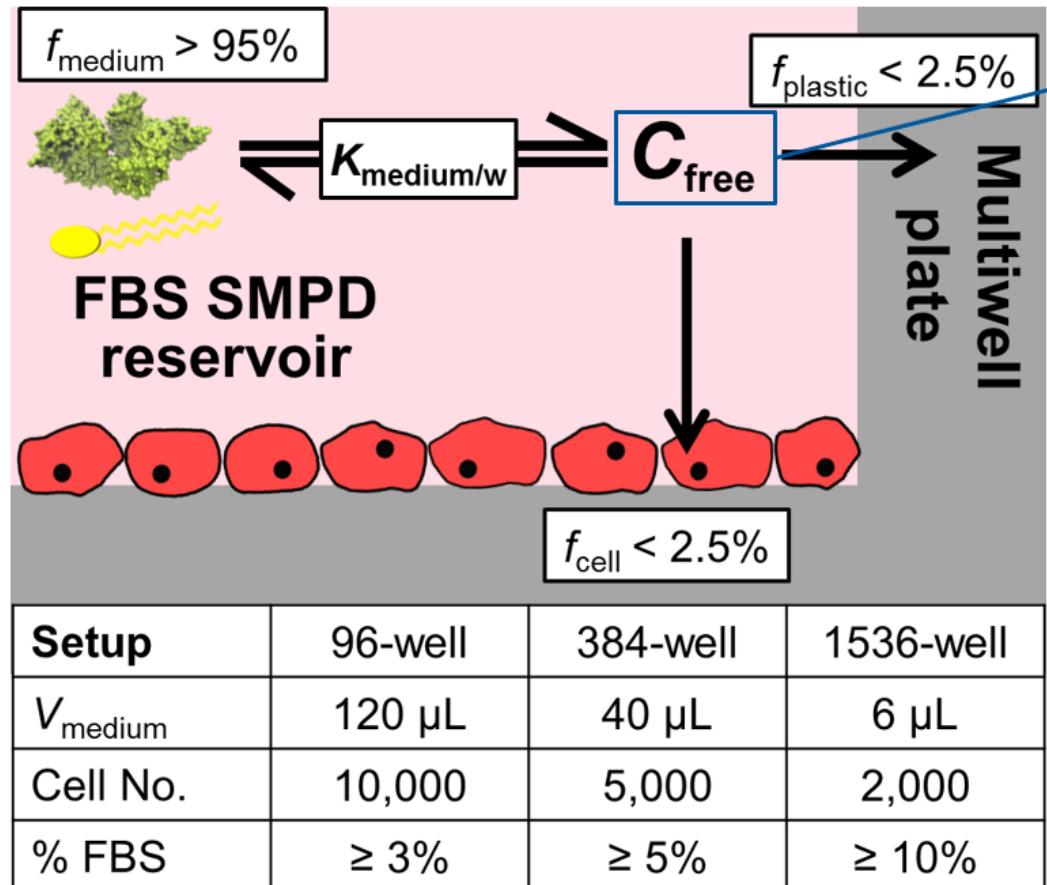


# Transport of chemicals into the plastic material of multi-well plates used for bioassays

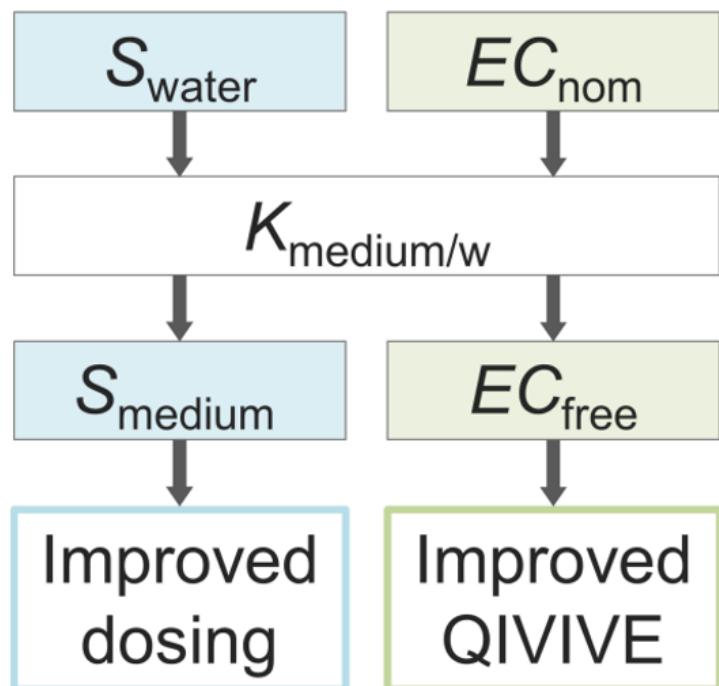


# Adjusting the FBS content in the medium to achieve controlled exposure conditions and easier modeling

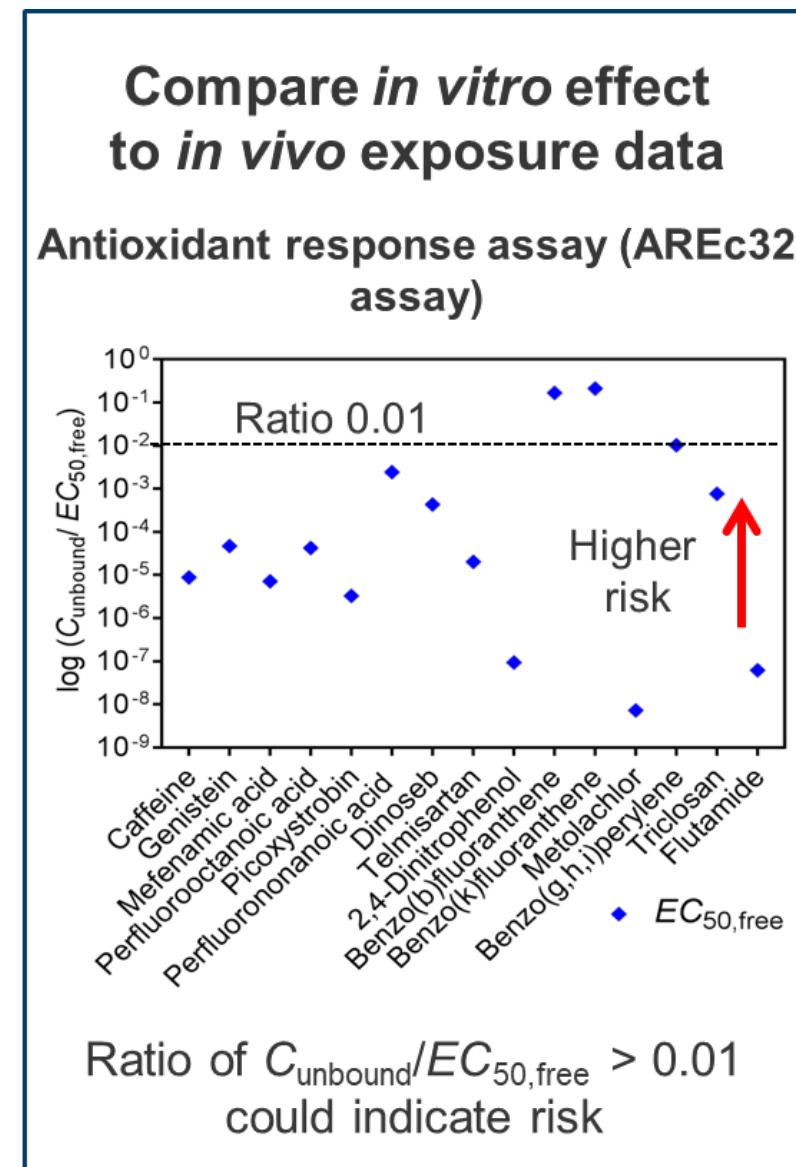
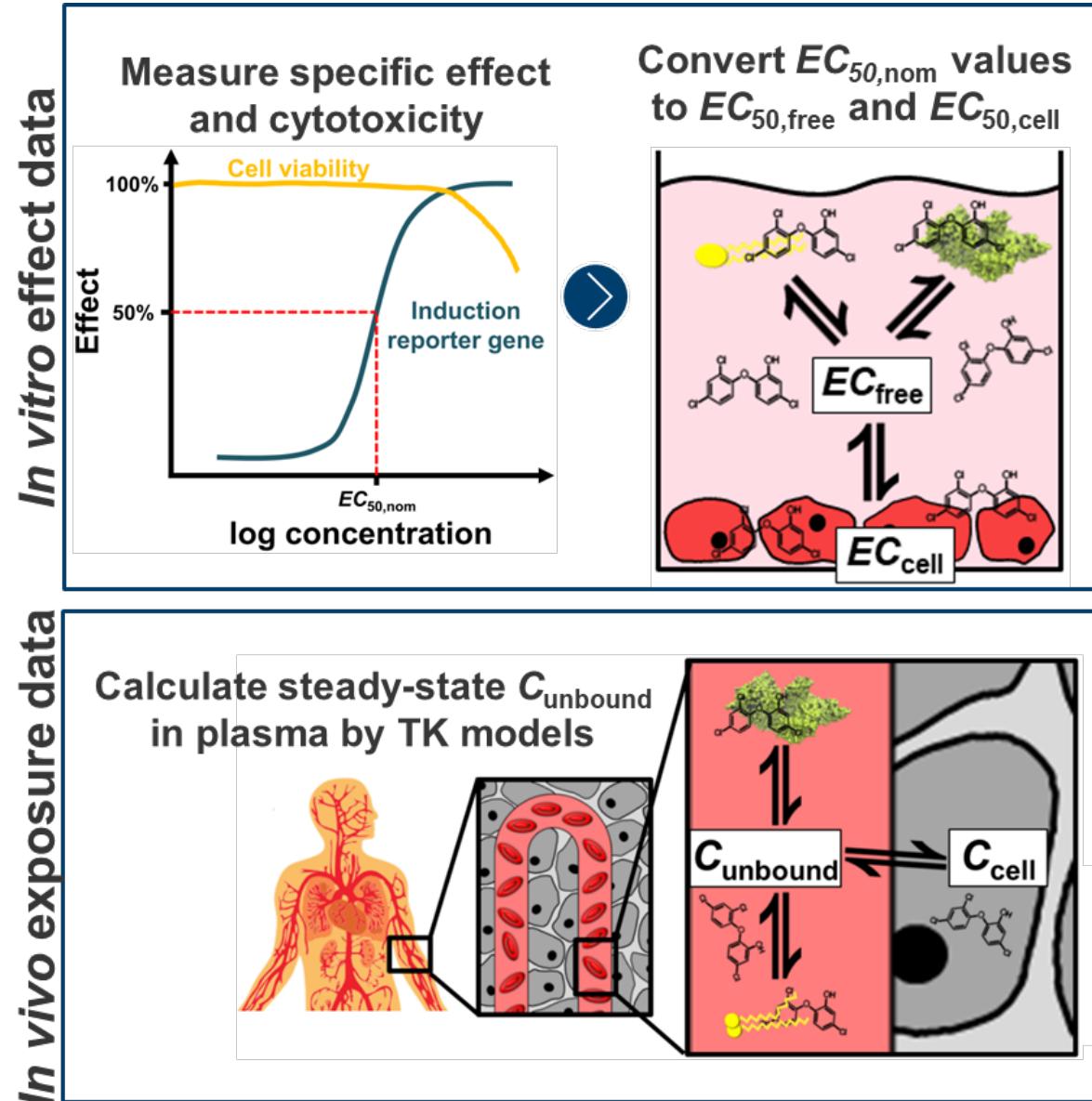
## Equilibrium model sufficiently accurate?



if  $C_{\text{free}}$  remains stable over time by SMPD

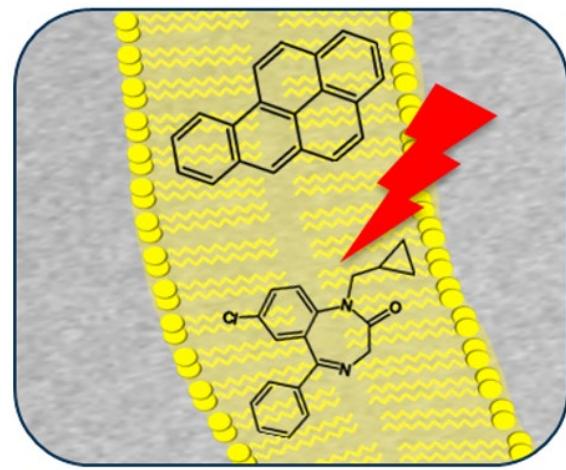
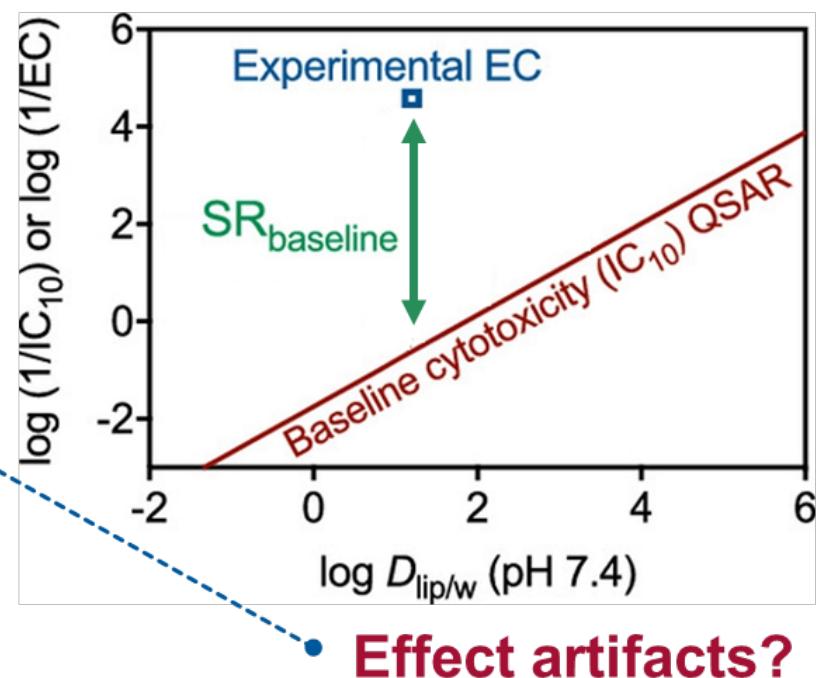
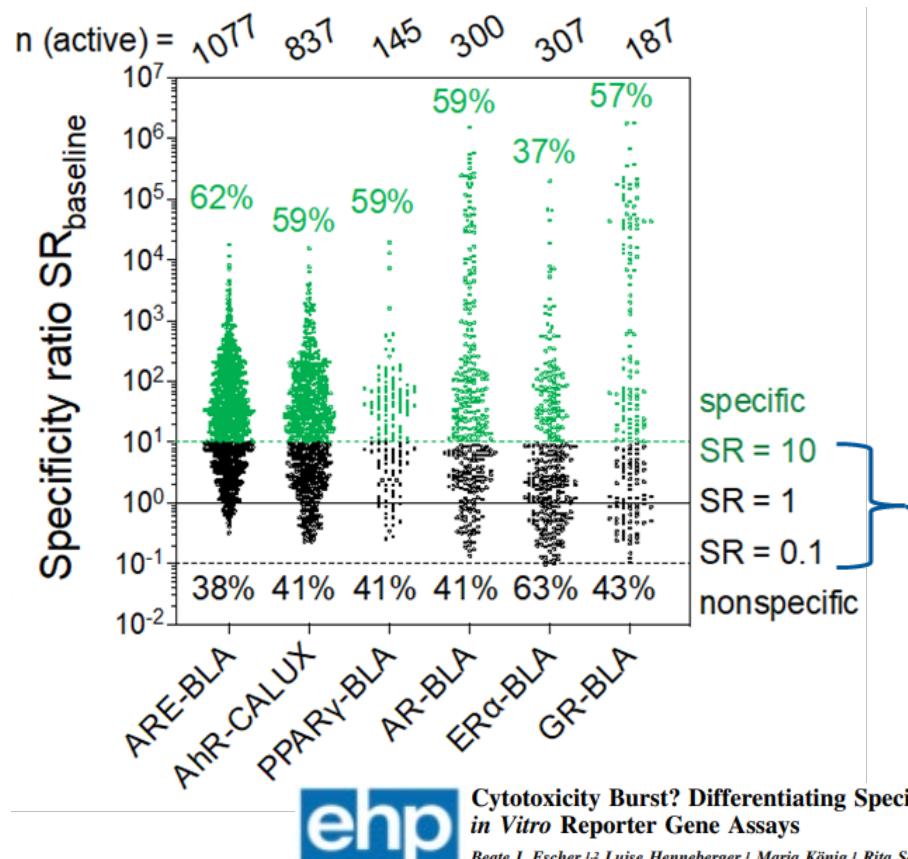


# Establishment of a quantitative extrapolation framework for risk assessment to protect human health



# Cytotoxicity burst – Specificity analysis of *in vitro* HTS effect data using mass balance modeling

- Automated processing of ~8,000 chemicals with ~380,000 data points in MATLAB
- Specificity analysis of ToxCast/Tox21 effect data to identify highly potent chemicals and *potentially* false-positive effects

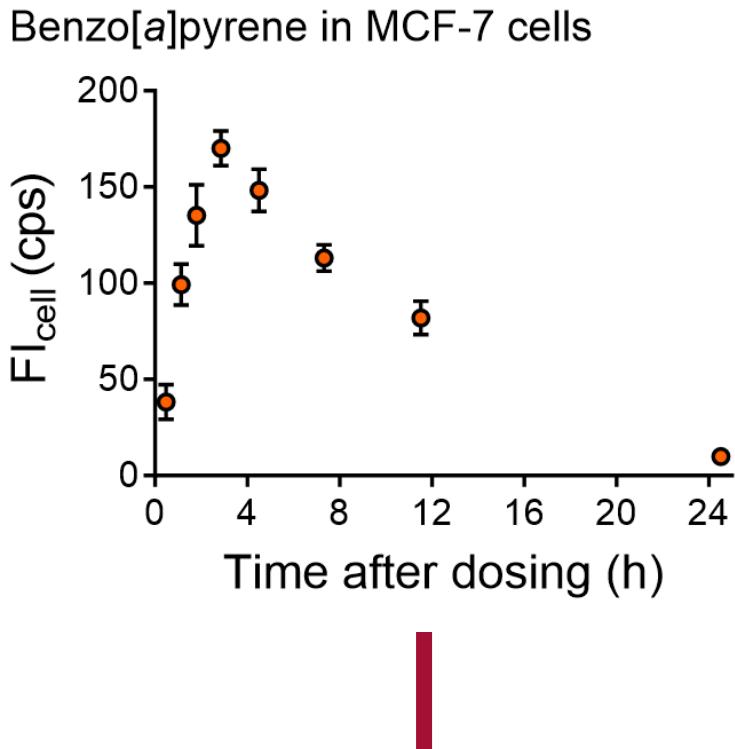
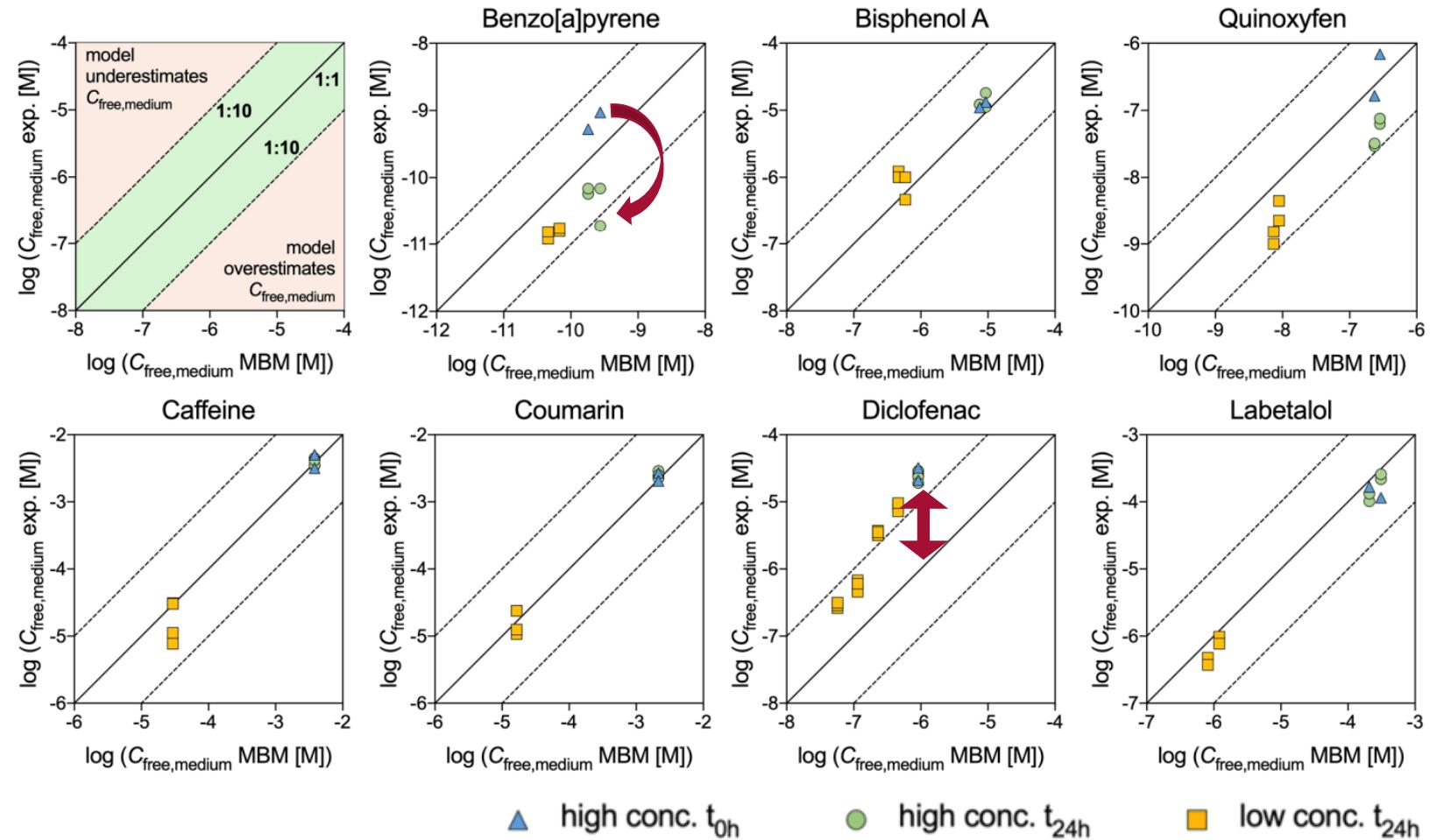


Concept applicable to other modes of action?



Cytotoxicity Burst? Differentiating Specific from Nonspecific Effects in Tox21  
*in Vitro* Reporter Gene Assays  
Beate I. Escher,<sup>1,2</sup> Luise Henneberger,<sup>1</sup> Maria König,<sup>1</sup> Rita Schlichting,<sup>1</sup> and Fabian C. Fischer<sup>1†</sup>

# Experimental evaluation of model performance: Measuring $C_{\text{free}}$ in *in vitro* cell assays

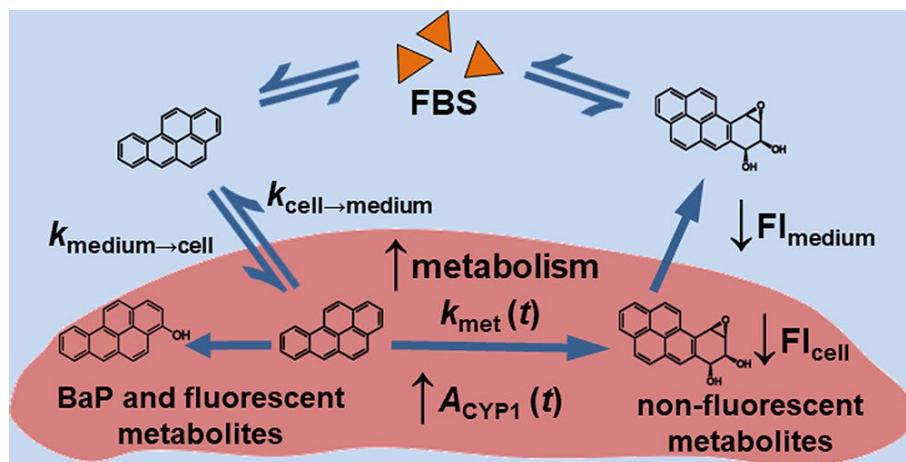
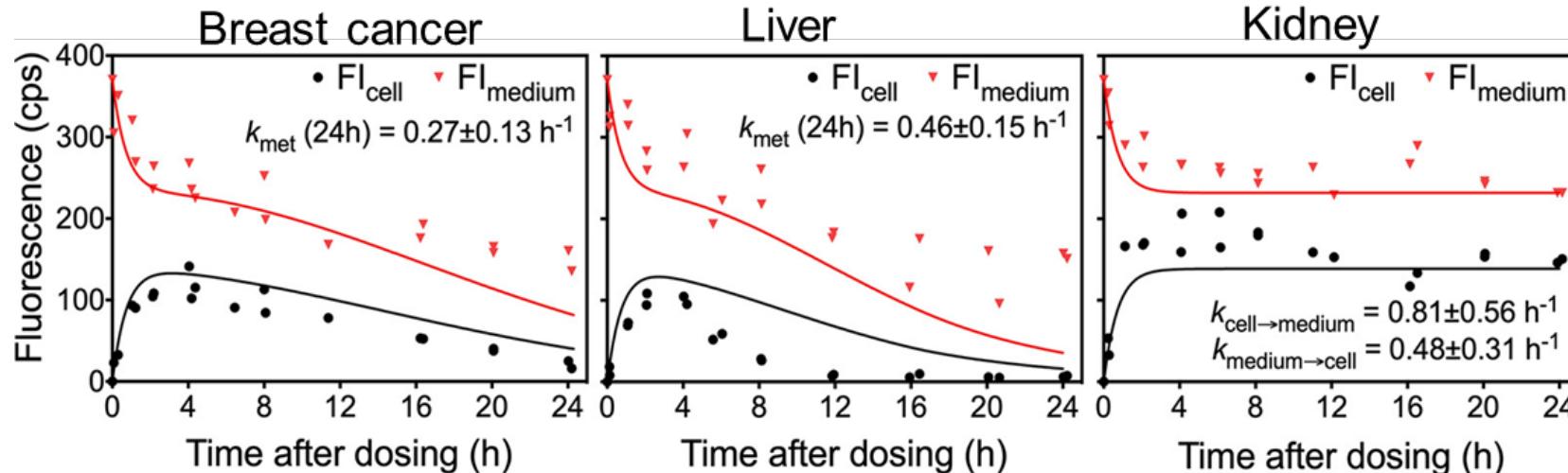


Metabolism?

Quantification of freely dissolved effect concentrations in *in vitro* cell-based bioassays

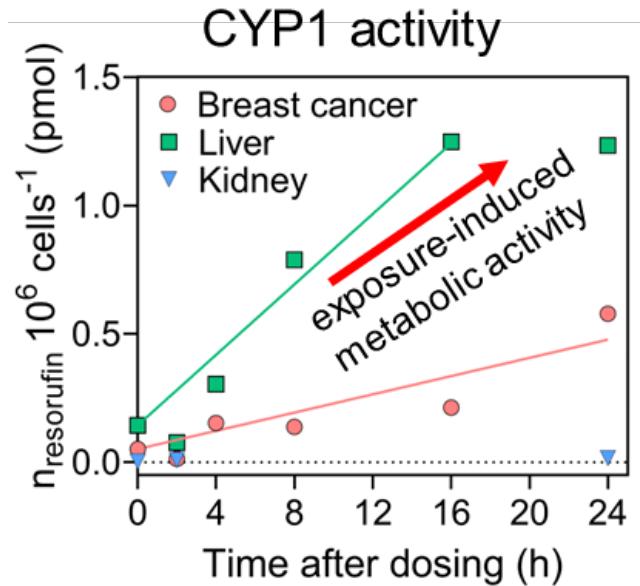
Luise Henneberger<sup>1</sup> · Marie Mühlenbrink<sup>1</sup> · Maria König<sup>1</sup> · Rita Schlichting<sup>1</sup> · Fabian C. Fischer<sup>1</sup> ·  
Beate I. Escher<sup>1,2</sup>

# Characterizing metabolism of *in vitro* cells: Fit experimental data to 2-comp kinetic transport model



$$\frac{d\text{FI}_{\text{medium}}}{dt} = k_{\text{cell} \rightarrow \text{medium}} \cdot \text{FI}_{\text{cell}}(t) - k_{\text{medium} \rightarrow \text{cell}} \cdot \text{FI}_{\text{medium}}(t)$$

$$\frac{d\text{FI}_{\text{cell}}}{dt} = k_{\text{medium} \rightarrow \text{cell}} \cdot \text{FI}_{\text{medium}}(t) - (k_{\text{cell} \rightarrow \text{medium}} + k_{\text{met}}(t)) \cdot \text{FI}_{\text{cell}}(t)$$

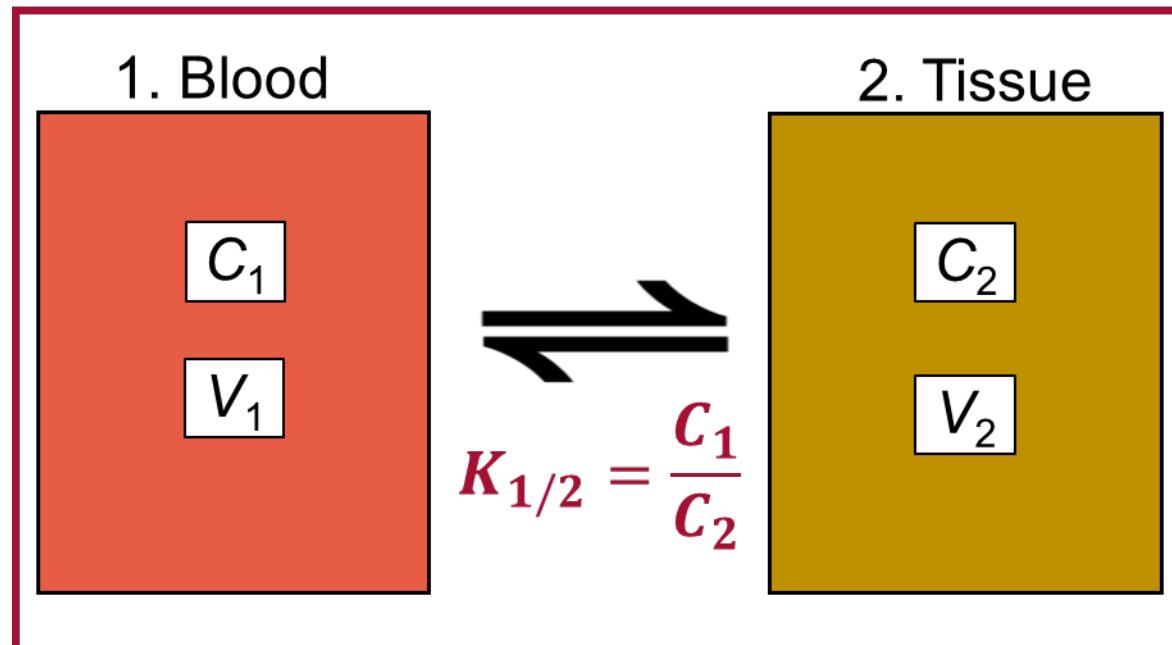


$$k_{\text{met}}(t) = k_{\text{met}} t \frac{m_{\text{CYP}}}{A_{\text{control}}}$$

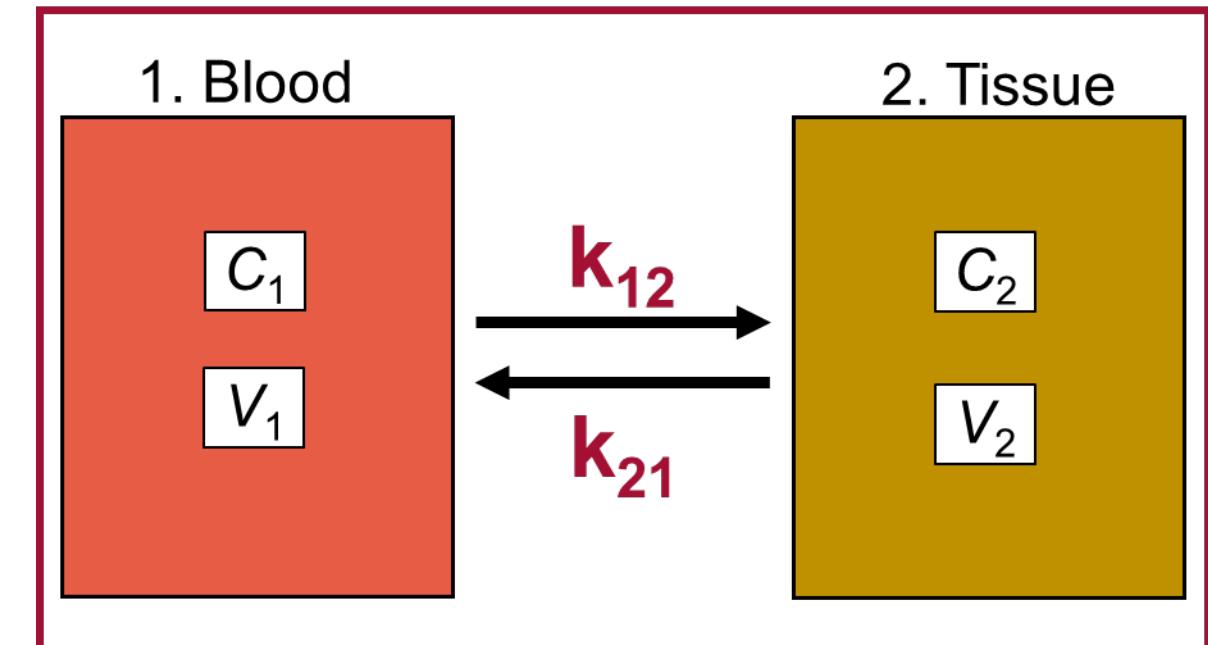
# Chemical transport models to investigate the distribution of chemicals in *in vitro* cell assays

- ? Can the process be approximated with an equilibrium or kinetic model?
- ? How much complexity is necessary to predict/explain exposure?

## Equilibrium mass balance model



## Kinetic model (rate-limited)

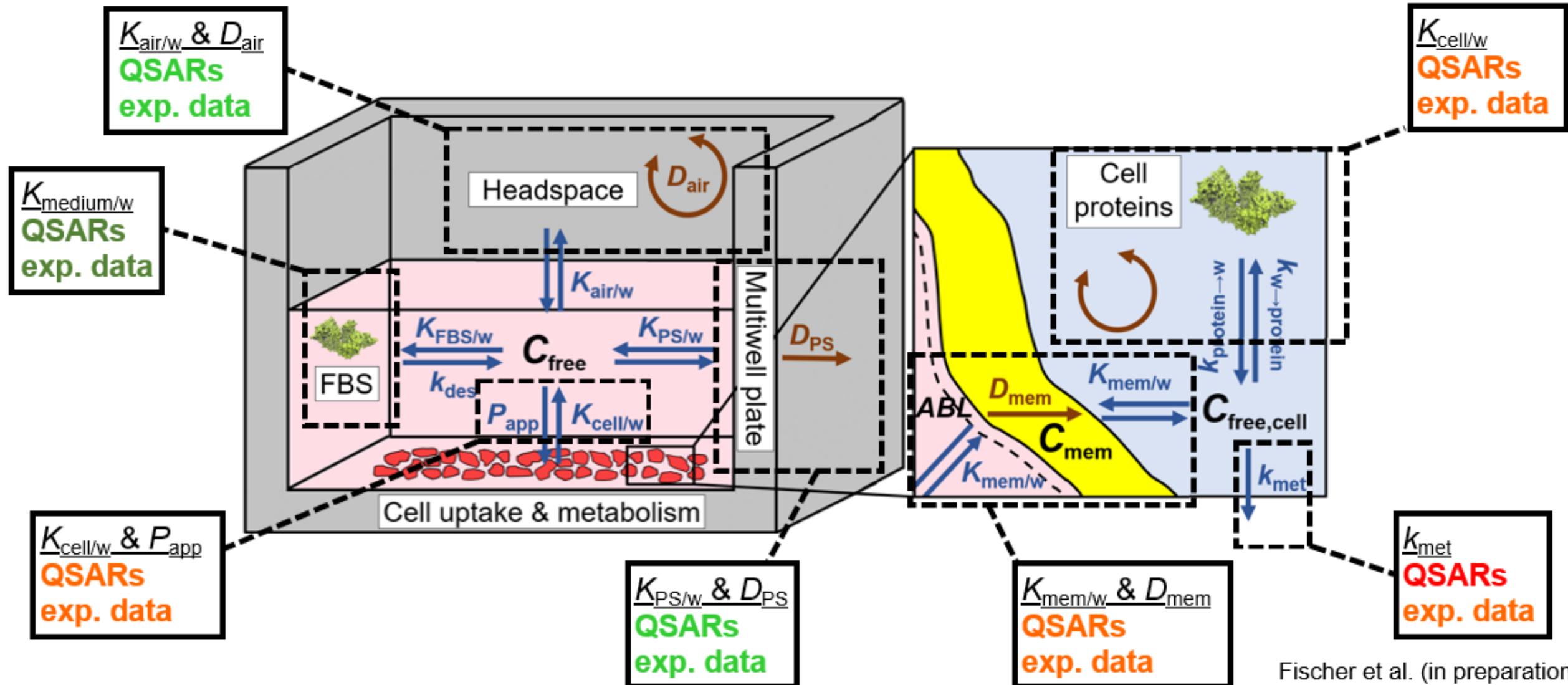


Can we simplify?  
For *in vitro* exposure, YES\*!

\*but there are exceptions

# Work in progress: Implementation of a kinetic transport model for *in vitro* cell-based bioassays

- Limited parameters available to parameterize a comprehensive *in vitro* kinetic model



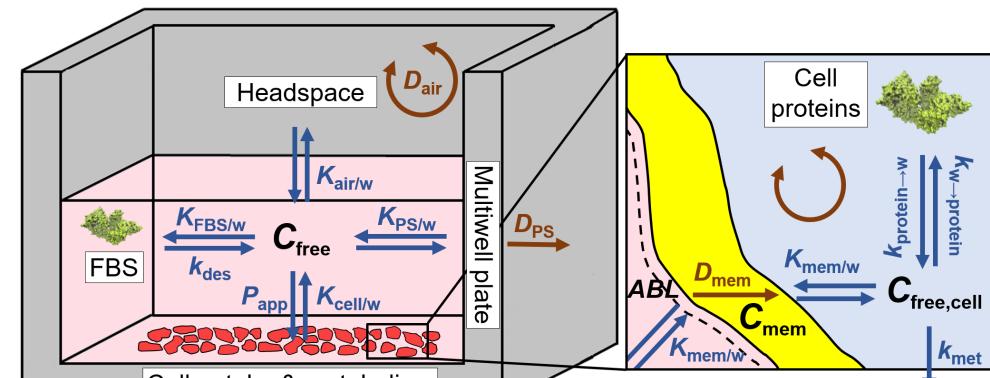
# Work in progress: Implementation of a kinetic transport model for *in vitro* cell-based bioassays

```
#### SECTION 2: DEFINITION OF CHEMICAL PARAMETERS ####
Mw      = 250                                # Molecular weight of chemical
Mw_BSA  = 66500                             # Molecular weight of BSA
D_BSA   = 7.0 * 10^-5 * Mw_BSA^(-0.45)

# Literature partition coefficients
K_alb_w = 1 * 10^4                            # albumin-water partition coefficient (L_w/L_alb)
K_lip_w = 1 * 10^4                            # phospholipid-water partition coefficient (L_w/L_lip)
K_well_w = 1 * 10^4                            # well plastic-water partition coefficient (L_w/L_well)
K_aw    = 5 * 10^-7                            # air-water partition coefficient (L_w/L_air)

# Kinetic parameters
D_water  = 9.9 * 10^-5 * Mw^(-0.45)
D_ABL    = D_water + K_alb_w * V_prot_medium * D_BSA
D_mem    = 10^(-5.13-0.453*log10(Mw))
P_app    = 1 / ((x_mem/(D_mem * K_lip_w))+(x_w/D_ABL))
D_well   = 1 * 10^-12
k_met    = 0.01
k_abiot  = 1 * 10^-5

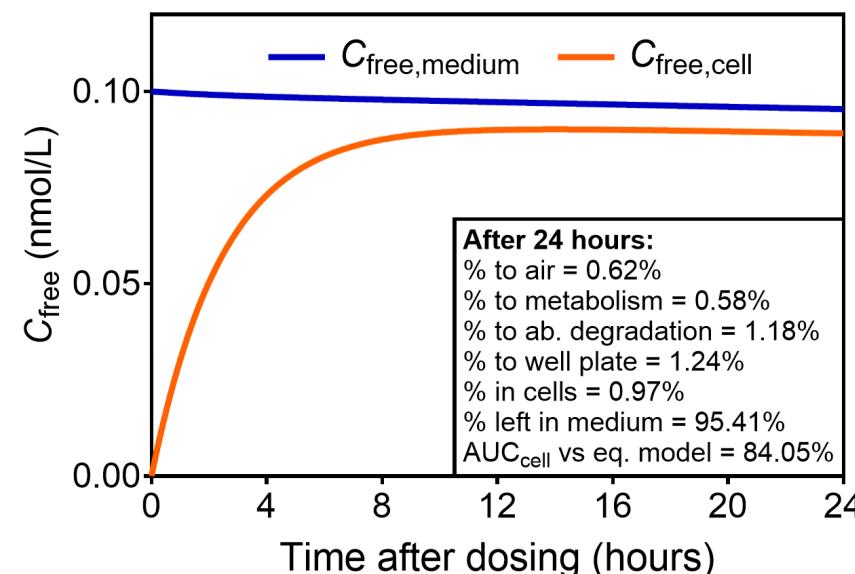
# Diffusion coefficient of chemical in water (cm^2/s)
# Diffusion coefficient of chemical in ABL (cm^2/s)
# Diffusion coefficient of chemical in membrane (cm^2/s)
# Apparent permeability of chemical (cell uptake) (cm/s)
# Diffusion coefficient of chemical in well plastic (cm^2/s)
# Elimination rate constant (cell metabolism) (1/s)
# Abiotic degradation rate constant in medium (1/s)
```



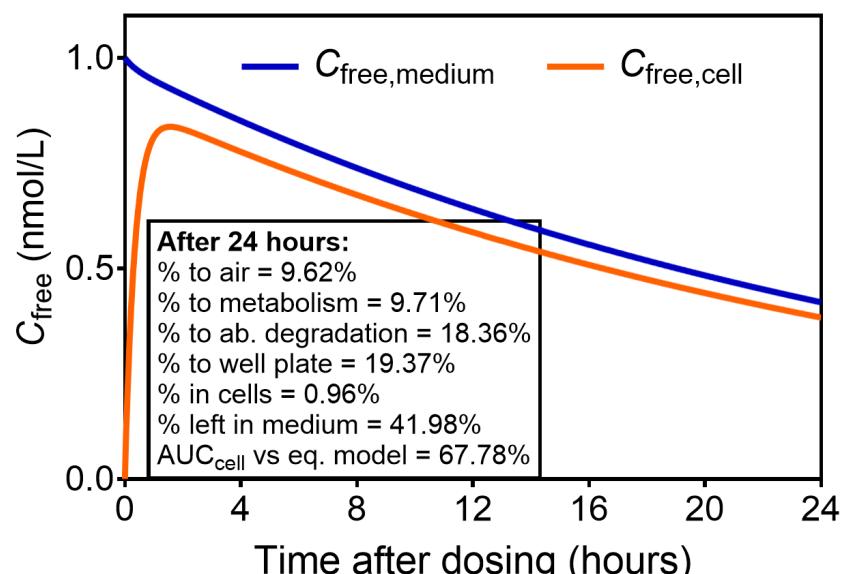
Fischer et al. (in preparation)

➤ 96-well plate, 120 µL OptiMEM + 10% FBS

A.  $K_{\text{medium/w}} = 10,000$

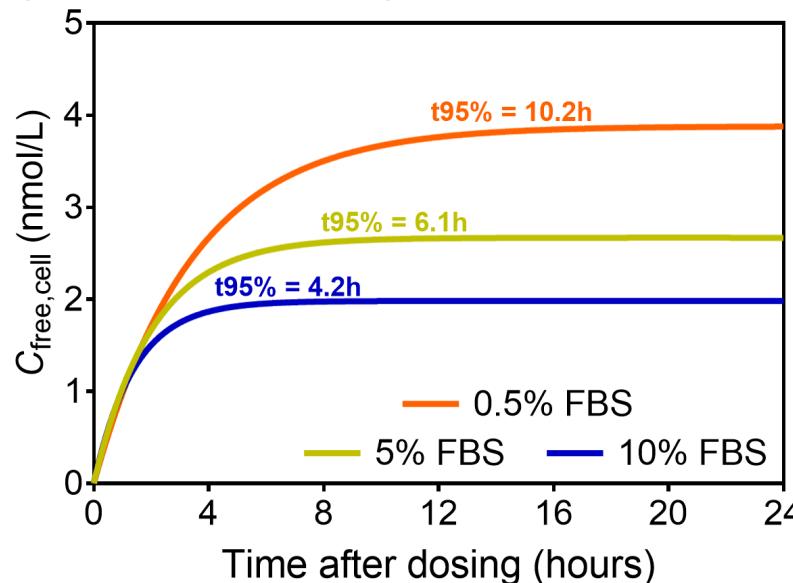


B.  $K_{\text{medium/w}} = 1,000$



➤ 0.5% - 10% FBS

C. BaP at different FBS





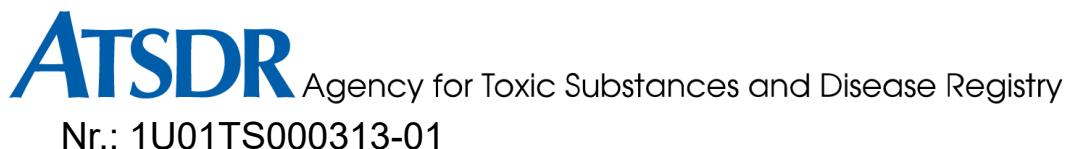
Harvard John A. Paulson  
School of Engineering  
and Applied Sciences



## Collaborators and Colleagues



## Funding



# Acknowledgments



BROWN



Thank you for your attention! Questions?



Biogeochemistry of  
Global Contaminants  
HARVARD

# Impact of Chemical-PDMS Interactions on IVIVE from Microphysiological Systems

PI: M. Shane Hutson

Co-Investigators:

Lisa J. McCawley

Dmitry Markov

QA Manager

Phillip Fryman

Students

Nate Hermann

Richard Ficek

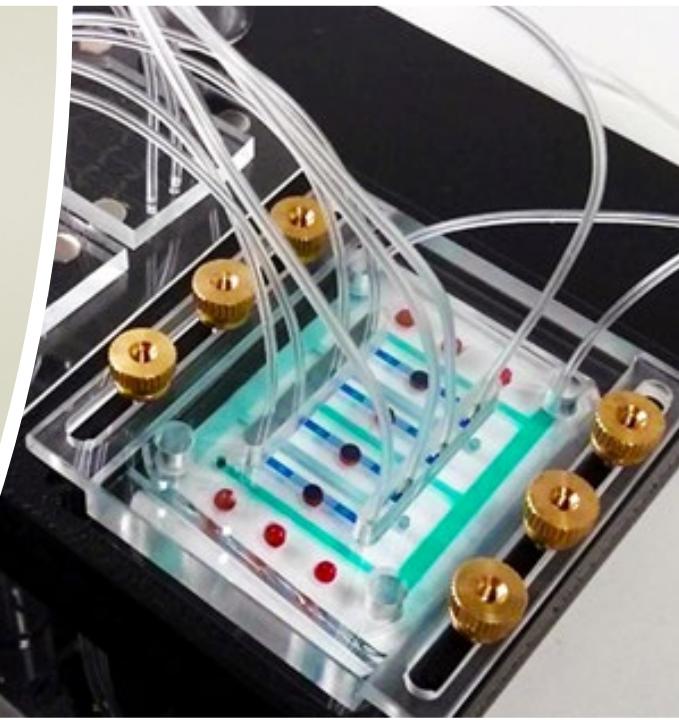
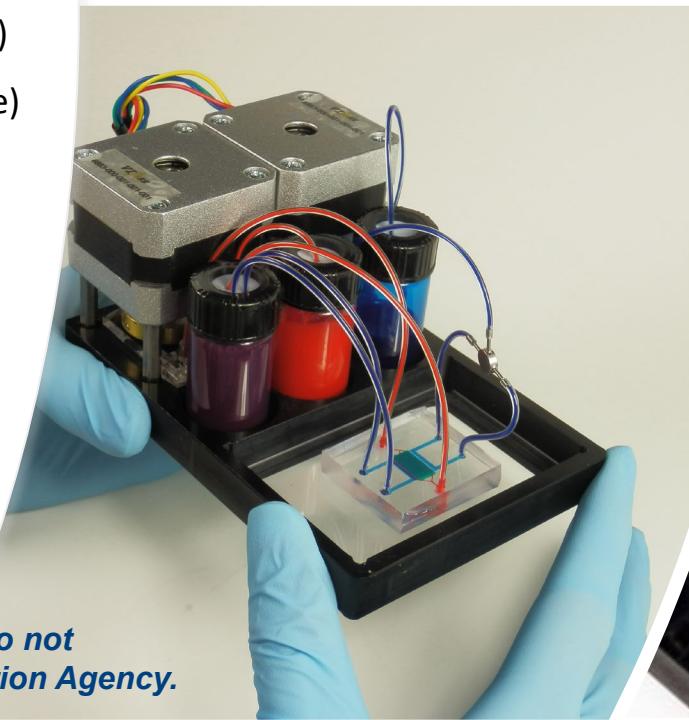
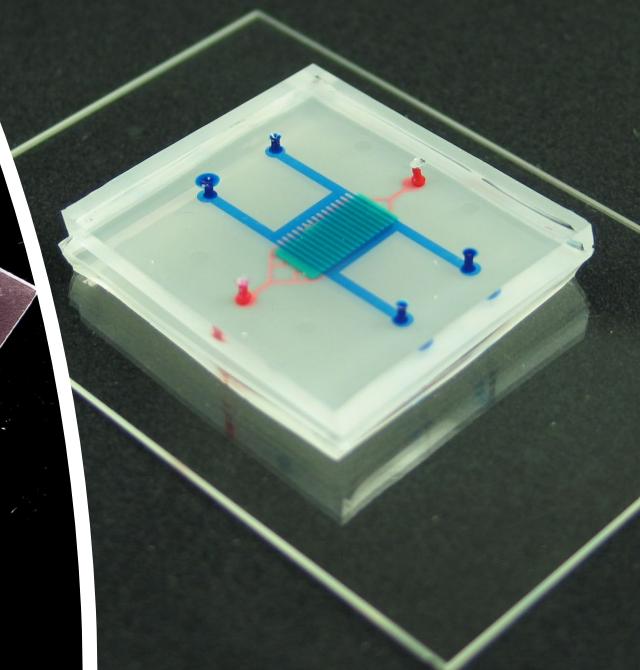
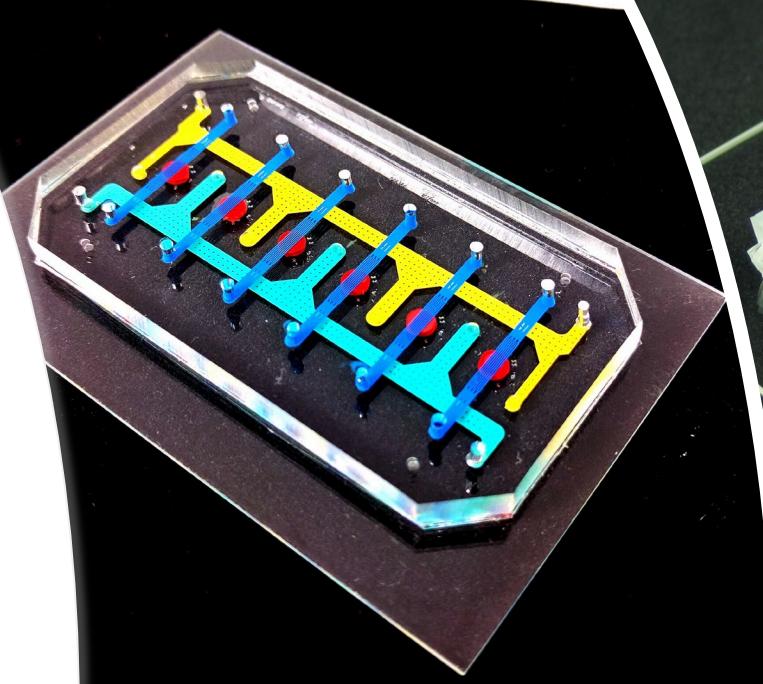
Kazi Tasneem (Genentech)

Alex Auner (Boston College)



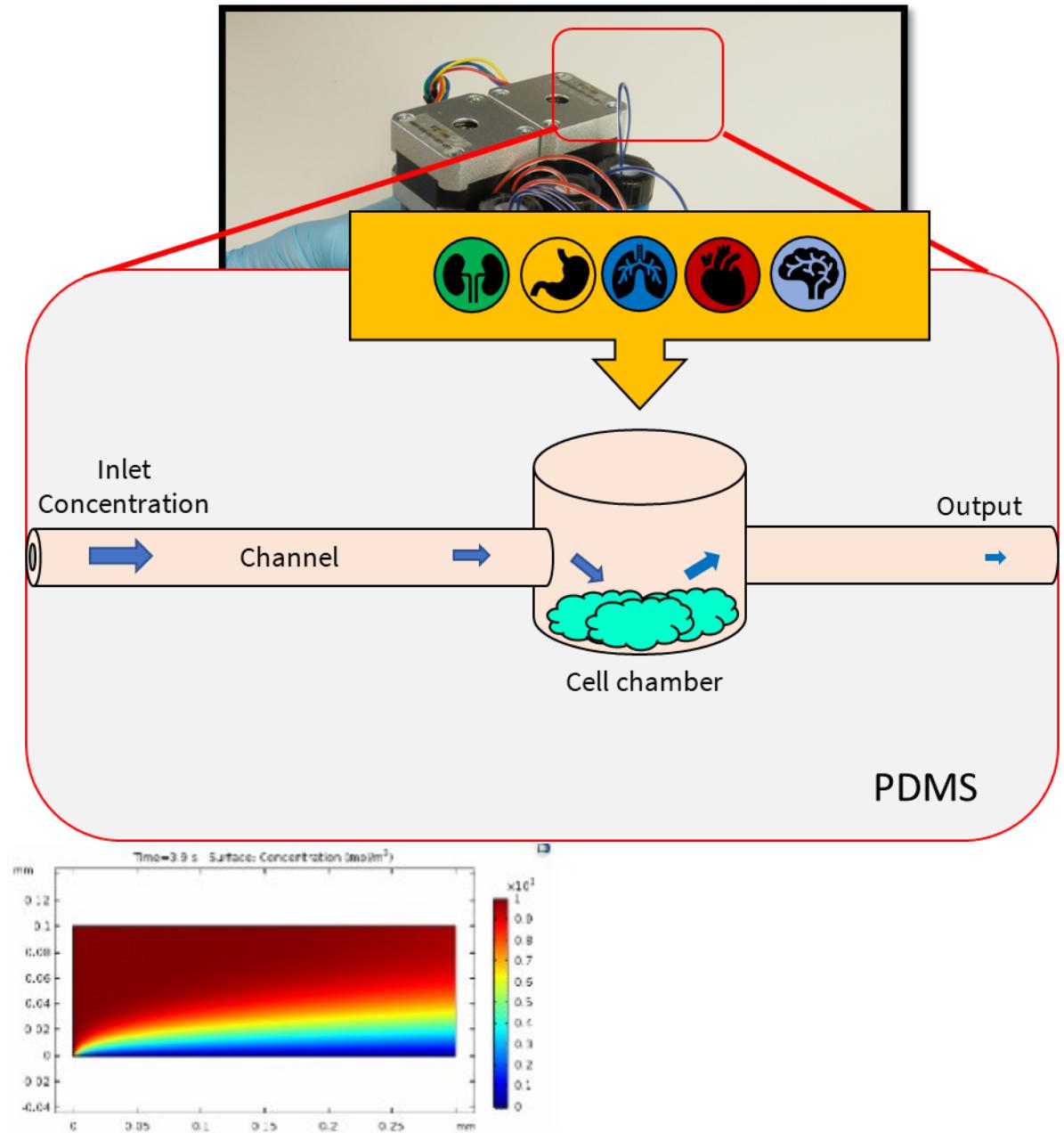
Thanks to U.S. EPA for funding support!

*The views expressed in this presentation are those of the author[s] and do not necessarily reflect the views or policies of the U.S. Environmental Protection Agency.*



# Microphysiological Systems (Organ-on-Chip Devices)

- A New Approach Method (NAM) to fill the bridge between cell-based assays and animal trials.
  - Better representation of human response
  - Useful for high throughput testing
  - Addresses ethical concerns
- Typically made of polydimethylsiloxane (PDMS):
  - Inexpensive
  - Flexible
  - Transparent
  - Gas permeable
- **But PDMS tends to sequester hydrophobic compounds.**
- Need to avoid, mitigate, or measure and model.



# Talk Outline

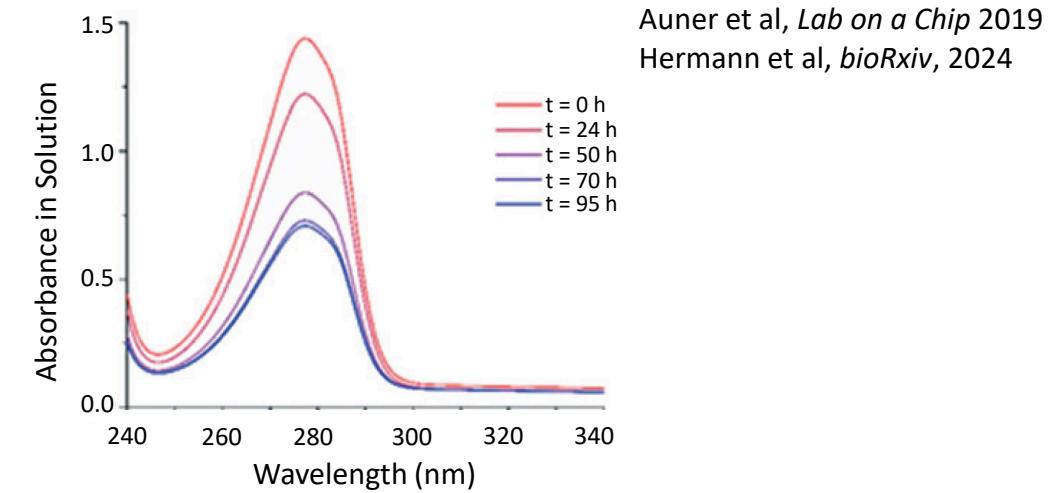
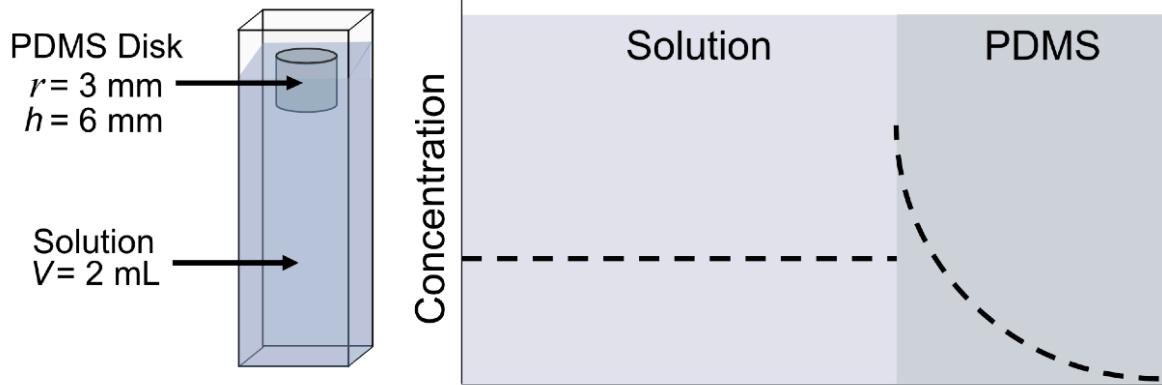
1. Measure chemical-PDMS interaction and transport parameters.
  - a. Partition coefficients:  $k = C_{PDMS}/C_{sol}$
  - b. Diffusion coefficients in PDMS:  $D_p$
  - c. Evaluate correlations with chemical properties
2. Be wary of read-across methods.
3. Validate multiphysics models for in-device toxicokinetics.
4. Investigate the effectiveness of mitigation strategies for reducing chemical sequestration in microfluidic devices:
  - a. SEBS co-polymers as PDMS alternatives; and
  - b. media with carrier proteins.

# Talk Outline

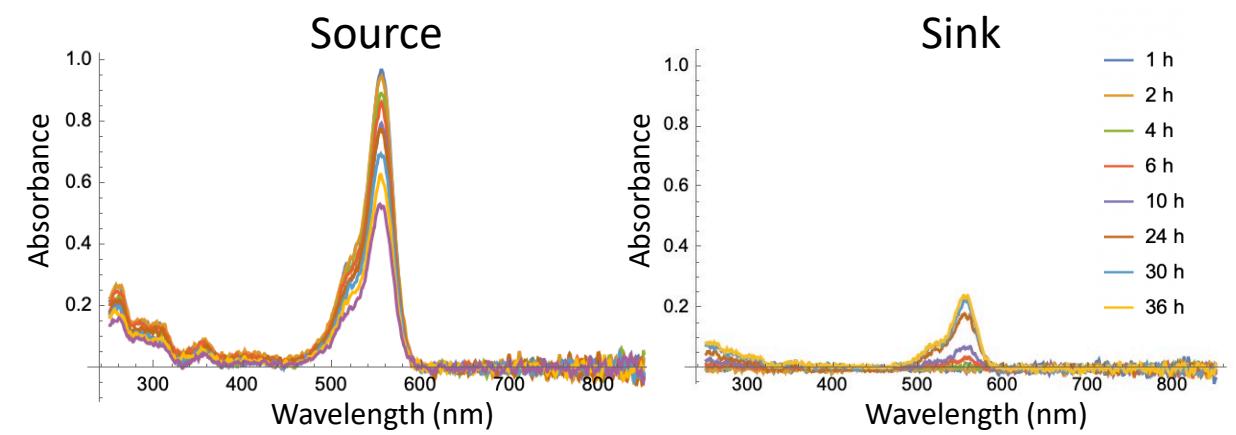
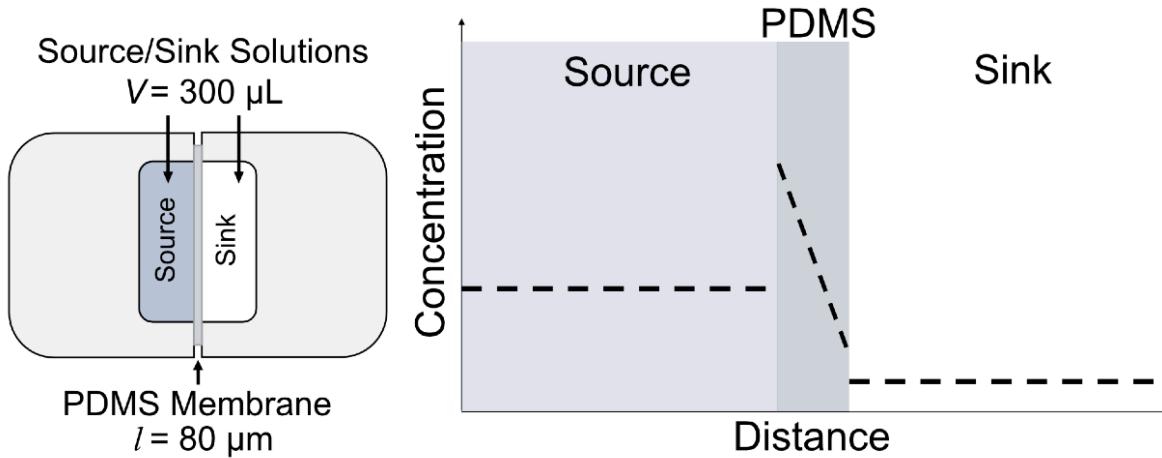
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# Measuring Chemical-PDMS Interaction and Transport Parameters

## Disk Soak Experiments

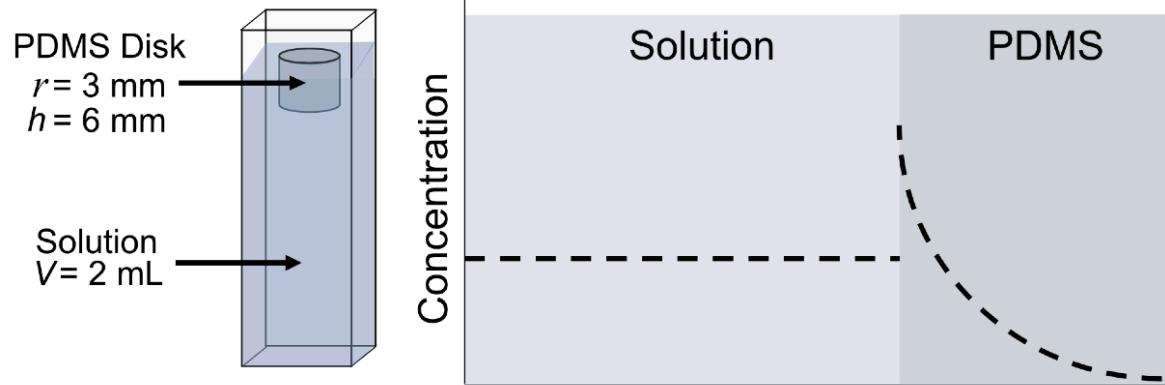


## Membrane Experiments



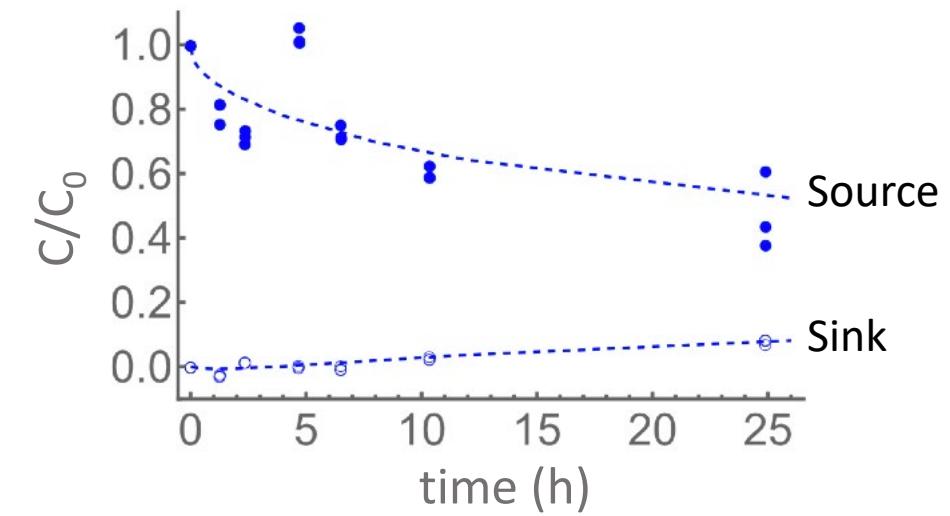
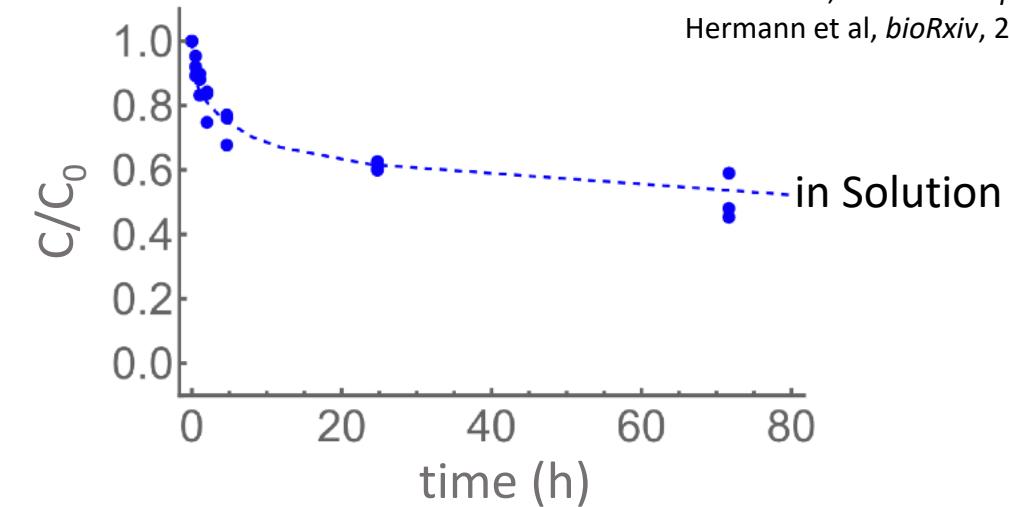
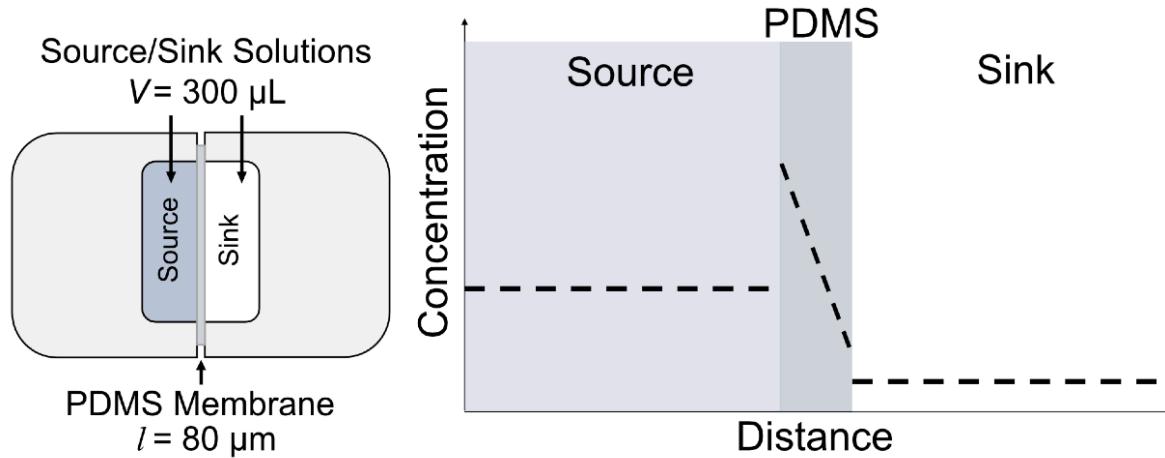
# Measuring Chemical-PDMS Interaction and Transport Parameters

## Disk Soak Experiments



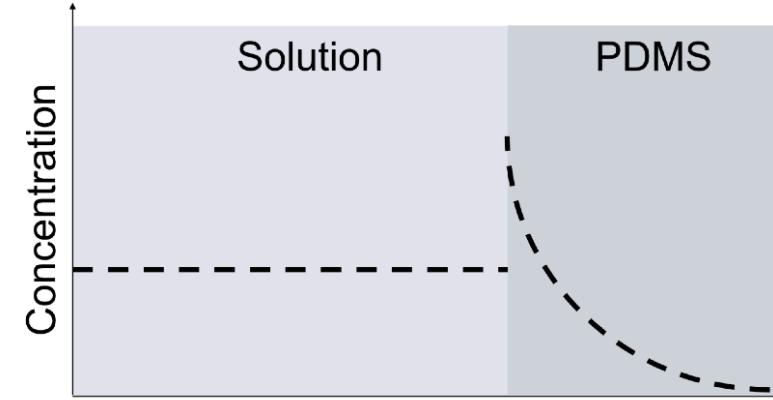
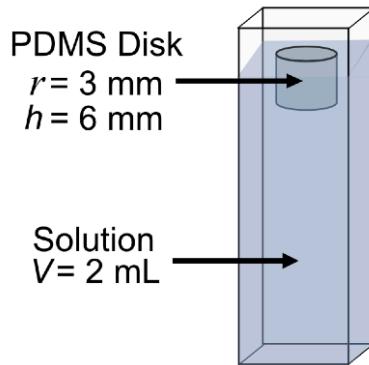
Auner et al, *Lab on a Chip* 2019  
Hermann et al, *bioRxiv*, 2024

## Membrane Experiments

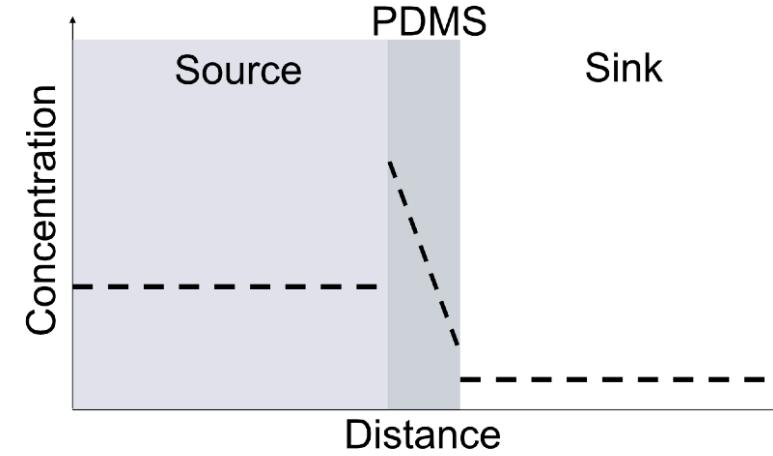
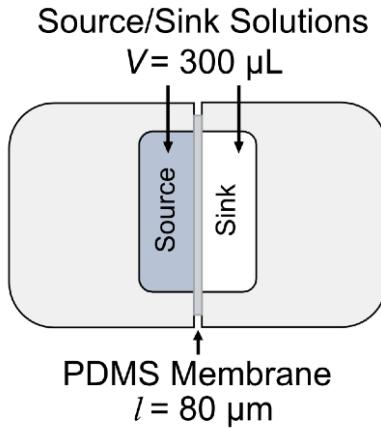


# Measuring Chemical-PDMS Interaction and Transport Parameters

## Disk Soak Experiments



## Membrane Experiments



## Aqueous Solution

$$\text{flux} = -H(kC_{sol} - C_p)$$

$$\frac{\partial C_{sol}}{\partial t} = D_{sol} \frac{\partial^2 C_{sol}}{\partial x^2}$$

$$\frac{\partial C_p}{\partial t} = D_p \frac{\partial^2 C_p}{\partial x^2}$$

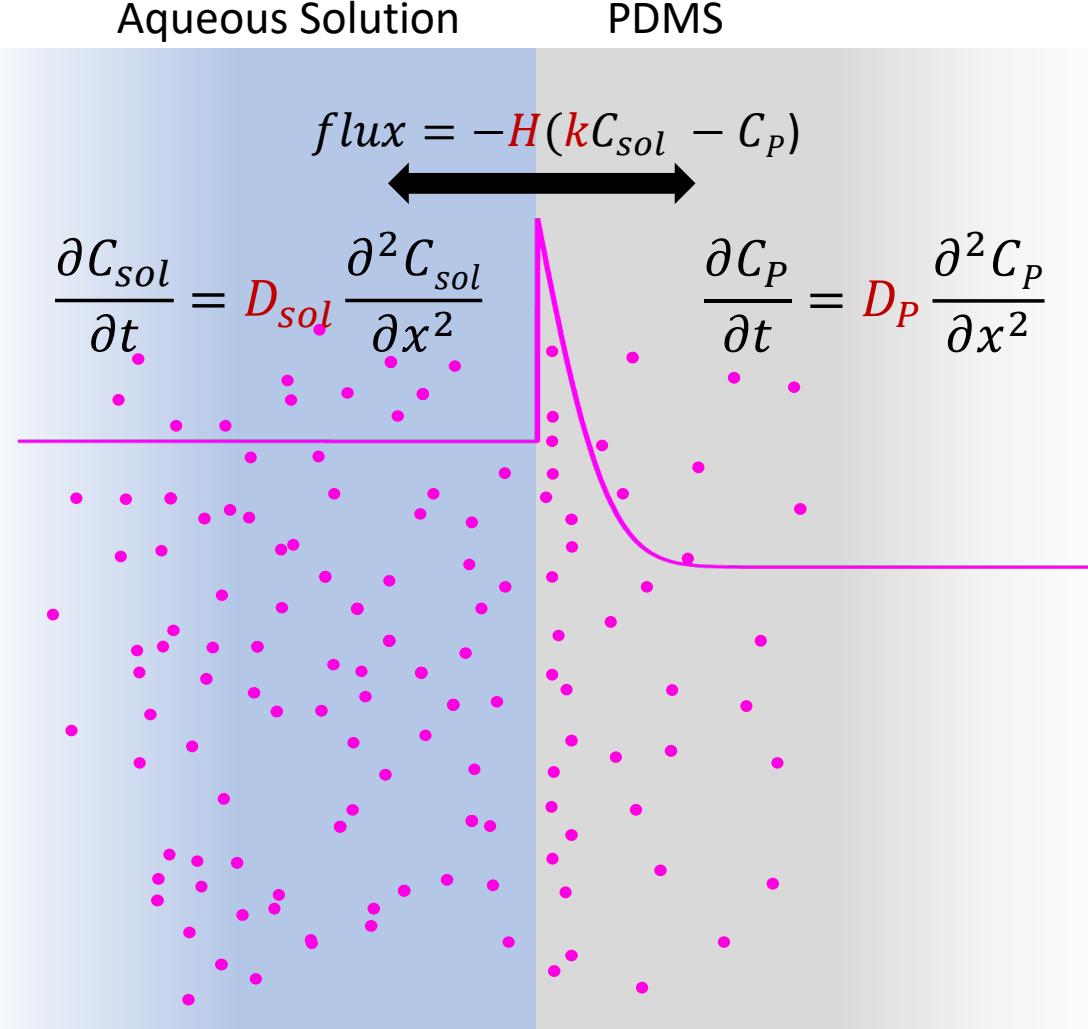


Table 1 CAS numbers, relevance, and selected properties for chemicals tested. Chemical properties sourced from PubChem.<sup>[34]</sup>

Test set of 10 chemicals

Chemical name	CAS number	Relevance	logP	H-Bond Donors	Molar Mass (amu)
Rhodamine 6G	989-38-8	Fluorescent dye	6.4	2	479.0
Benzo[a]pyrene	50-32-8	PAH/Combustion byproduct	6.1	0	252.3
Chlorpyrifos	2921-88-2	Organophosphate pesticide	5.0	0	350.6
Fluorescein-5-isothiocyanate	3326-32-7	Fluorescent dye	4.8	2	389.4
Parathion	56-38-2	Organophosphate pesticide	3.8	0	291.3
Amodiaquine	82-46-0	Pharmaceutical	3.7	2	355.9
Fluorescein	2321-07-5	Fluorescent dye	3.4	2	332.3
Indole	120-72-9	Indole	2.1	1	117.2
Paraoxon	311-45-5	Organophosphate pesticide metabolite	2.0	0	275.2
Rhodamine B	81-88-9	Fluorescent dye	1.9	1	479.0

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Test set of 10 chemicals

7 were sufficiently soluble to perform experiments directly in phosphate-buffered saline

Chemical name	CAS number	Relevance	logP	H-Bond Donors	Molar Mass (amu)
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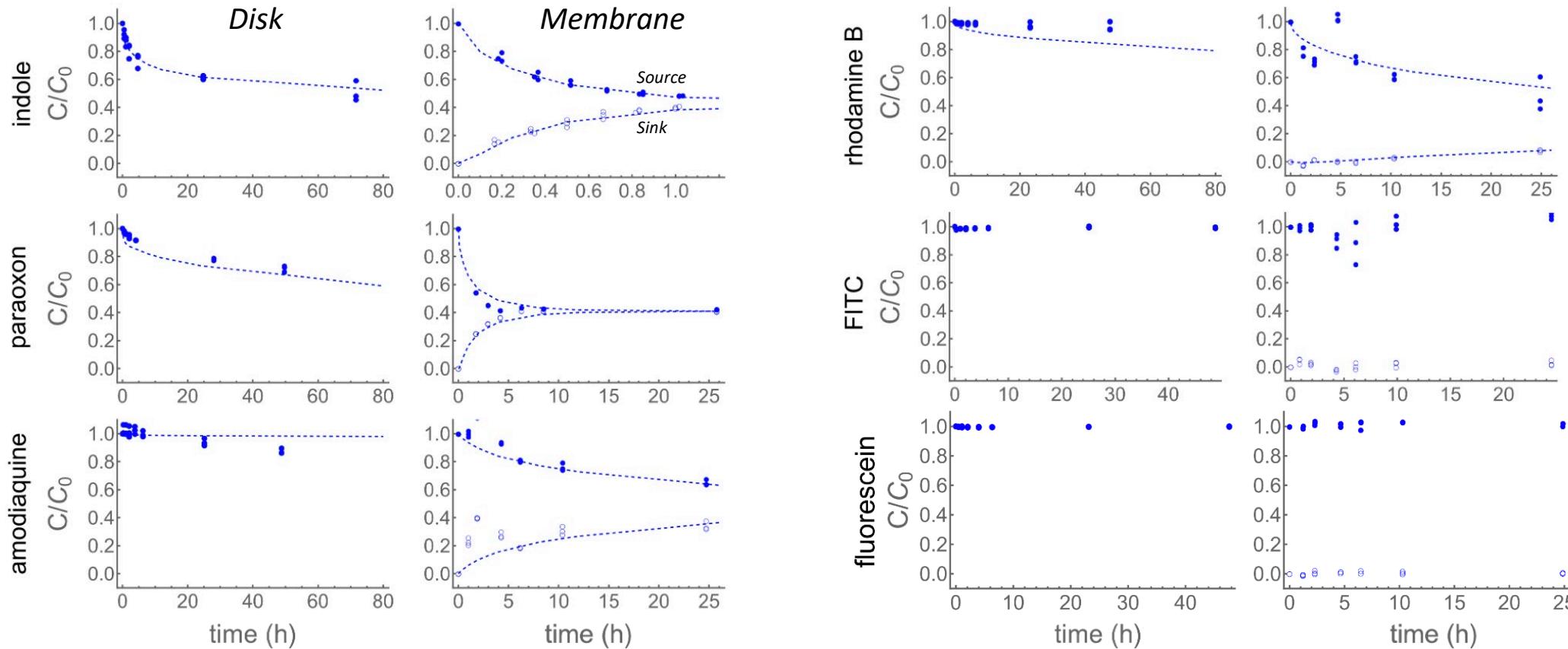


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Test set of 10 chemicals

3 required the addition of DMSO as a cosolvent

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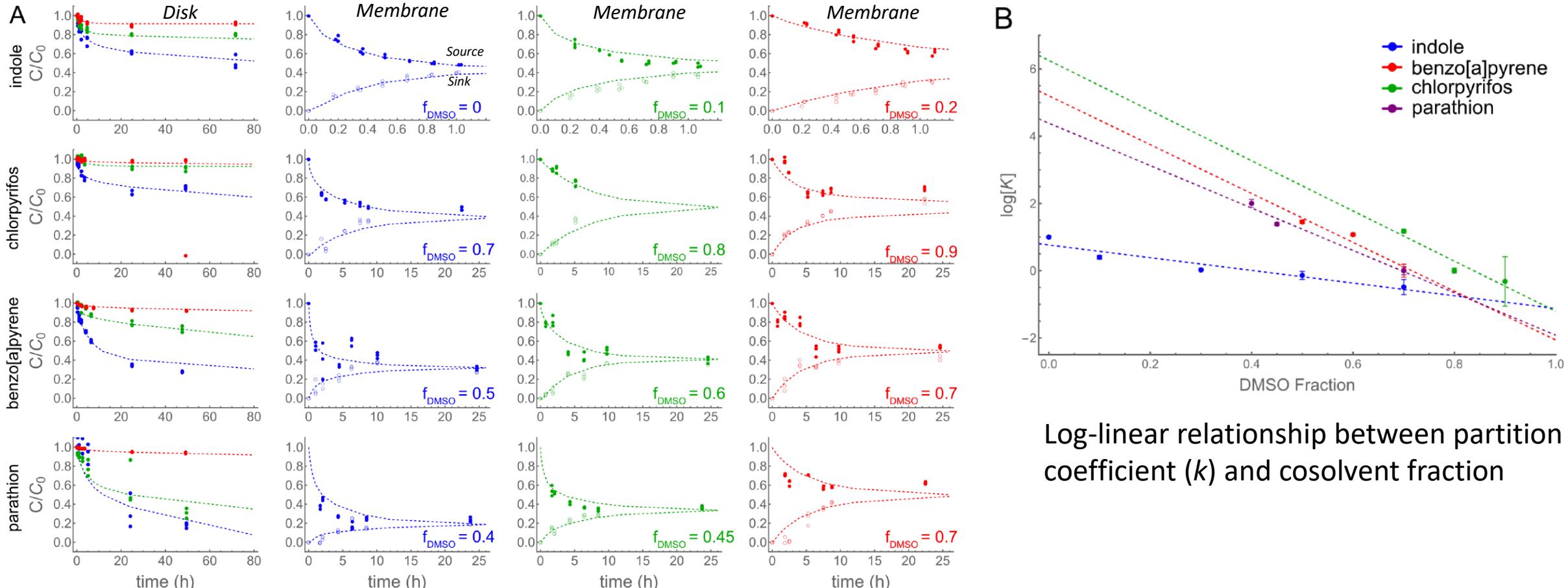
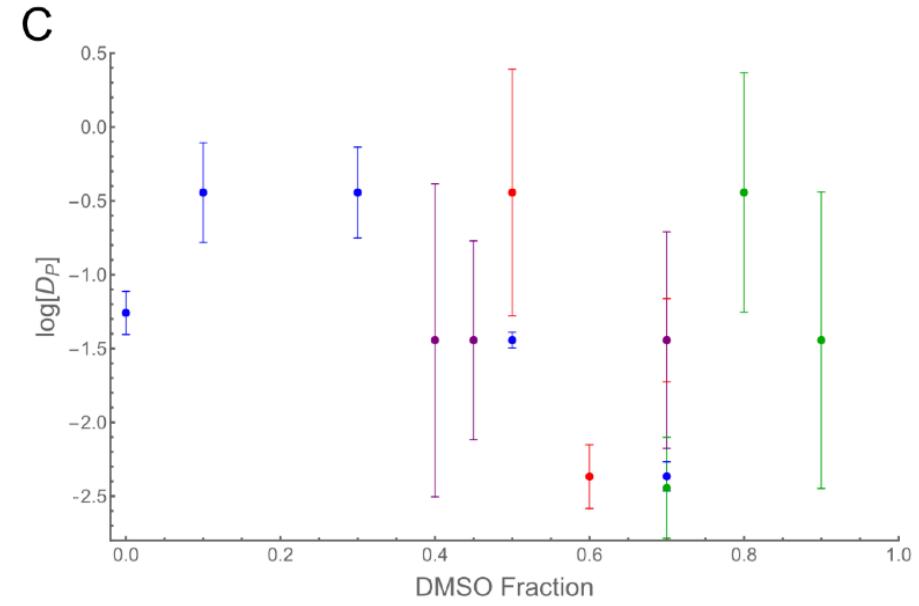
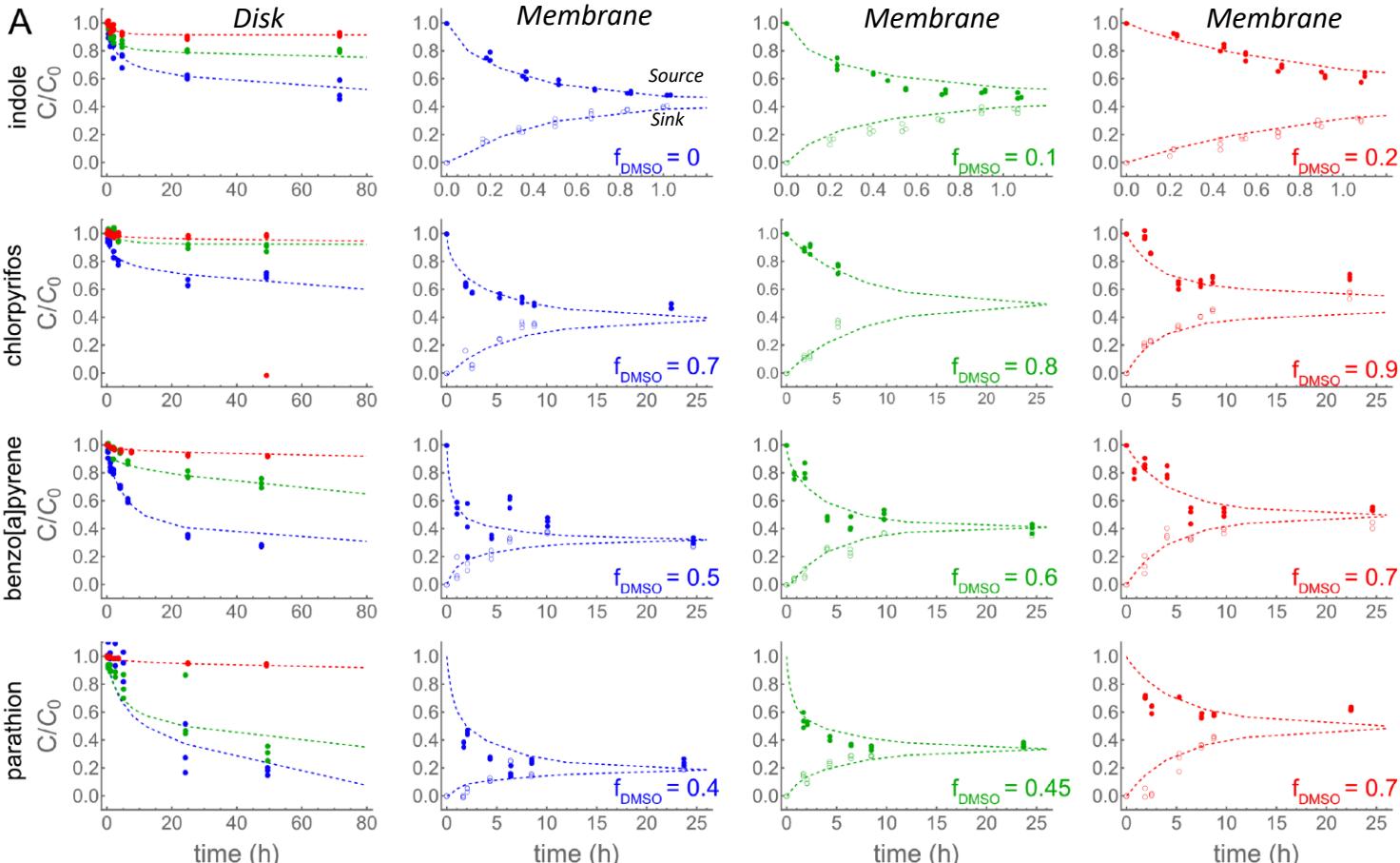


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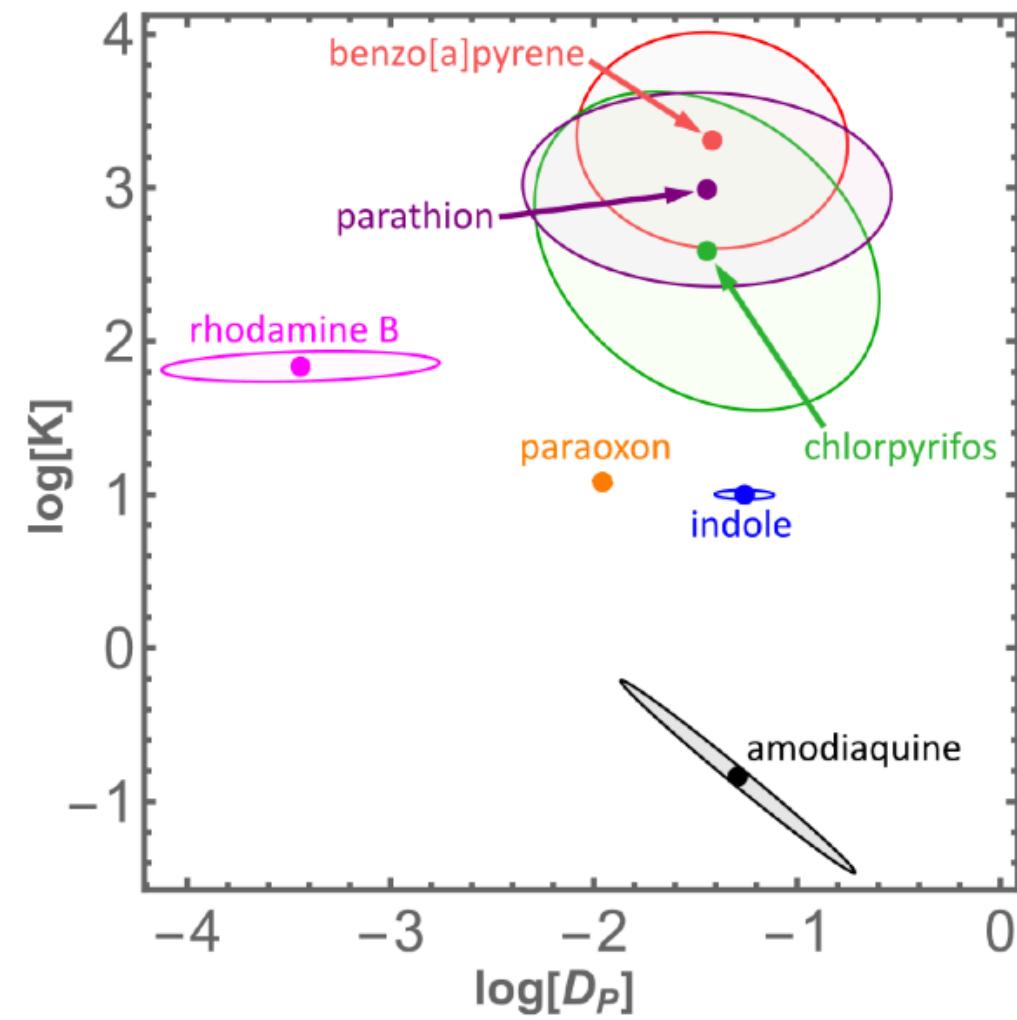
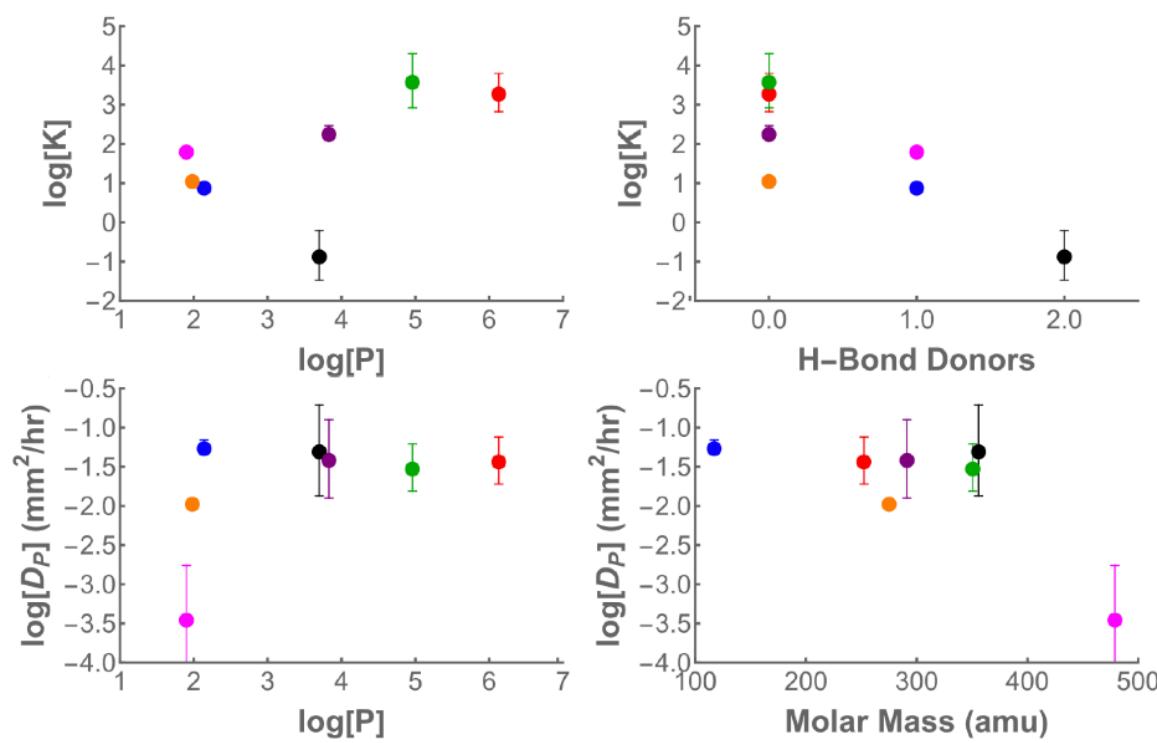


No clear relationship between diffusion constant in PDMS ( $D_P$ ) and cosolvent fraction

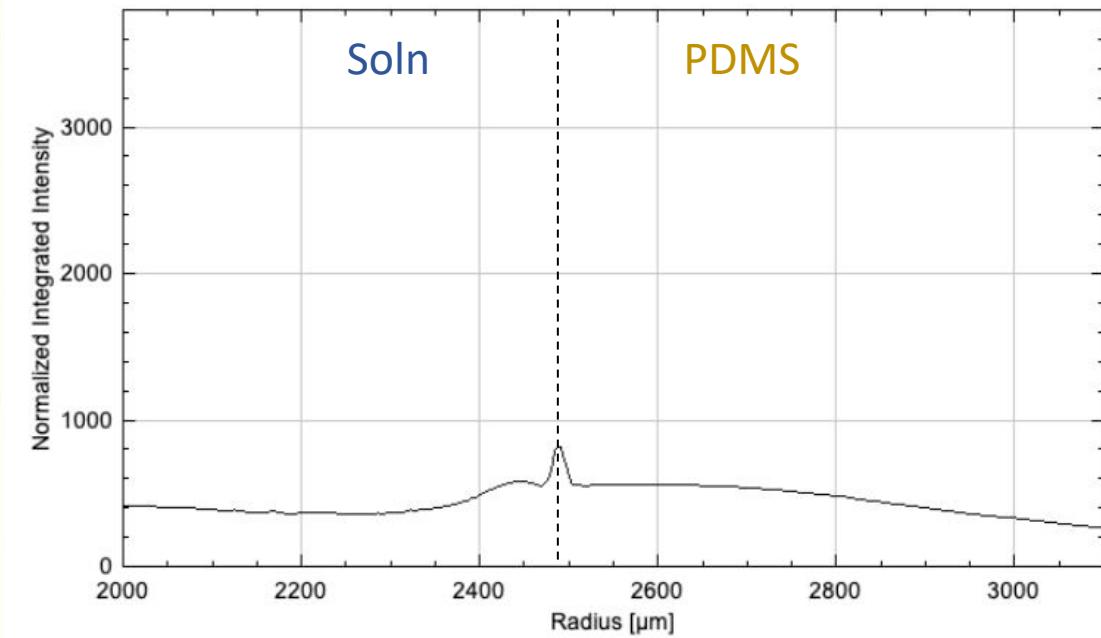
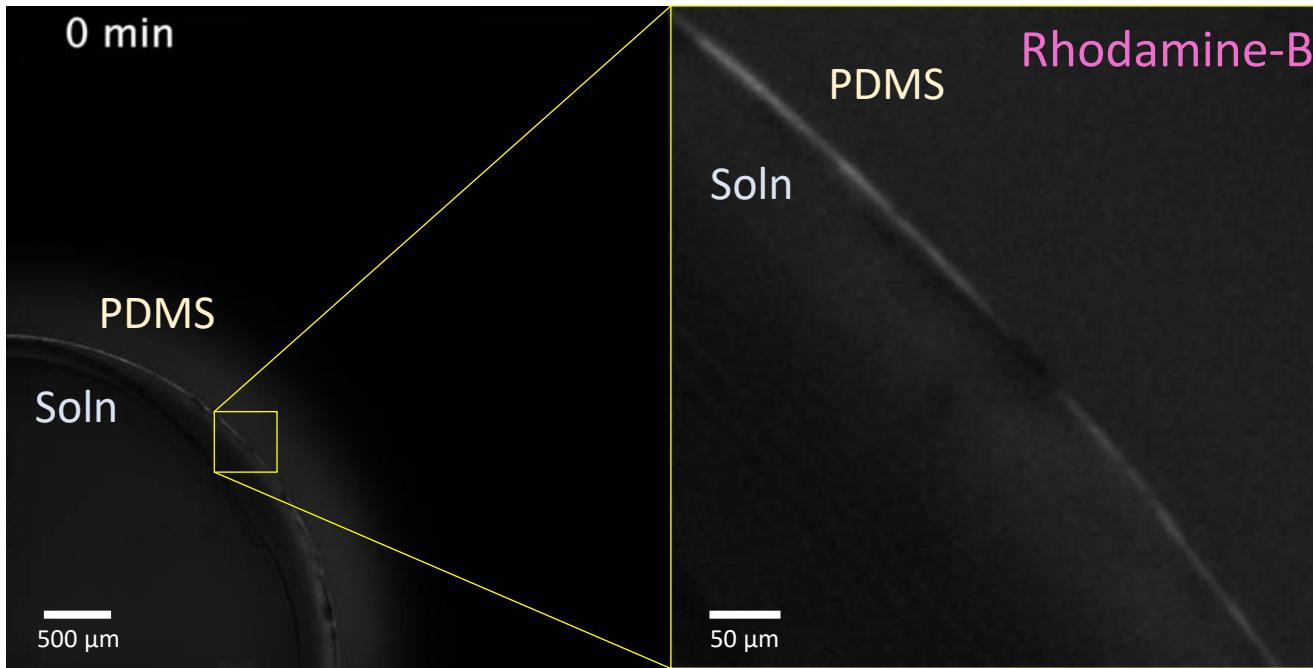
# The best-fit chemical-PDMS interaction parameters span several orders of magnitude

Table 2 Best-fit parameter values for the partition-diffusion model

Chemical	$\log K_{PW}$	$\log K_{PD}$	$\log D_P$ ( $\text{mm}^2 \text{ h}^{-1}$ )
Chlorpyrifos	$6.25 \pm 1.98$	$-1.21 \pm 0.53$	$-1.51 \pm 0.30$
Benzo[a]pyrene	$5.20 \pm 1.22$	$-2.06 \pm 0.82$	$-1.42 \pm 0.30$
Parathion	$4.39 \pm 0.50$	$-1.92 \pm 0.47$	$-1.40 \pm 0.50$
Rhodamine B	$1.83 \pm 0.10$	–	$-3.44 \pm 0.68$
Paraoxon	$1.08 \pm 0.03$	–	$-1.96 \pm 0.05$
Indole	$0.91 \pm 0.03$	$-1.12 \pm 0.26$	$-1.25 \pm 0.09$
Amodiaquine	$-0.84 \pm 0.63$	–	$-1.29 \pm 0.58$

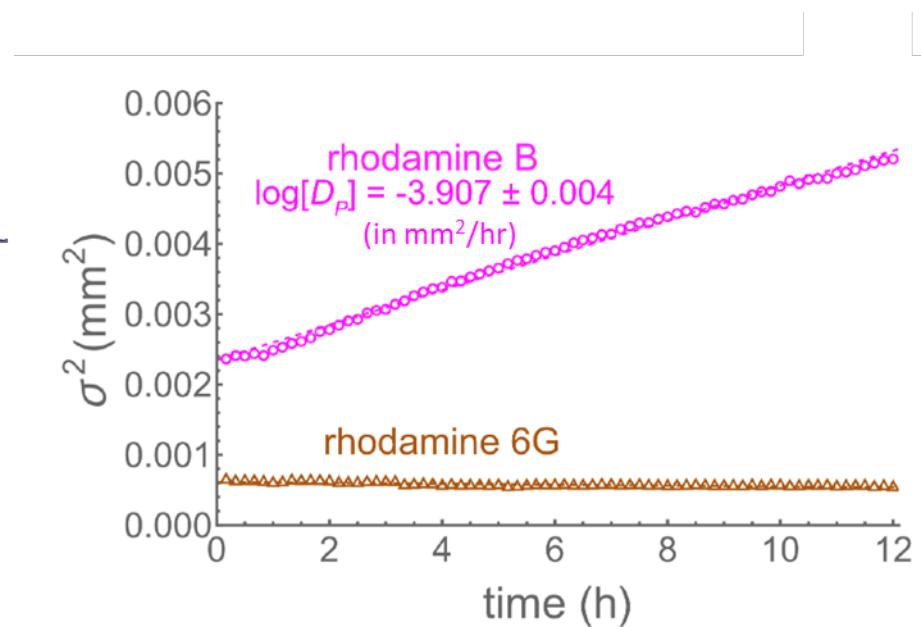
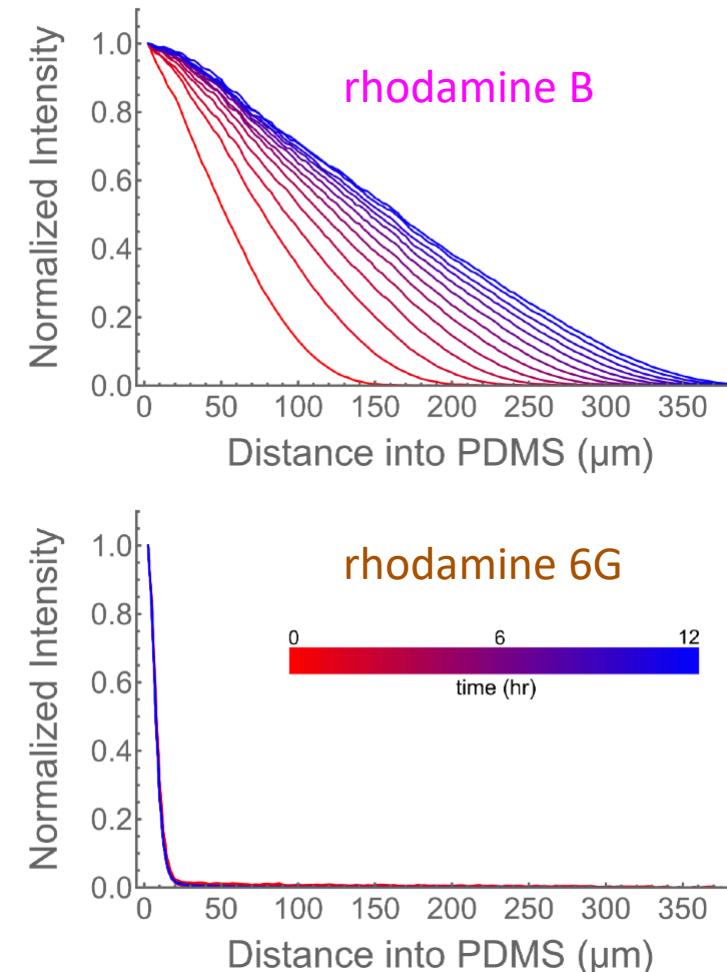
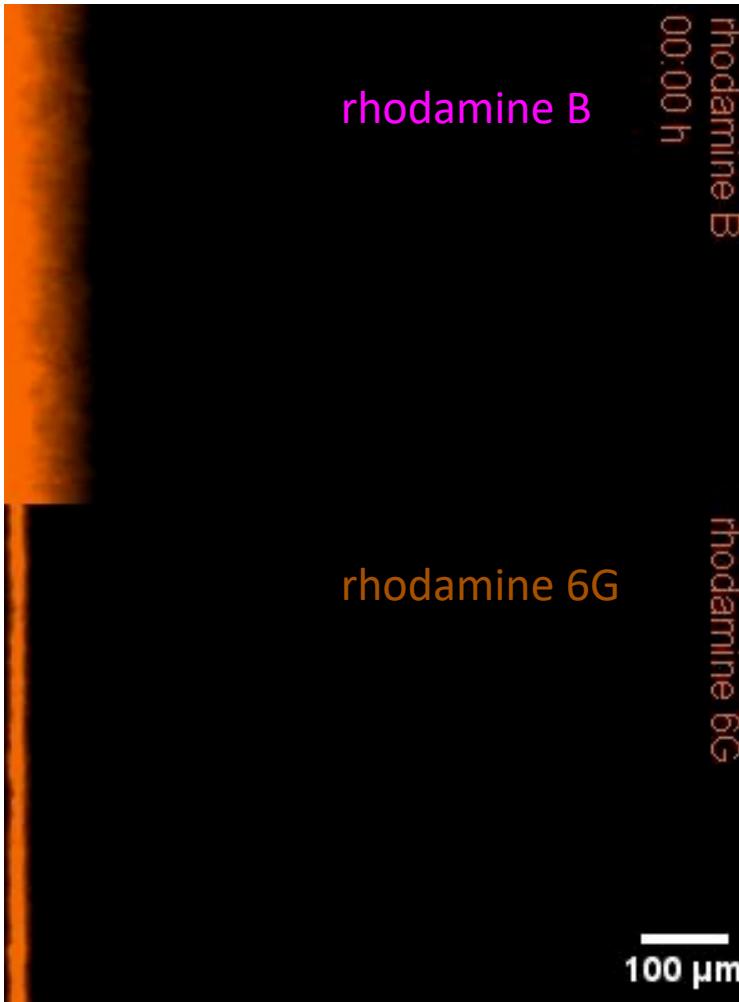


## Validate $D_P$ by visualizing diffusion into the PDMS bulk



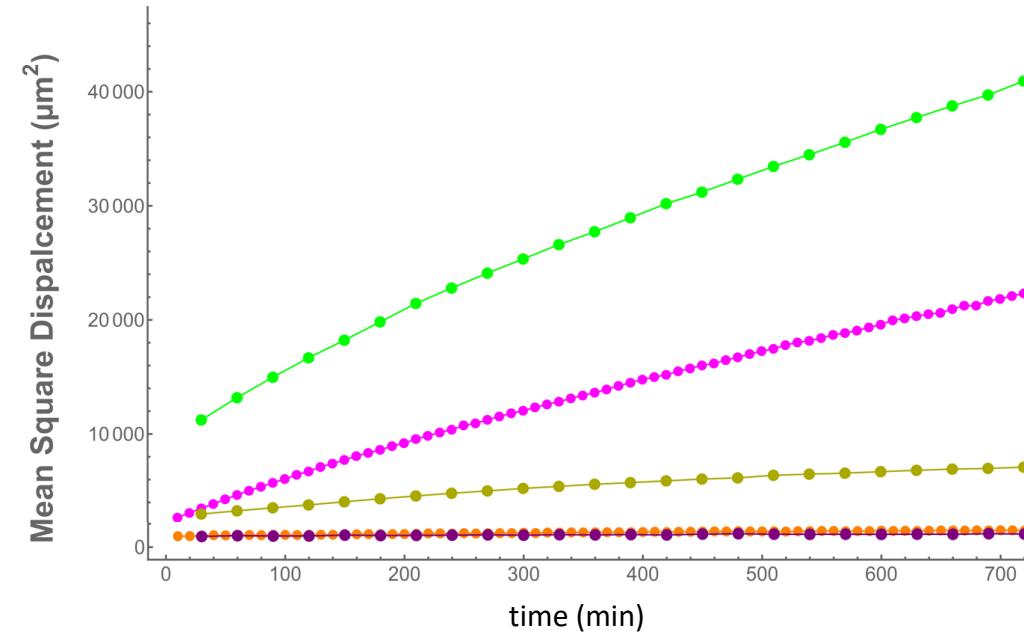
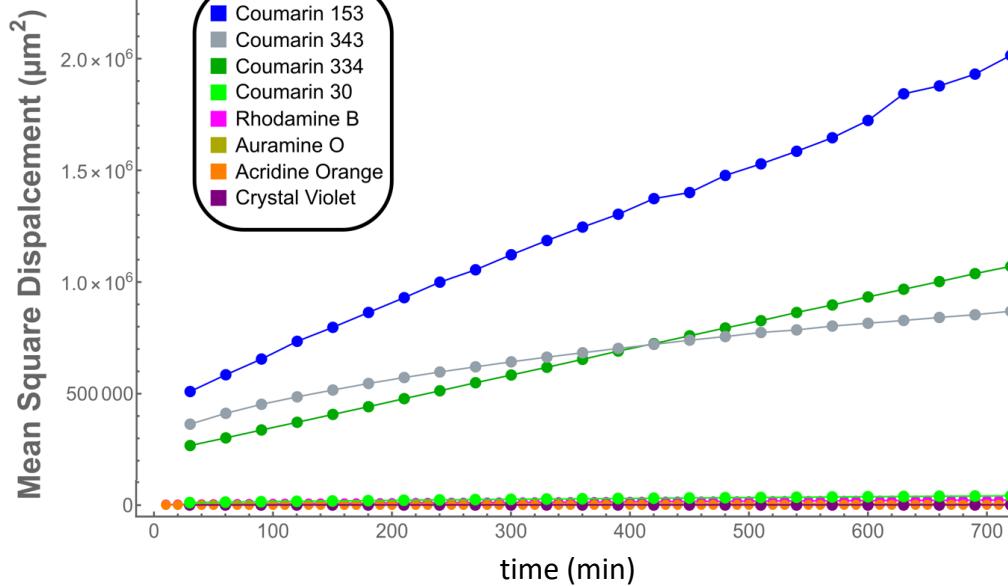
Spreads  $\sim 30 \mu\text{m}$  into PDMS within 3 hours:  $D_P \sim \frac{l^2}{t} \sim 300 \mu\text{m}^2/\text{h} \sim 3 \times 10^{-4} \text{ mm}^2/\text{h}$   
 $\log[D_P] \sim -3.5$

# Validate $D_p$ by visualizing diffusion into the PDMS bulk



Result from disk/membrane experiments for rhodamine B:  
 $\log[D_p] = -3.44 \pm 0.68$  (in  $\text{mm}^2/\text{hr}$ )

# Tracking additional dyes: $D_P$ can vary over 4 orders of magnitude



Chemical name	$\log D_P$ (in $\text{mm}^2/\text{hr}$ )
Coumarin 153	-1.192
Coumarin 334	-1.461
Coumarin 343	-1.562
Coumarin 30	-2.838
Rhodamine B	-3.042
Auramine O	-3.683
Acridine Orange	-4.536
Crystal Violet	-4.912

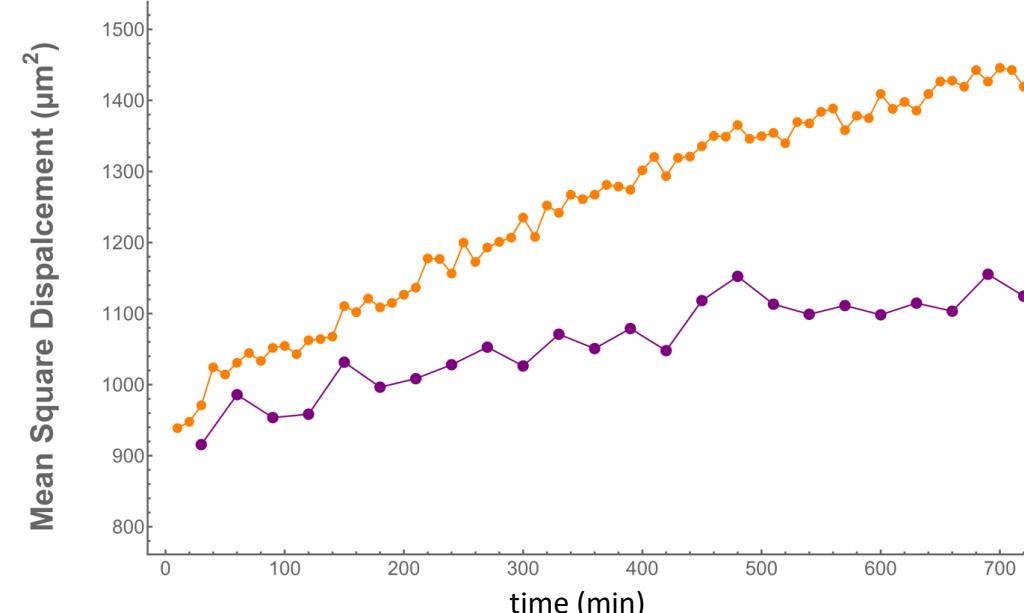


Table 1: Fluorescent dyes and selected properties. (\*) Calculated using RDKit [13]. All other properties sourced from PubChem [14]

Chemical name	logP	Mass (amu)	H-Bond Donors	H-Bond Acceptors	TPSA (Å <sup>2</sup> )
Rhodamine 6G	6.4	479.0	2	5	61.5
X-34	5.9	402.4	4	6	115
Eosin B Diphenol	5.6	580.1	2	9	168
Fluorescein-5-isothiocyanate	4.8	389.4	2	7	120
Coumarin 30	4.0	347.4	0	4	47.4
Auramine O	3.7*	303.8	2	3	30.3
Fluorescein	3.4	332.3	2	5	76
Acridine Orange	3.4	265.35	0	3	19.4
Coumarin 153	3.2	309.28	0	6	29.5
Coumarin 343	3.1	285.29	1	5	66.8
Coumarin 334	2.7	283.32	0	4	46.6
Rhodamine B	1.9	479.0	1	5	52.8
Crystal Violet	1.5*	408.0	0	3	9.5
Merocyanine 540	0.7*	569.7	0	8	151

Dyes without a color-code did not diffuse into PDMS (but some bound to surface)

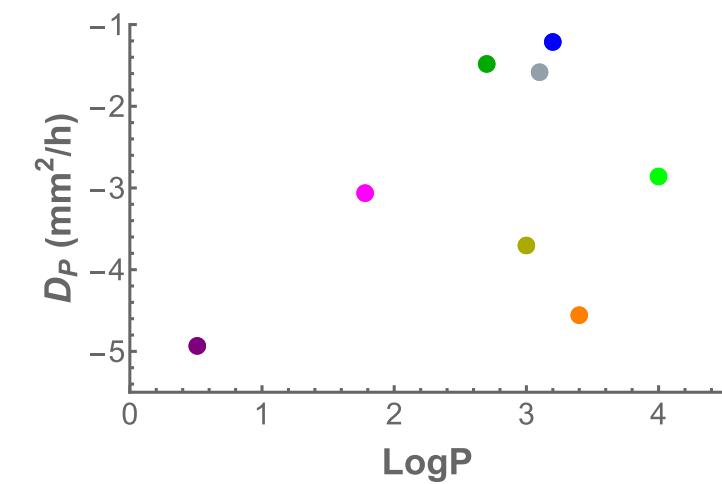
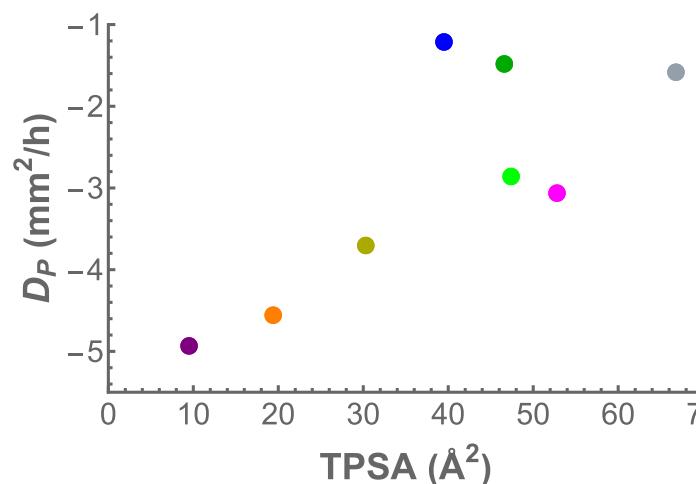
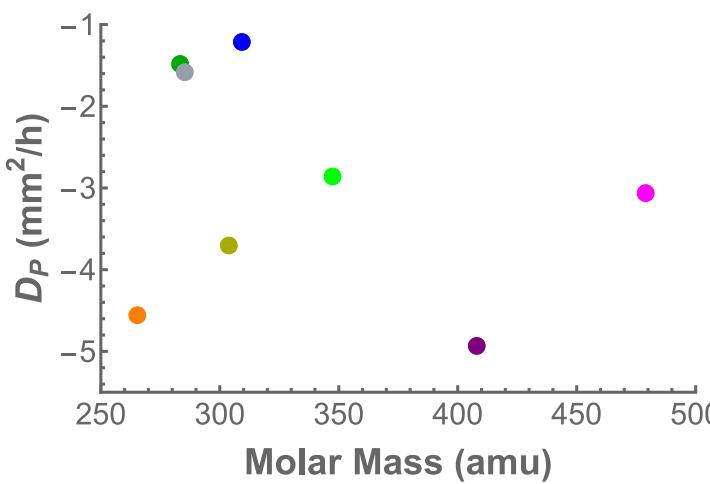
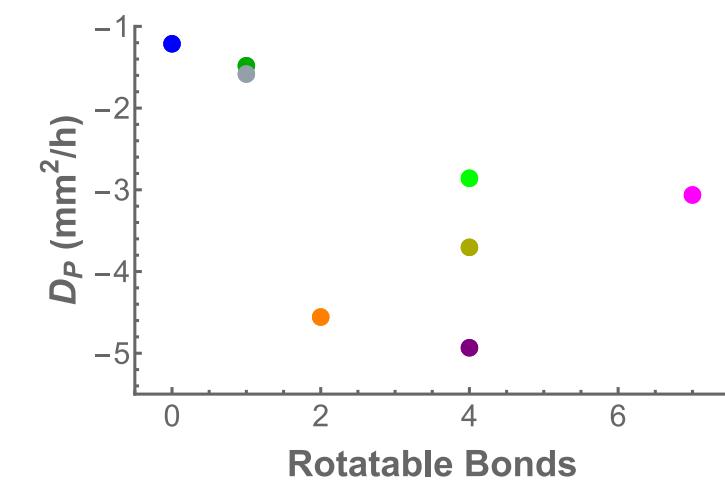
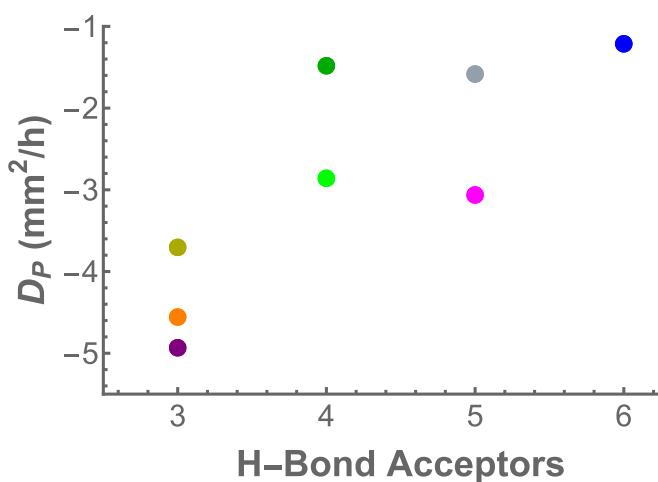
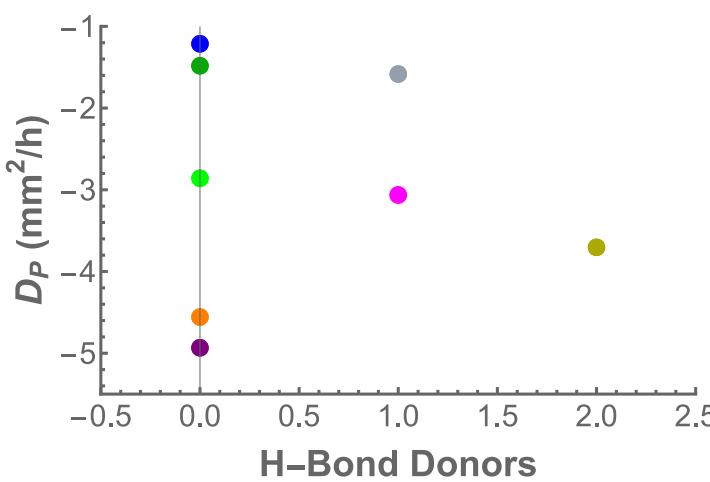


Table 1: Fluorescent dyes and selected properties. (\*) Calculated using RDKit [13]. All other properties sourced from PubChem [14]

Chemical name	logP	Mass (amu)	H-Bond Donors	H-Bond Acceptors	TPSA (Å <sup>2</sup> )
Rhodamine 6G	6.4	479.0	2	5	61.5
X-34	5.9	402.4	4	6	115
Eosin B Diphenol	5.6	580.1	2	9	168
Fluorescein-5-isothiocyanate	4.8	389.4	2	7	120
Coumarin 30	4.0	347.4	0	4	47.4
Auramine O	3.7*	303.8	2	3	30.3
Fluorescein	3.4	332.3	2	5	76
Acridine Orange	3.4	265.35	0	3	19.4
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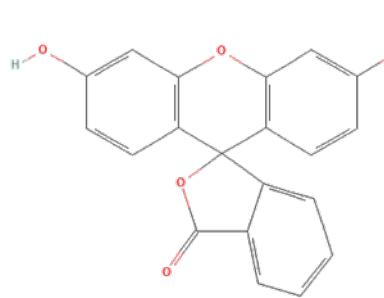
Dyes without a color-code did not diffuse into PDMS (but some bound to surface)



# Talk Outline

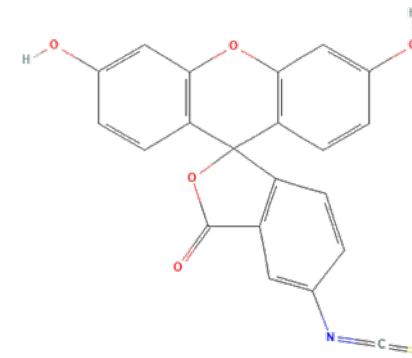
1. Measure chemical-PDMS interaction and transport parameters.
  - a. Partition coefficients:  $k = C_{PDMS}/C_{sol}$
  - b. Diffusion coefficients in PDMS:  $D_P$
  - c. Evaluate correlations with chemical properties
2. Be wary of read-across methods.
3. Validate multiphysics models for in-device toxicokinetics.
4. Investigate the effectiveness of mitigation strategies for reducing chemical sequestration in microfluidic devices:
  - a. SEBS co-polymers as PDMS alternatives; and
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## Some chemically similar pairs do have similar PDMS interactions.



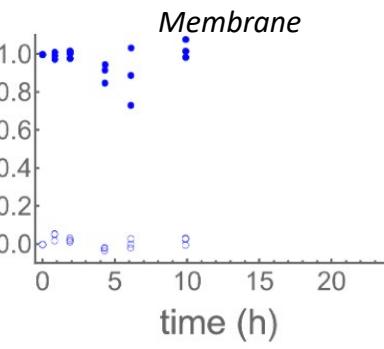
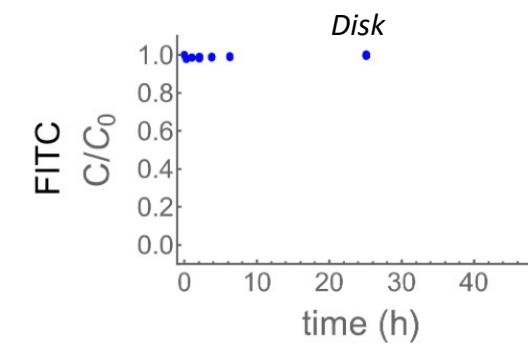
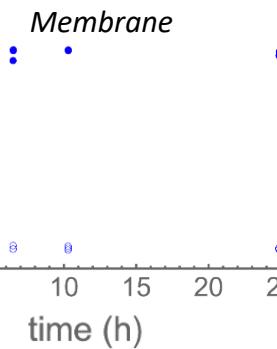
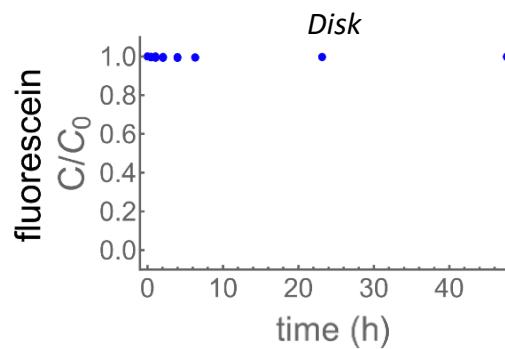
**Fluorescein**

MW = 332.3 amu  
 LogP = 3.4  
 H-Bond Donors = 2  
 TPSA = 76 Å



**Fluorescein-5-isothiocyanate (FITC)**

MW = 389.4 amu  
 LogP = 4.8  
 H-Bond Donors = 2  
 TPSA = 120 Å



Both are quite hydrophobic,  
 but neither interacts measurably with PDMS.

# Similar chemicals can interact with PDMS quite differently!

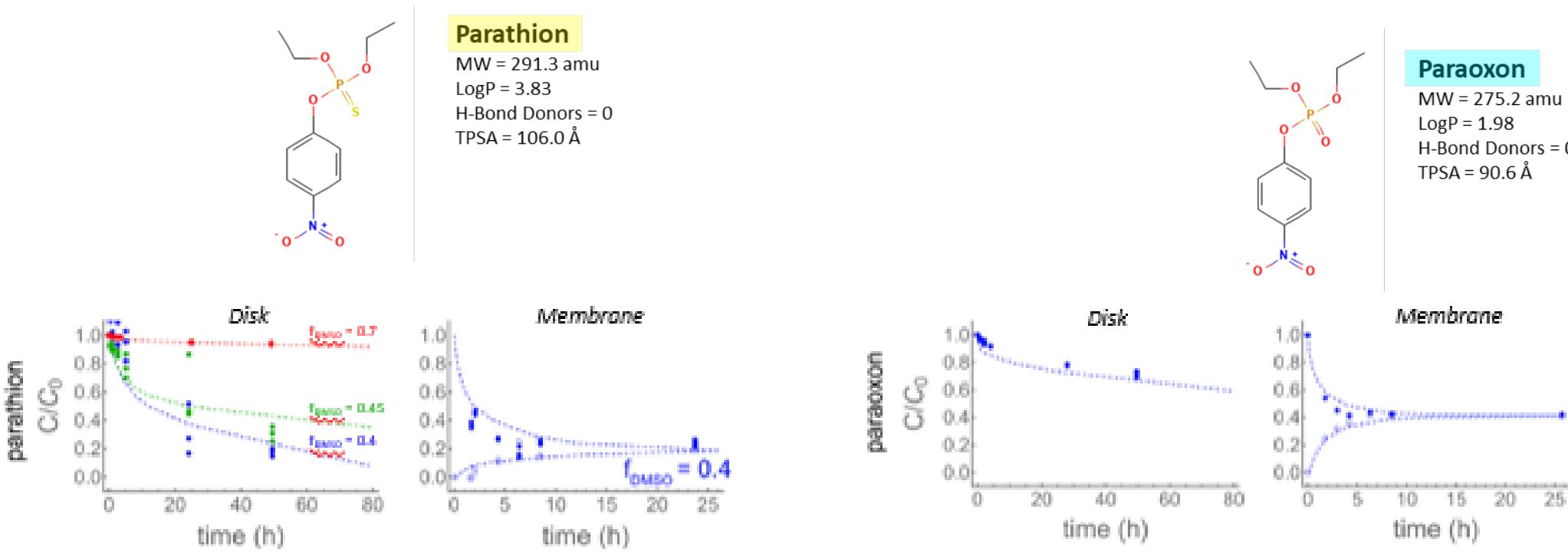


Table 2 Best-fit parameter values for the partition-diffusion model

Chemical	$\log K_{PW}$	$\log K_{PP}$	$\log D_P$ ( $\text{mm}^2 \text{ h}^{-1}$ )
Chlorpyrifos	$6.25 \pm 1.98$	$-1.21 \pm 0.53$	$-1.51 \pm 0.30$
Benzo[a]pyrene	$5.20 \pm 1.22$	$-2.06 \pm 0.82$	$-1.42 \pm 0.30$
Parathion	$4.39 \pm 0.50$	$-1.92 \pm 0.47$	$-1.40 \pm 0.50$
Rhodamine B	$1.83 \pm 0.10$	-	$-3.44 \pm 0.68$
Paraoxon	$1.08 \pm 0.03$	-	$-1.96 \pm 0.05$
Indole	$0.91 \pm 0.03$	$-1.12 \pm 0.26$	$-1.25 \pm 0.09$
Amodiaquine	$-0.84 \pm 0.63$	-	$-1.29 \pm 0.58$

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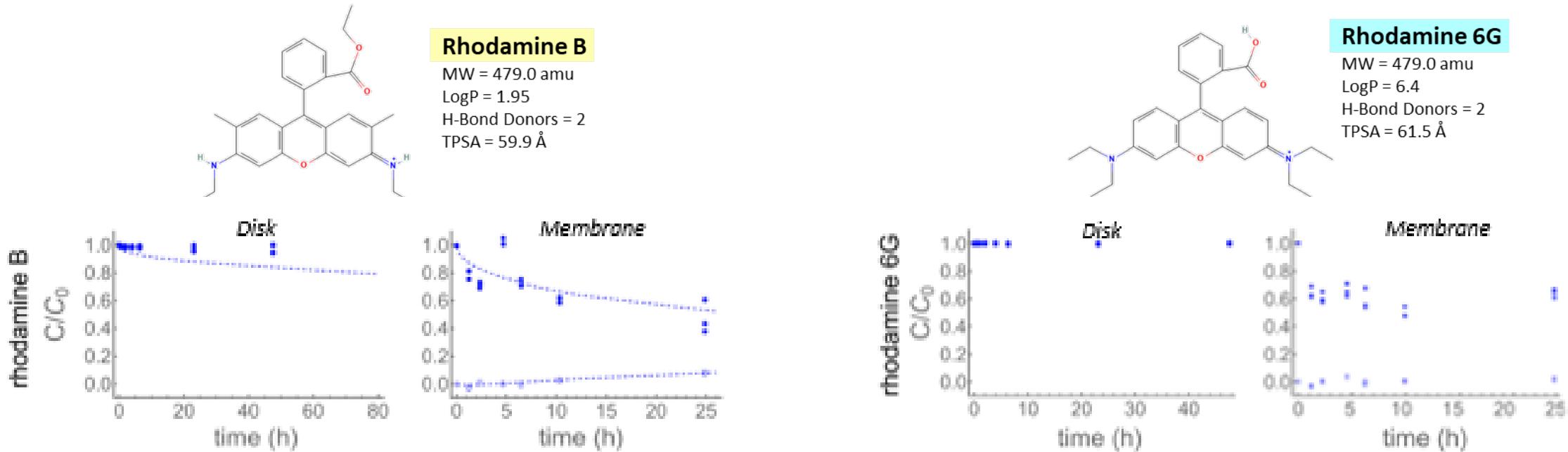


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<b>Rhodamine B</b>	<b><math>1.83 \pm 0.10</math></b>	<b>—</b>	<b><math>-3.44 \pm 0.68</math></b>
Paraoxon	$1.08 \pm 0.03$	—	$-1.96 \pm 0.05$
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Amodiaquine	$-0.84 \pm 0.63$	—	$-1.29 \pm 0.58$

Rhodamine 6G binds to PDMS surfaces, but does not diffuse into PDMS bulk.

# Similar chemicals can interact with PDMS quite differently!

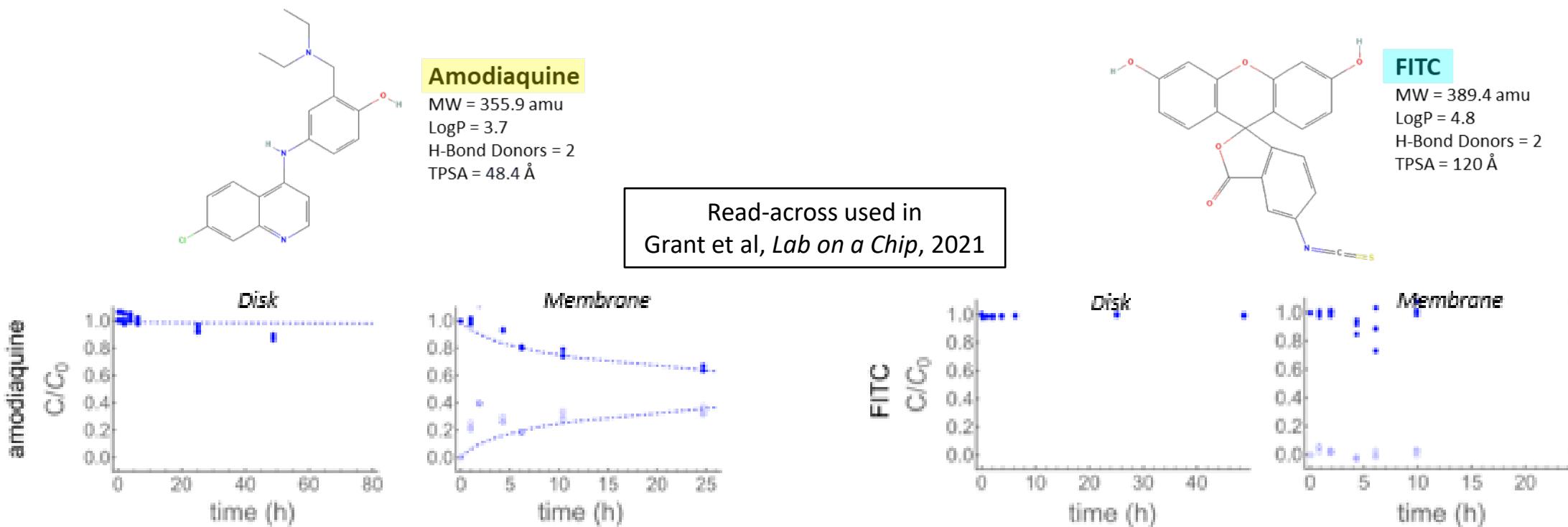


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<b>Amodiaquine</b>	<b><math>-0.84 \pm 0.63</math></b>	-	<b><math>-1.29 \pm 0.58</math></b>

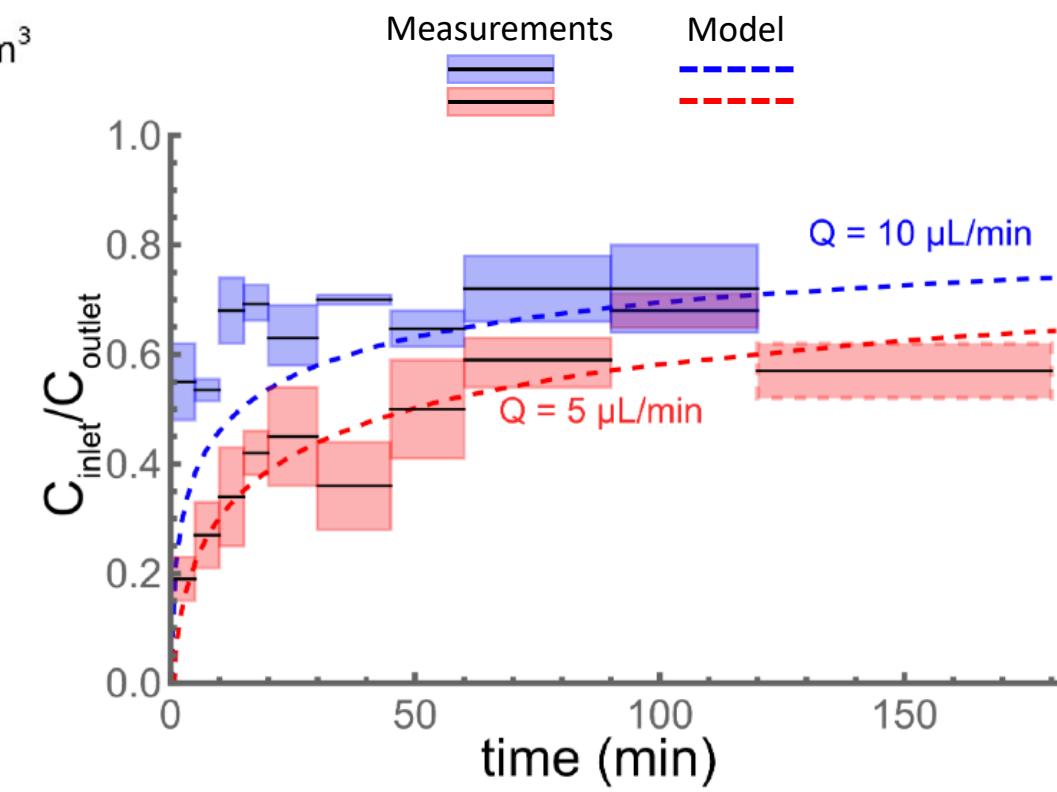
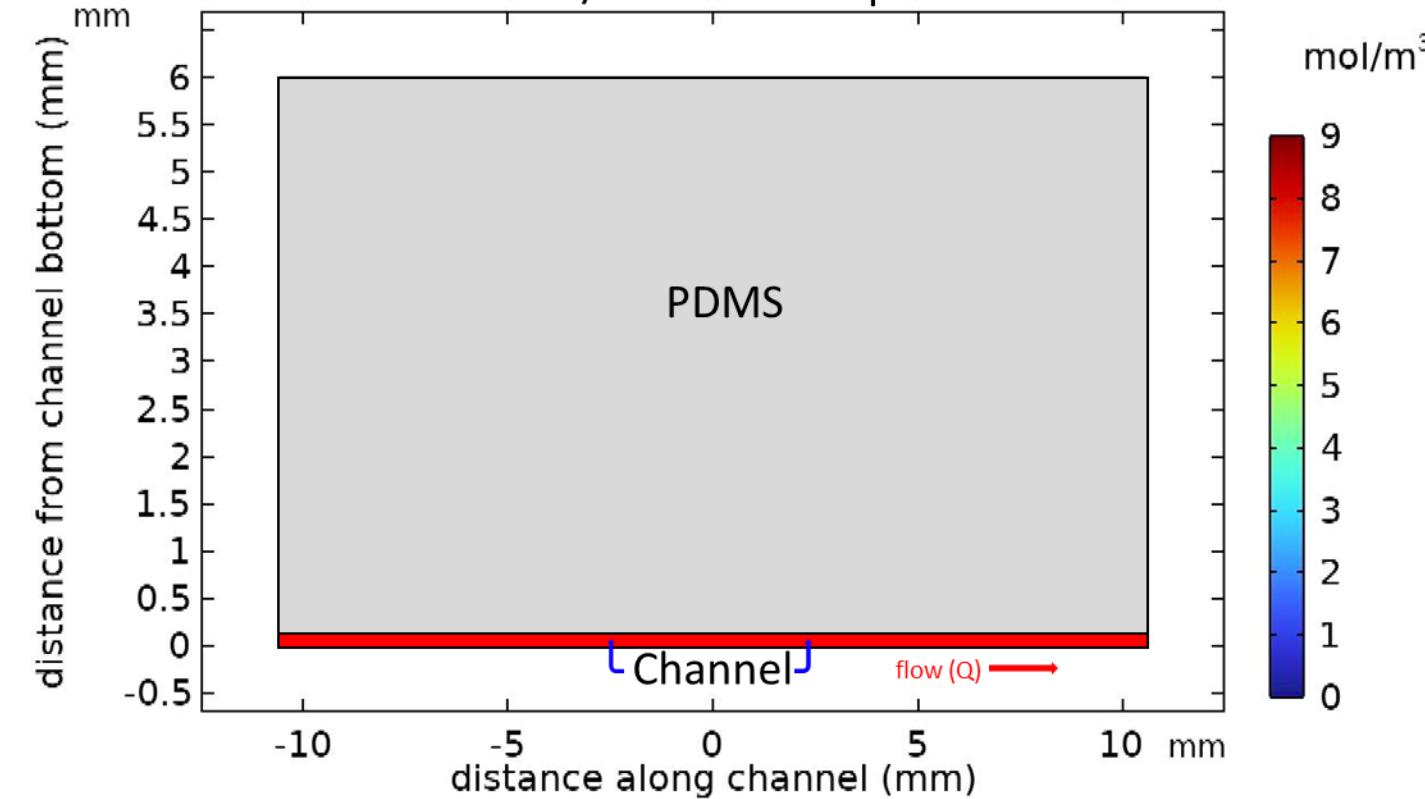
FITC does not measurably bind to, partition into or diffuse through PDMS.

# Talk Outline

1. Measure chemical-PDMS interaction and transport parameters.
  - a. Partition coefficients:  $k = C_{PDMS}/C_{sol}$
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  - c. Evaluate correlations with chemical properties
2. Be wary of read-across methods.
3. **Validate multiphysics models for in-device toxicokinetics.**
4. Investigate the effectiveness of mitigation strategies for reducing chemical sequestration in microfluidic devices:
  - a. SEBS co-polymers as PDMS alternatives; and
  - b. media with carrier proteins.

# Modelling chemical distribution during flow through a microfluidic channel

COMSOL model based on parameters  
from disk/membrane experiments

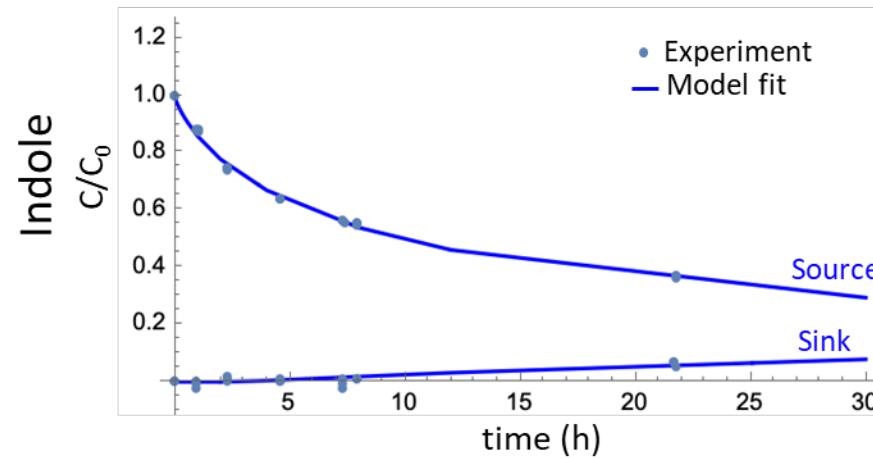


# Talk Outline

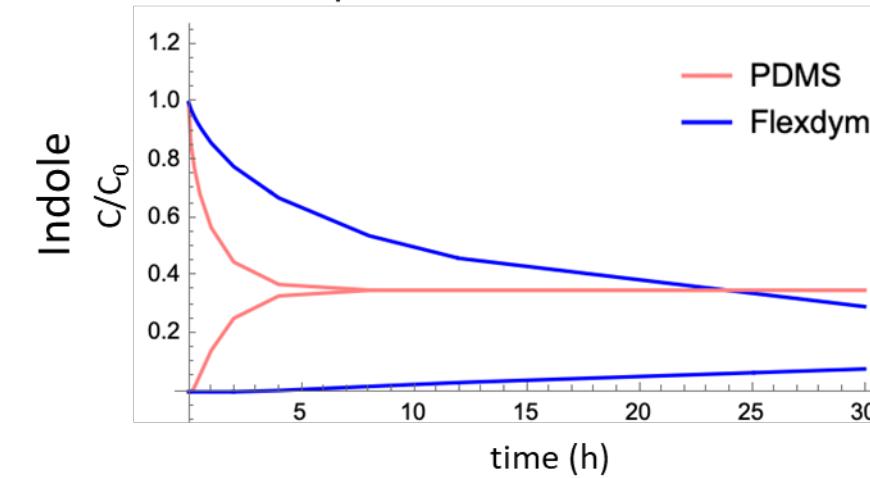
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Chemicals still partition into Flexdym™ (SEBS), but diffuse through it more slowly.

Indole diffusing into and across a 250- $\mu\text{m}$  thick Flexdym™ membrane



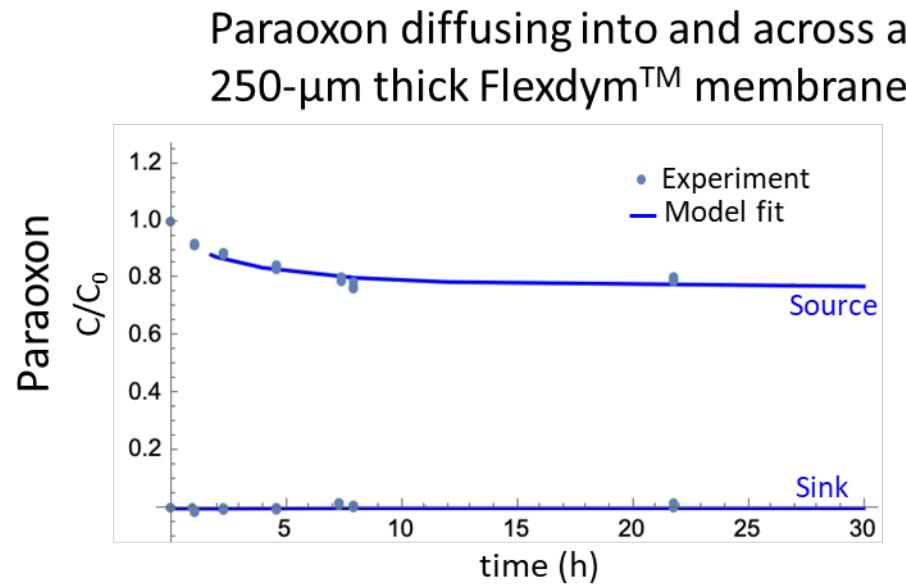
Compared to predictions for 250- $\mu\text{m}$  thick PDMS membrane



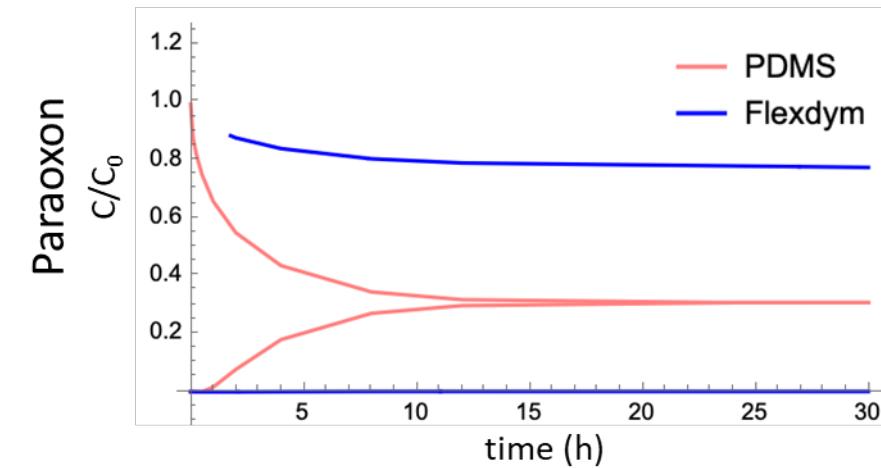
Indole's interaction with	$\log[k]$	$\log[D_p]$ in $\text{mm}^2/\text{h}$
Flexdym™	$1.60 \pm 0.03$	$-2.57 \pm 0.11$
PDMS	$0.91 \pm 0.03$	$-1.25 \pm 0.09$

Greater partition coefficient for Flexdym™,  
but with slower diffusion.

Chemicals still partition into Flexdym™ (SEBS), but diffuse through it more slowly.



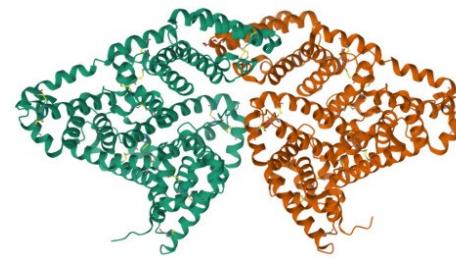
Compared to predictions for 250- $\mu\text{m}$  thick PDMS membrane



Paraoxon's interaction with	$\log[k]$	$\log[D_p]$ in $\text{mm}^2/\text{h}$
Flexdym™	$1.19 \pm 0.36$	$-6 \pm 19$
PDMS	$1.08 \pm 0.03$	$-1.96 \pm 0.05$

Similar partition coefficient for Flexdym™, but with no detectable diffusion.

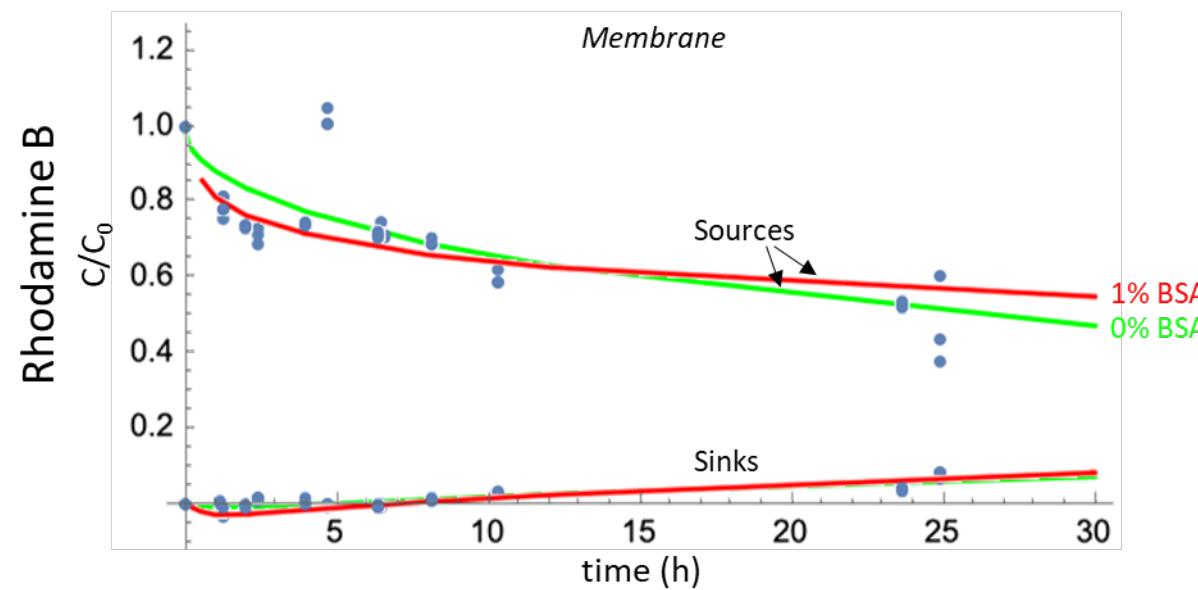
## Adding BSA to media can reduce partitioning into PDMS



Bovine serum albumin

PDB DOI: <https://doi.org/10.2210/pdb4F5S/pdb>

Mol Wt = 66,433 Da  $\longrightarrow$  1% w/v = 15 mM



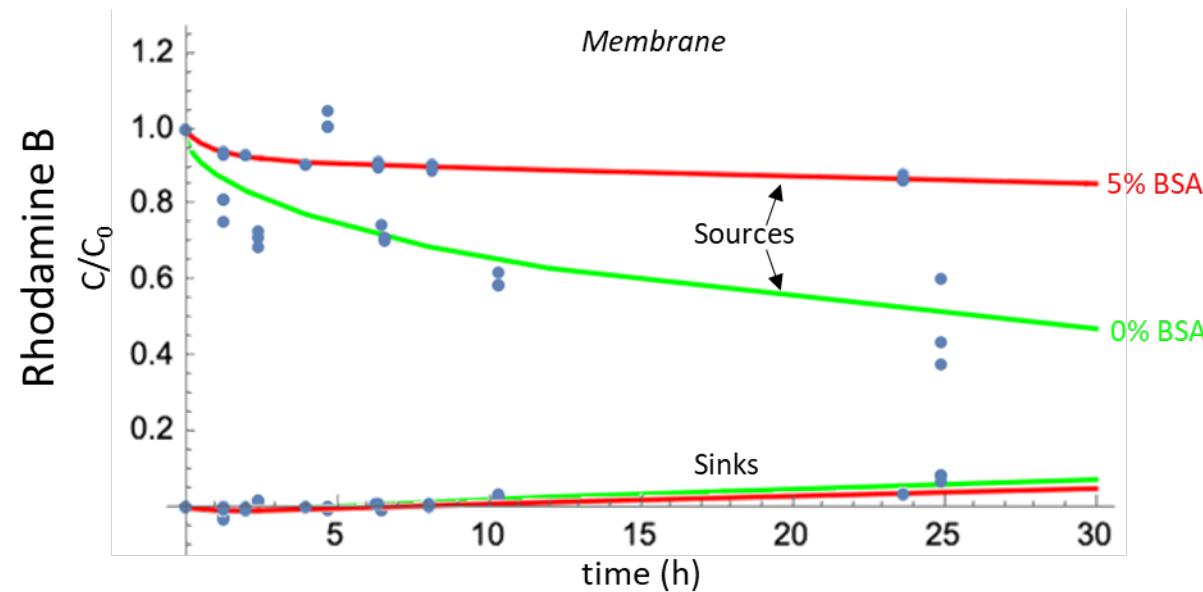
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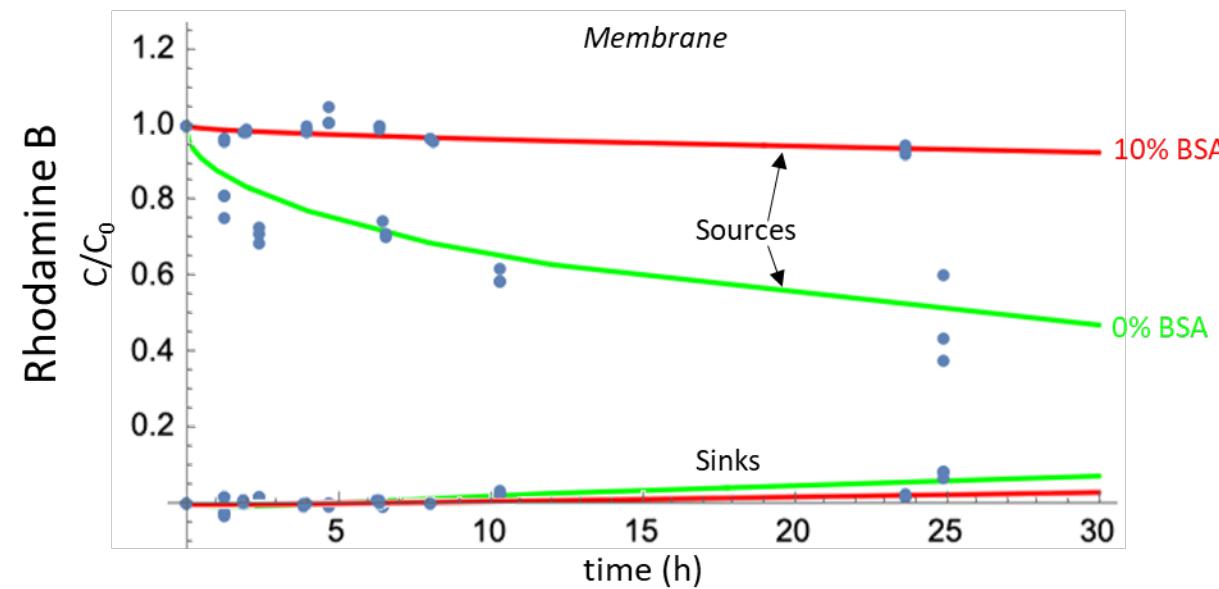
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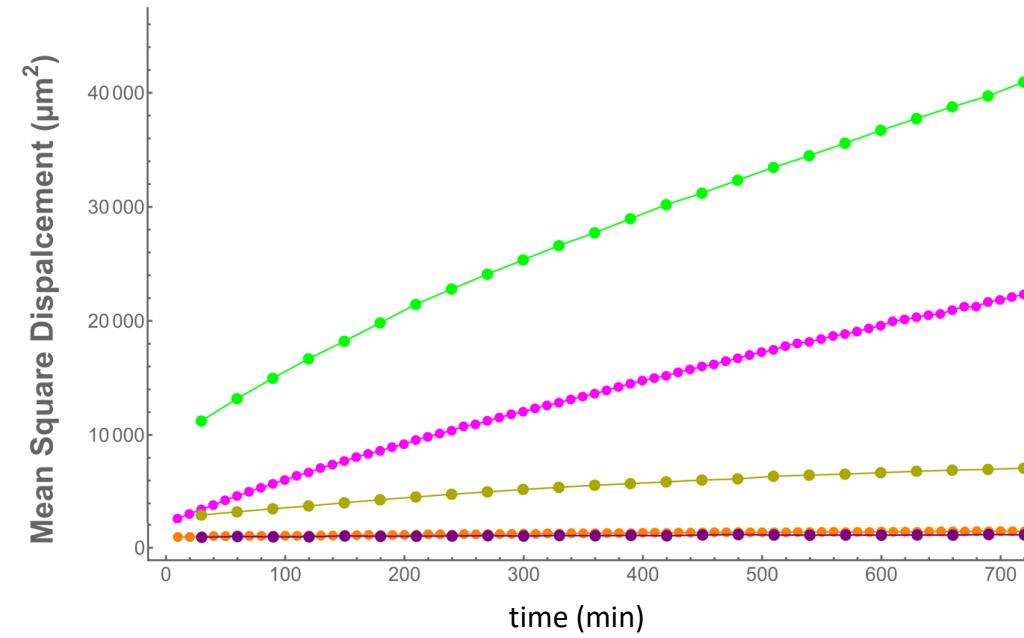
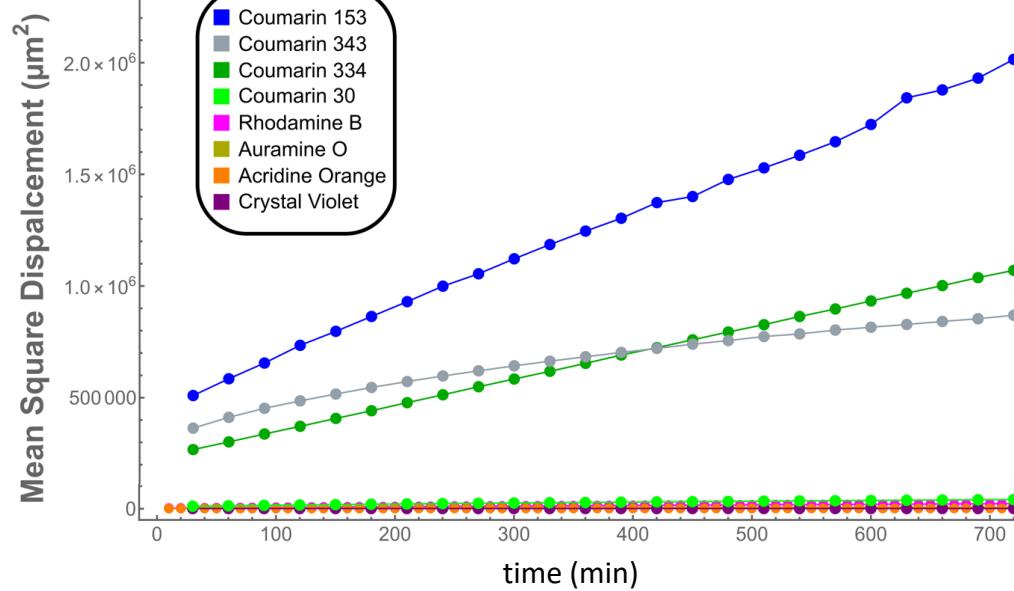
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# Summary

- Disk-soak and membrane-diffusion experiments are working well.
  - Partition and diffusion coefficients ( $k$  and  $D_p$ ) can be extracted by fitting data to numerical solutions of PDEs.
  - Diffusion coefficients are confirmed by direct observation of dyes.
- PDMS-interaction properties show the same trends noted in Auner et al: hydrophobicity is important, and yet some chemicals with very high LogP have surprisingly weak interactions – **be wary of simple read-across methods!**
- Multiphysics models based on measured  $k$  and  $D_p$  values do a good job of predicting chemical distributions under flow within a microfluidic device – **measure and model is a viable strategy!**
- In terms of mitigation, substituting SEBS co-polymer reduces but does not eliminate diffusion through the polymer. Including a carrier protein like BSA in the culture medium can reduce chemical partitioning into PDMS.

# EXTRAS – ANOMALOUS DIFFUSION



Chemical name	$\log D_P$	$\text{AIC}_P$	$\log D_\alpha$	$\alpha$	$\text{AIC}_\alpha$
Coumarin 153	-1.192	130.7	-1.124	0.975	127.6
Coumarin 334	-1.461	68.5	-1.505	1.017	48.5
Coumarin 343	-1.562	198.5	-0.625	0.651	113.1
Coumarin 30	-2.838	106.6	-2.288	0.795	20.8
Rhodamine B	-3.042	392.2	-2.621	0.841	133.2
Auramine O	-3.683	61.6	-3.070	0.772	24.3
Acridine Orange	-4.536	215.2	-3.359	0.562	7.4
Crystal Violet	-4.912	5.5	-3.890	0.540	-19.9

$D_P$  in  $\text{mm}^2/\text{hr}$ ;  $D_\alpha$  in  $\text{mm}^2/\text{hr}^\alpha$

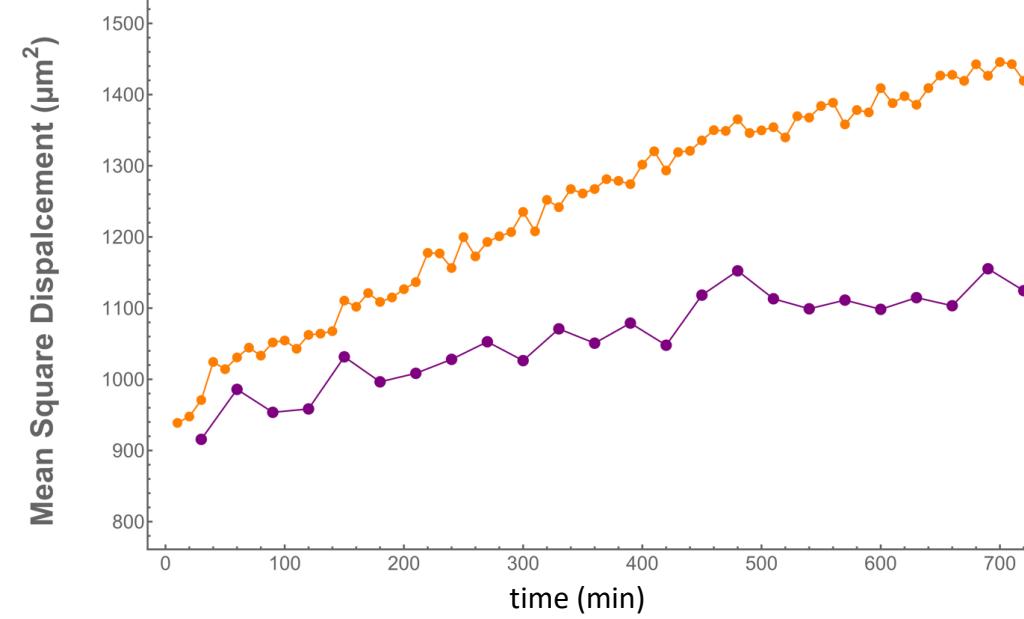


Table 1: Fluorescent dyes and selected properties. (\*) Calculated using RDKit [13]. All other properties sourced from PubChem [14]

Chemical name	logP	Mass (amu)	H-Bond Donors	H-Bond Acceptors	TPSA (Å <sup>2</sup> )
Rhodamine 6G	6.4	479.0	2	5	61.5
X-34	5.9	402.4	4	6	115
Eosin B Diphenol	5.6	580.1	2	9	168
Fluorescein-5-isothiocyanate	4.8	389.4	2	7	120
Coumarin 30	4.0	347.4	0	4	47.4
Auramine O	3.7*	303.8	2	3	30.3
Fluorescein	3.4	332.3	2	5	76
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Dyes without a color-code did not diffuse into PDMS (but some bound to surface)

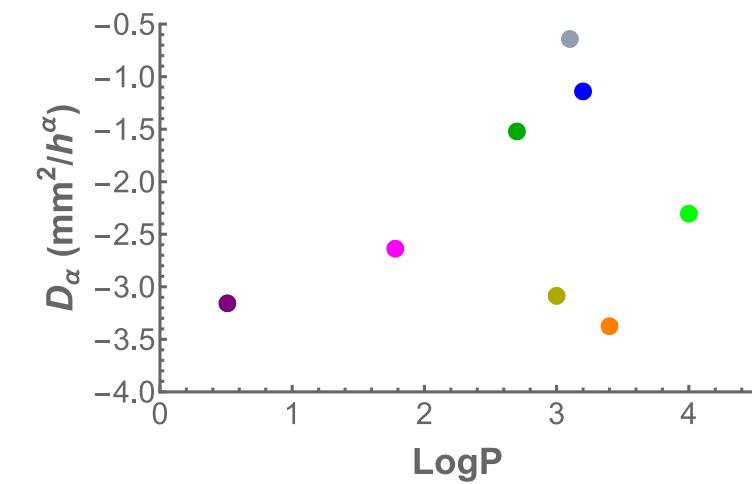
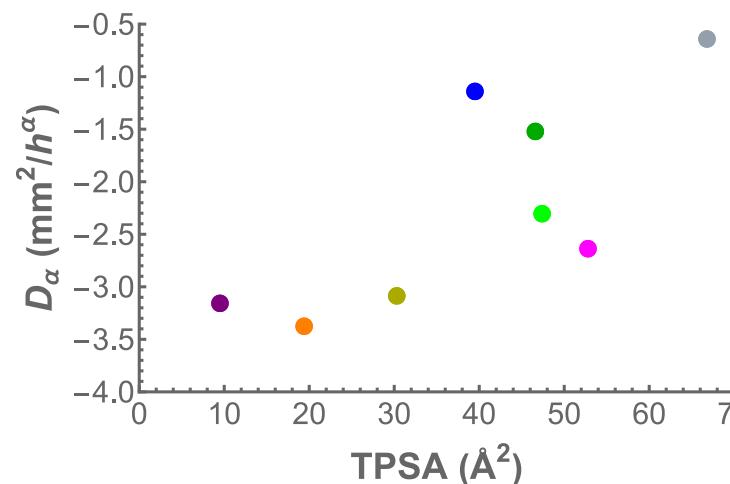
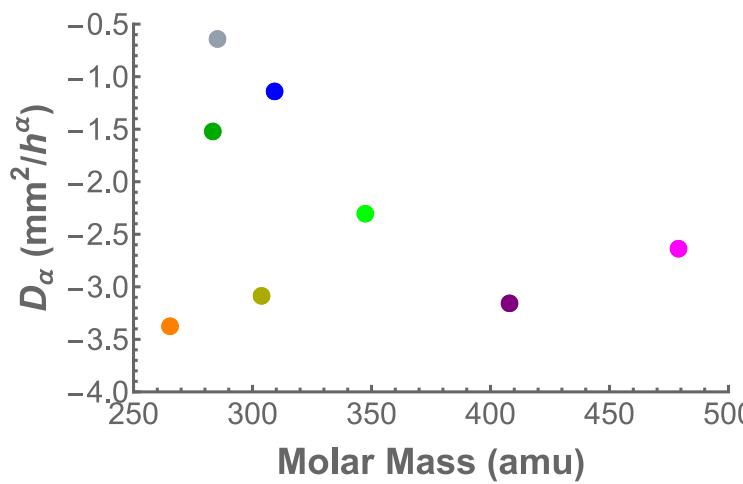


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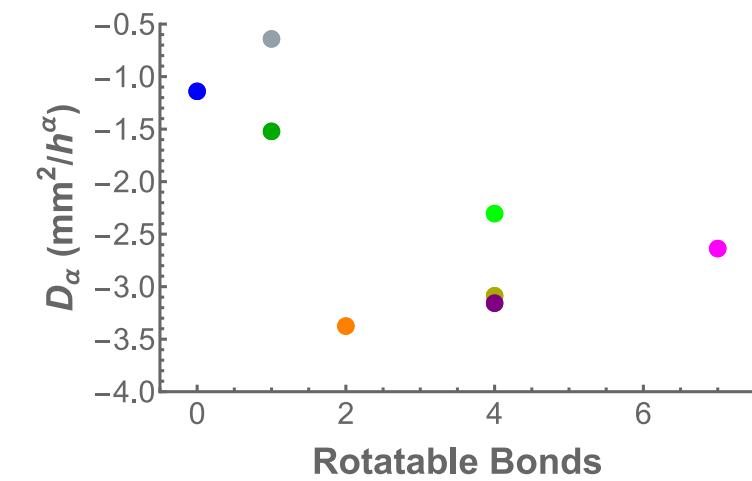
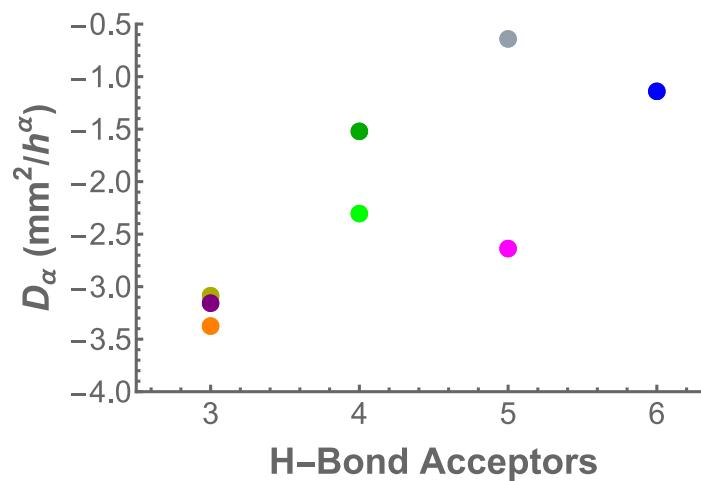
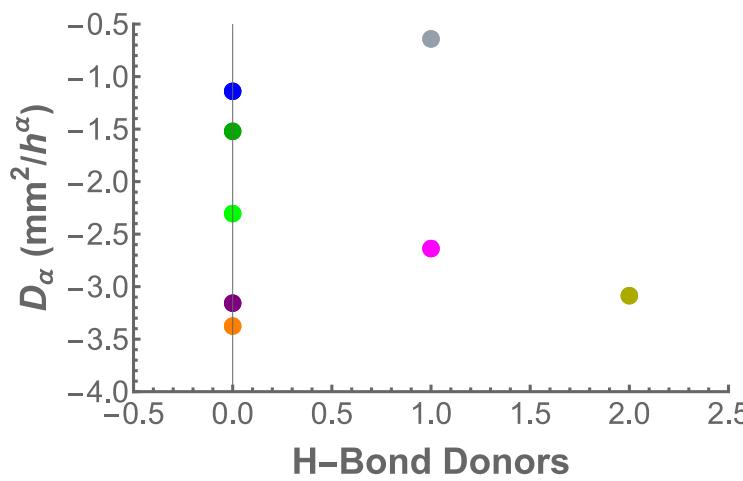


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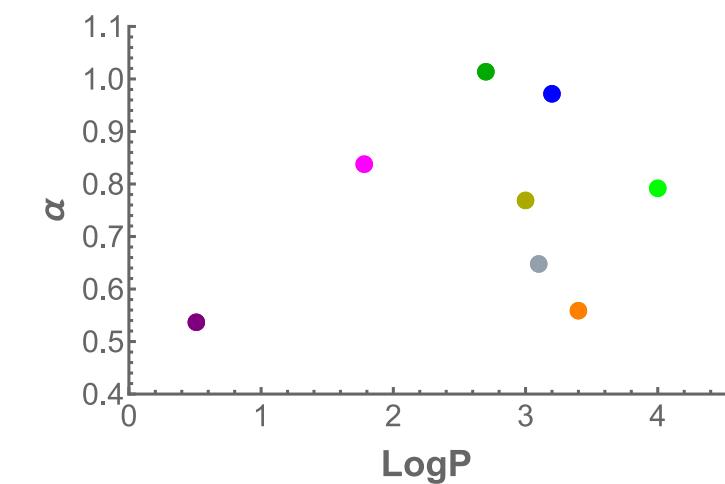
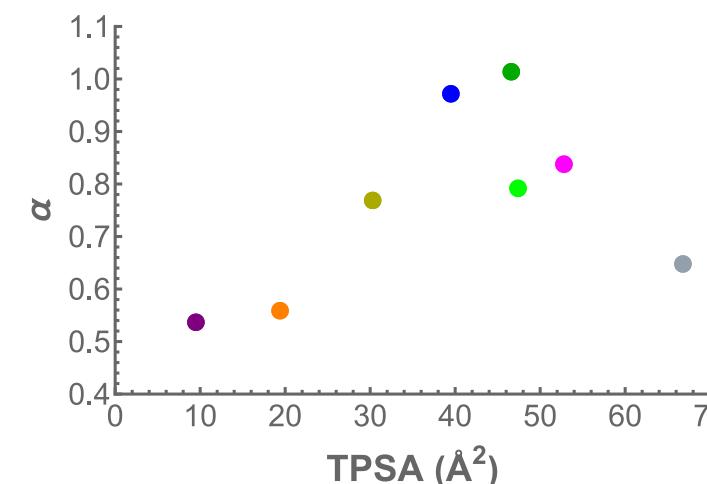
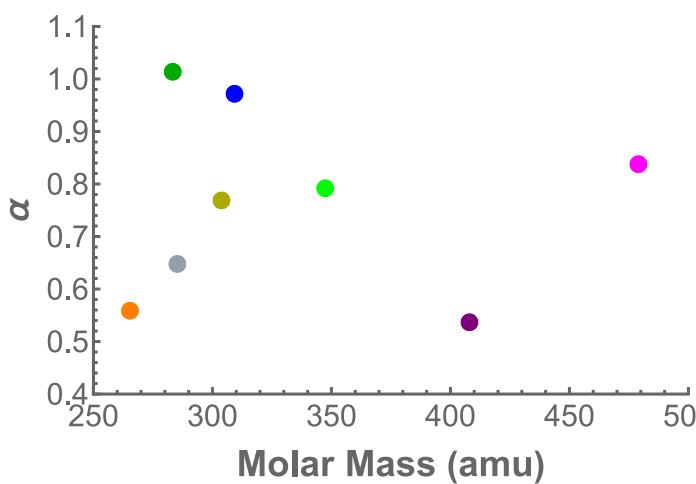
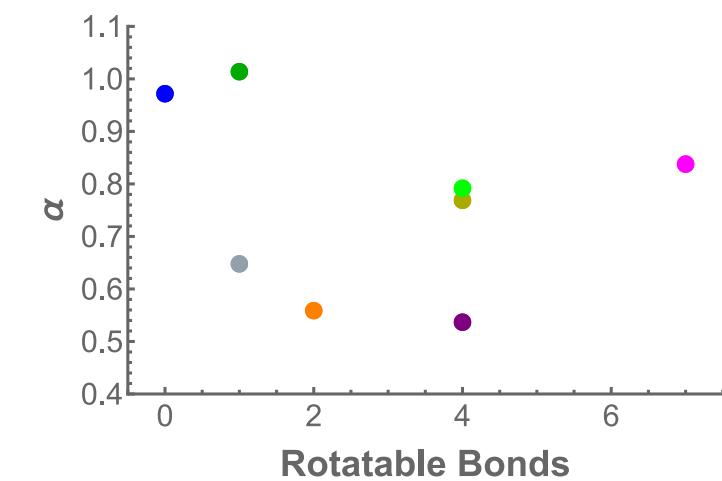
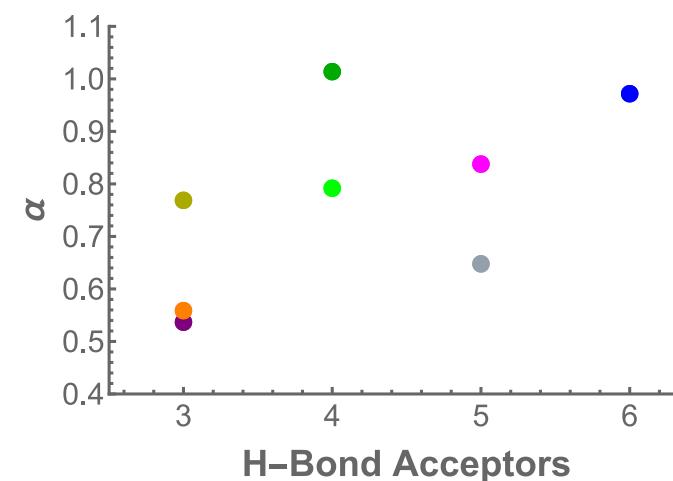
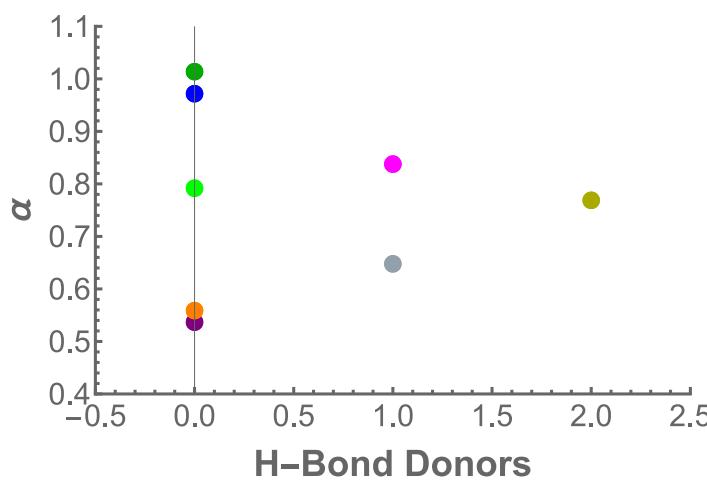


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Auramine O	3.7*	303.8	2	3	30.3
Fluorescein	3.4	332.3	2	5	76
Acridine Orange	3.4	265.35	0	3	19.4
Coumarin 153	3.2	309.28	0	6	29.5
Coumarin 343	3.1	285.29	1	5	66.8
Coumarin 334	2.7	283.32	0	4	46.6
Rhodamine B	1.9	479.0	1	5	52.8
Crystal Violet	1.5*	408.0	0	3	9.5
Merocyanine 540	0.7*	569.7	0	8	151

Dyes without a color-code did not diffuse into PDMS (but some bound to surface)





# EXISTING OECD GUIDANCE ON TOXICOKINETICS, IN VITRO- TO-IN VIVO EXTRAPOLATION, AND EXPLORING FUTURE OPPORTUNITIES

**Dr. Alicia Paini**

Senior Scientific Officer

Scientific Committee (SC)

Methodology and Scientific Support (MESE)



U.S. EPA 4th NAMs Conference

# BACKGROUND

- Chemical Risk Assessment can and should be based on non-animal data!
- This implies the need to use alternatives such as in vitro and in silico methods (New approach methodologies, NAMs)!
- Especially to interpret and use in vitro toxicity data in combination with biokinetic data!
- Biokinetic (ADME) data can be generated by in silico and in vitro models!
- PBK modeling is the way to accurately integrate and use in vitro data for the design of experiments and extrapolate in vitro effect data to in vivo for safety assessment by setting Point of Departure (PoD).

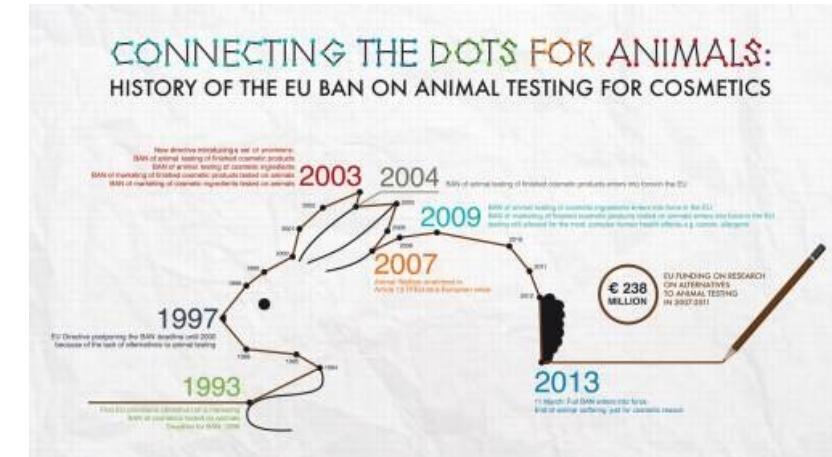
## Regulatory agencies reluctant to use mathematical models of organisms

### JRC survey highlights need for new PBK guidelines

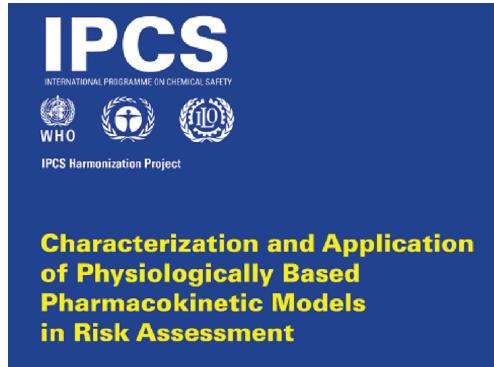
30 November 2017 / Alternative approaches to testing, Europe, United States

Regulatory agencies remain reluctant to use physiologically based kinetic (PBK) models, according to an expert survey by the European Commission's Joint Research Centre (JRC).

Widely used in industry and academia, PBK mathematical models describe how chemicals pass through the body, which is represented as a series of interconnected compartments. The models are becoming



# AN INTERNATIONAL EFFORT TO PROMOTE THE REGULATORY USE OF PHYSIOLOGICALLY BASED KINETIC (PBK) MODELS!



## Characterization and Application of Physiologically Based Pharmacokinetic Models in Risk Assessment

A screenshot of a scientific opinion document from the European Food Safety Authority (EFSA). The header includes the EFSA logo and the text 'European Food Safety Authority' and 'EFSA Journal 2014;12(3):3589'. The main title is 'SCIENTIFIC OPINION' followed by 'Scientific Opinion on good modelling practice in the context of mechanistic effect models for risk assessment of plant protection products<sup>1</sup>'. Below this, it says 'EFSA Panel on Plant Protection Products and their Residues (PPR)<sup>2,3</sup>' and 'European Food Safety Authority (EFSA), Parma, Italy'.

1. [View document](#)

2. [View document](#)

3. [View document](#)

U.S. Environmental Protection Agency (2006)

## Approaches for the Application of Physiologically Based Pharmacokinetic (PBPK) Models and Supporting Data in Risk Assessment

U.S. Food & Drug Administration (2018)

## Physiologically Based Pharmacokinetic Analyses — Format and Content Guidance for Industry

A screenshot of a guideline document from the European Medicines Agency (EMA). The header includes the EMA logo and the text '13 December 2018 EMA/CHMP/458101/2016 Committee for Medicinal Products for Human Use (CHMP)'. The main title is 'Guideline on the reporting of physiologically based pharmacokinetic (PBPK) modelling and simulation'.

1. [View document](#)

## Test No. 417: Toxicokinetics

This Test Guideline describes *in vivo* studies that provide information on mass balance, absorption, bioavailability, tissue distribution, metabolism, excretion, and basic toxicokinetic parameters [e.g. AUC], as well as supplemental approaches that may provide useful information on toxicokinetics. Information from toxicokinetic studies helps to relate concentration or dose to the observed toxicity and to understand its mechanism of toxicity. The test substance ('unlabelled' or 'radiolabelled' forms) is normally administered by an oral route, but other routes of administration may be applicable. Single dose administration of the substance (preferably a minimum of two dose levels) may be adequate, but repeated dose may be needed in some circumstances. Toxicokinetic studies should preferably be carried out in the same species as that used in other toxicological studies performed with the substance (normally the rat, a minimum of 4 animals of one sex for each dose). Initial estimation of absorption can be achieved by mass balance determination, but further investigations such as intravenous (IV) administration and biliary excretion studies might be

OECD Template #56: Biotransformation and kinetics (Version [9.6]-[August 2024])

Template #56: Biotransformation and kinetics (Version [9.6]-[August 2024])

The following table gives a detailed description of the type of information prompted for by the data entry fields.

Line no.	Field name	Field type Display type	Picklist Freetext template	Help text	Remarks Guidance Cross-reference
1.	Administrative data	Header 1			
2.		Confidentiality			
3.	Endpoint				

Guidance document on the characterisation, validation and reporting of Physiologically Based Kinetic (PBK) models for regulatory purposes



Series on Testing and Assessment  
No. 331



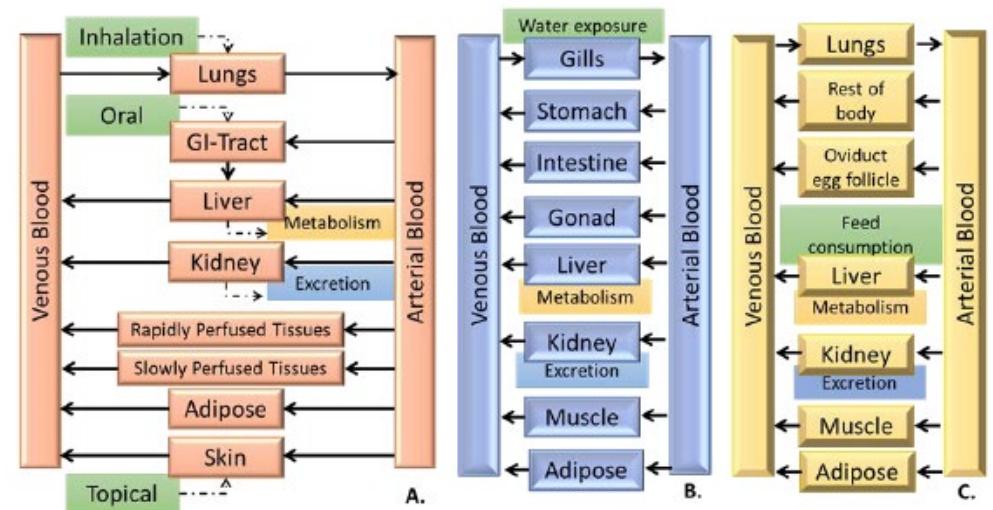
# OECD GD 331 GUIDANCE ON CHARACTERIZATION, VALIDATION AND REPORTING OF PBK MODELS FOR REGULATORY PURPOSES

- The GD provides contextual information on the scientific process of model characterization and evaluation, but not a technical guidance on model development or applications
- The GD is not prescriptive guidance on model acceptance; the level of confidence required for a model should depend on the regulatory context of use
- The GD is applicable to most chemicals and all species, provided that appropriate methods/data exist to parameterize a model
- The GD is a living document

Guidance document on the characterisation, validation and reporting of Physiologically Based Kinetic (PBK) models for regulatory purposes

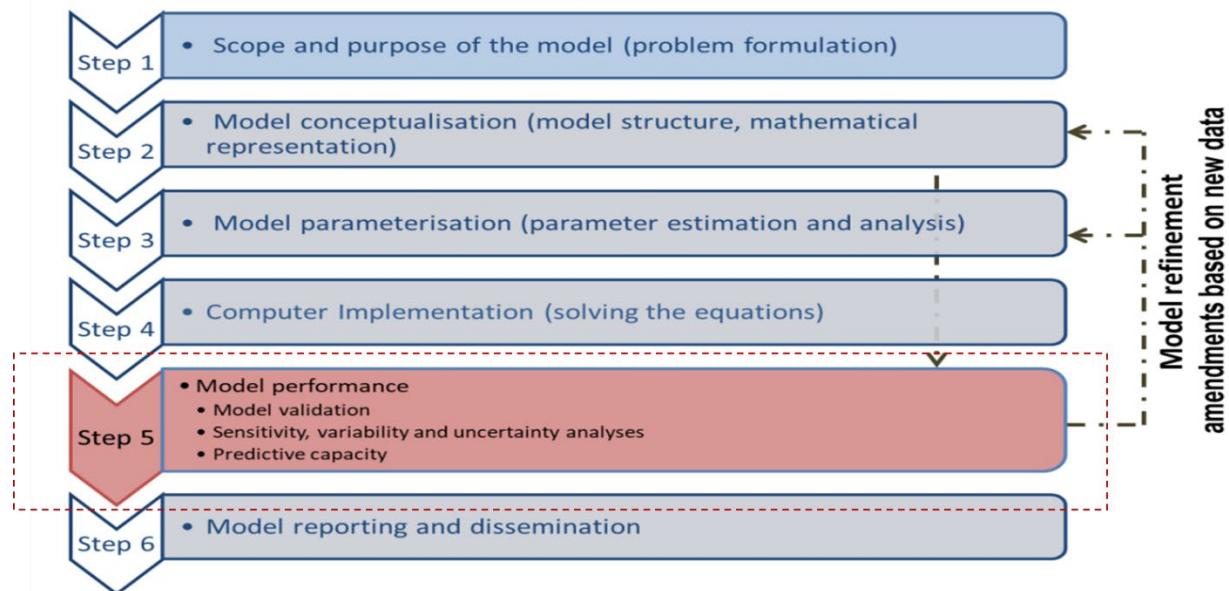


Series on Testing and Assessment  
No. 331



# GD 331 ELEMENTS

1. Provide a **scientific workflow** for characterizing and validating PBK models, with emphasis on models that are constructed without using *in vivo* data
2. Provide knowledge sources on *in vitro* and *in silico* methods that can be used to generate model parameters



# MODEL PARAMETRISATION STEP 3



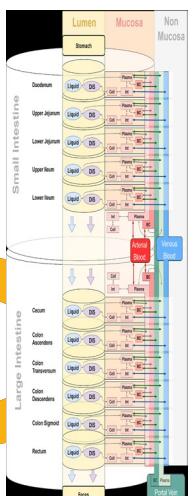
## Input parameters



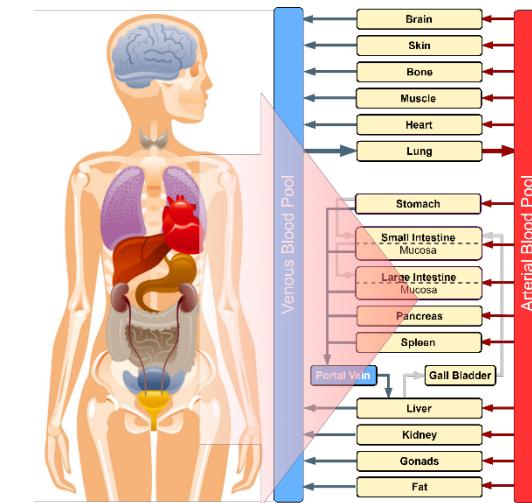
## *In vitro* ADME models



**Before using an *in vitro* parameter, one must up-scale the value - *in vitro* to *in vivo* using scaling factors.**



Absorption ( $P_{app}$ )  
Distribution ( $P_t$ ,  $f_u$ ,  $P_{app}$ )  
Metabolism ( $V_{max}$ ,  $K_m$ ,  $CL_{int}$ )  
Excretion ( $P_{ba}$ ,  $K_m$ ,  $V_{max}$ )



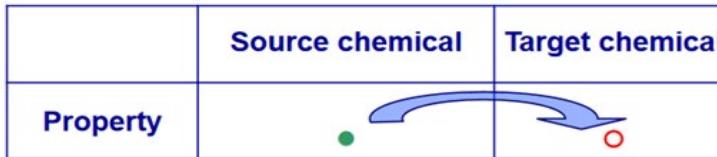
**For each of the ADME parameter, the OECD GD reports pointers for the modeler and assessor.**



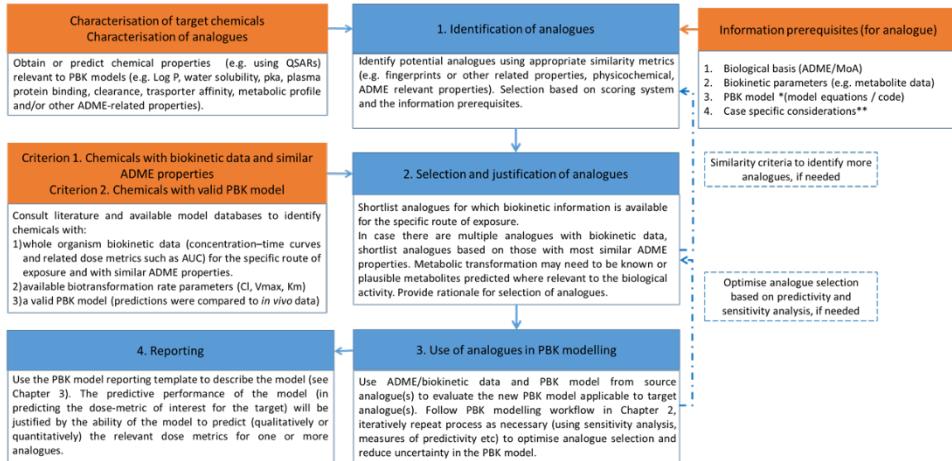
# MODEL PERFORMANCE STEP 5

1. Model Validation
2. Sensitivity, Uncertainty Analysis
3. Predictive capacity

Assessment of model predictive capacity by using a read-across approach



Identify an analogue with biokinetic data and /or has an already available PBK model to fill data gaps for a target chemical in the context of a chemical safety assessment. Problem formulation is informed by the intended application of the PBK model, including exposure scenario (see Table 2.1).



\* Ideally an existing PBK model should be available; where this is absent but there are sufficient data for the analogue, from which a PBK model could be generated, then this is a possibility

\*\* Phenotypes, population variability, lifestyle, genomics etc.



# GD 331 ELEMENTS

3. Provide an assessment framework for evaluating PBK models for intended purposes
4. Provide a template for documenting PBK models
5. Provide a checklist to support the evaluation of PBK model applicability according to context of use.

Context & Implementation



- Regulatory purpose
- Model applications
- Software implementation
- Peer input / review
- Documentation

Model validity



- *Biological basis (model structure and parameters)*
  - *Theoretical basis of model equations*
  - *Reliability and relevance of input parameters*
    - *Sensitivity of output to parameters*
    - *Goodness-of-fit and predictivity*



PBK Model Reporting Template sections	
A. Name of model	
B. Model developer and contact details	
C. Summary of model characterisation, development, validation, and regulatory applicability	
D. Model characterisation (modelling workflow)	
Step 1 – Scope and purpose of the model (problem formulation)	
Step 2 – Model conceptualisation (model structure, mathematical representation)	
Step 3. Model parameterisation (parameter estimation and analysis)	
Step 4 – Computer implementation (solving the equations)	
Step 5 – Model Performance	
Step 6 – Model Documentation	
E. Identification of uncertainties	
• model structure	
• input parameters	
• model output	
• other uncertainties	
F. Model implementation details	
• software (version no)	
• availability of code	
• software verification / qualification	
G. Peer engagement (input/review)	
H. Parameter tables	
I. References and background information	
• publications	
• links to other resources	

Part II Checklist for model evaluation

PBK Model Evaluation Checklist		Checklist assessment	Comments
Name of the PBK model (as in the reporting template)		Generic PBK model for farm animal species	
Model developer and contact details		(1) Leonie Lautz, (1) Jan Hendrik, (1) Ad Raga, (2) Jean Lou Deme 1) Radboud University, Nijmegen, Netherlands; 2) European Food Safety Authority, Parma, Italy	
Name of person reviewing and contact details		A Paini	
Date of checklist assessment		10/01/2020	
<b>A.1 Regulatory Purpose</b>			
1. What is the acceptable degree of confidence/uncertainty (e.g. high, medium or low) for the envisaged application (e.g. priority setting, screening, full assessment)?		high	
2. Is the degree of confidence/uncertainty in application of the PBK model for the envisaged purpose greater or less than that for other assessment options (e.g. reliance on PBK model and <i>in vitro</i> data vs. no experimental data)?		high	
<b>A.2 Documentation</b>			
3. Is the model documentation adequate, i.e. does it address the essential content of model reporting template, including the following:		YES	
• Clear indication of the chemical, or chemicals, to which the model is applicable?		YES	
• Is the model being applied for the same scientific purpose as it was developed, or has it been repurposed somehow?		YES	
• Model assumptions?		YES	
• Graphical representation of the proposed mode of action, if known?		NO	
• Graphical representation of the conceptual model?		YES	
• Supporting tabulation for parameters (names, meanings, values, mean and standard deviations, units and sources)?		YES	
• Relevance and reliability of model parameters?		YES	
• Uncertainty and sensitivity analysis?		YES	
• Mathematical equations?		YES	
• PBK model code?		Reported in the peer reviewed articles and EFSA	

## Checklist for Evaluation of Model Applicability



## PBK Model Reporting Template

## TAKE HOME MSG PART 1

- Models are **designed for purpose**; required confidence varies as a function of the application
- Important Aspects of PBK Model Description and Assessment
  - Quality of the source data
  - Model characterization and implementation
  - Model uncertainty and sensitivity
  - Extent of critical input/prior acceptance
- Best Practice in Developing and Assessing PBK Models to Support Regulatory Application
  - Collaboration with regulatory authorities in development
    - Design for purpose addressing aspects important for the regulatory community
  - Transparency in documentation



# OECD DNT-IVB Recommendations

Leads: EFSA and US EPA

Regulatory and technical experts contributed

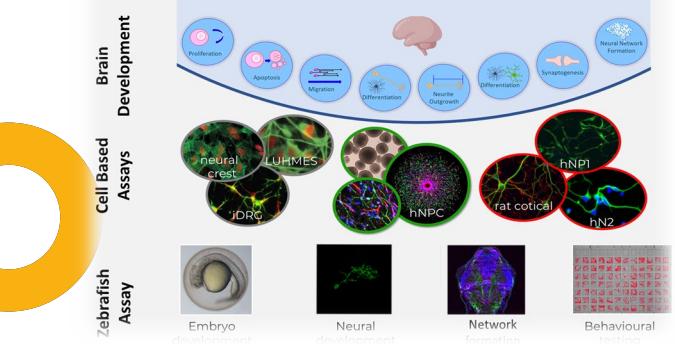
>1000  
Comments addressed



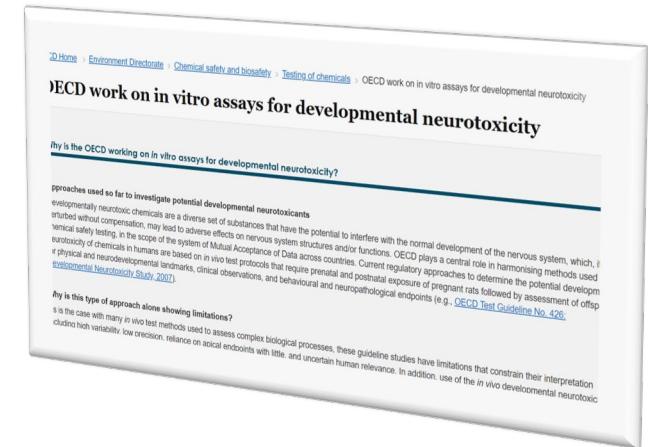
81

Compounds tested in all the assays of the DNT-IVB

## In vitro testing for DNT

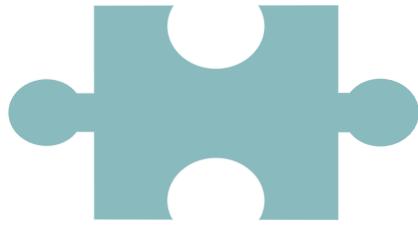


1  
OECD Document with Initial Recommendations



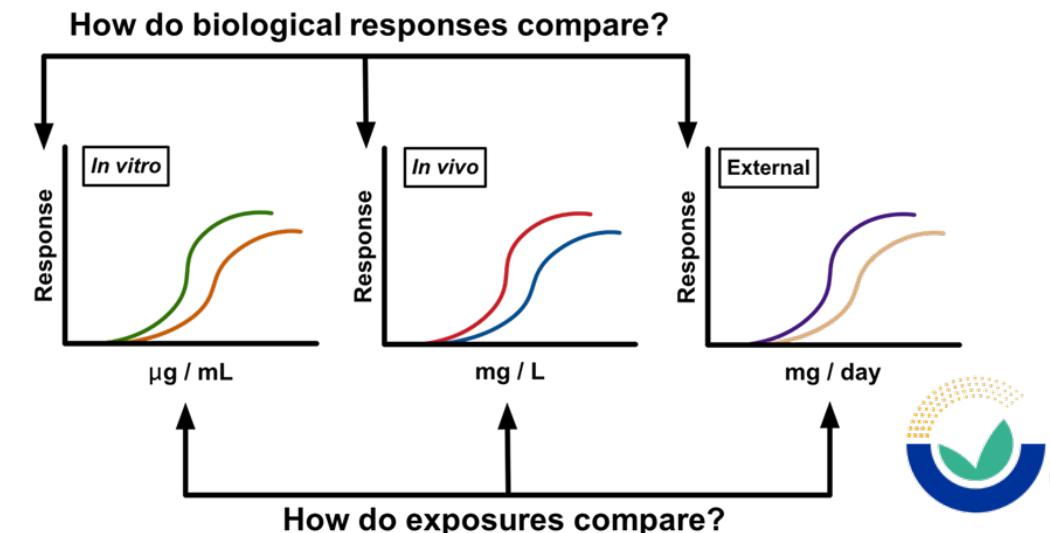
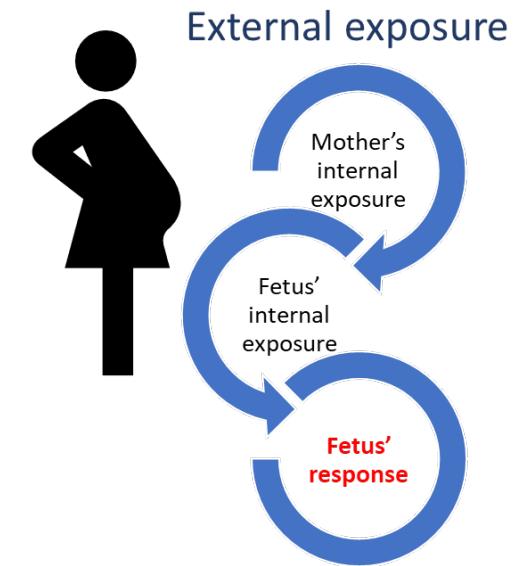
<https://www.oecd.org/env/ehs/testing/developmental-neurotoxicity.htm>





- Exposure
- Kinetics

- How can an in vitro concentration associated with bioactivity be converted to an external exposure level?
- How can a point of departure (PoD) from DNT-IVB be derived, and how can its corresponding in vivo tissue or plasma/blood concentration be determined?



# PRINCIPLES OF QUANTITATIVE IN VITRO TO IN VIVO EXTRAPOLATION (QIVIVE) APPLYING PBK MODELLING FOR DNT IVB

The **aim** of this document is to provide an overview of the principles of QIVIVE through the application of PBK modelling to facilitate the incorporation of data from the Developmental Neurotoxicity In Vitro Battery (**DNT IVB**) into chemical hazard characterization and human health risk assessments.

This document **should be read in conjunction** with the Initial Recommendations on Evaluation of Data from the Developmental Neurotoxicity (DNT) In Vitro Battery (IVB).

This **document is not an** exhaustive technical guidance for conducting QIVIVE in regulatory applications.

In Draft

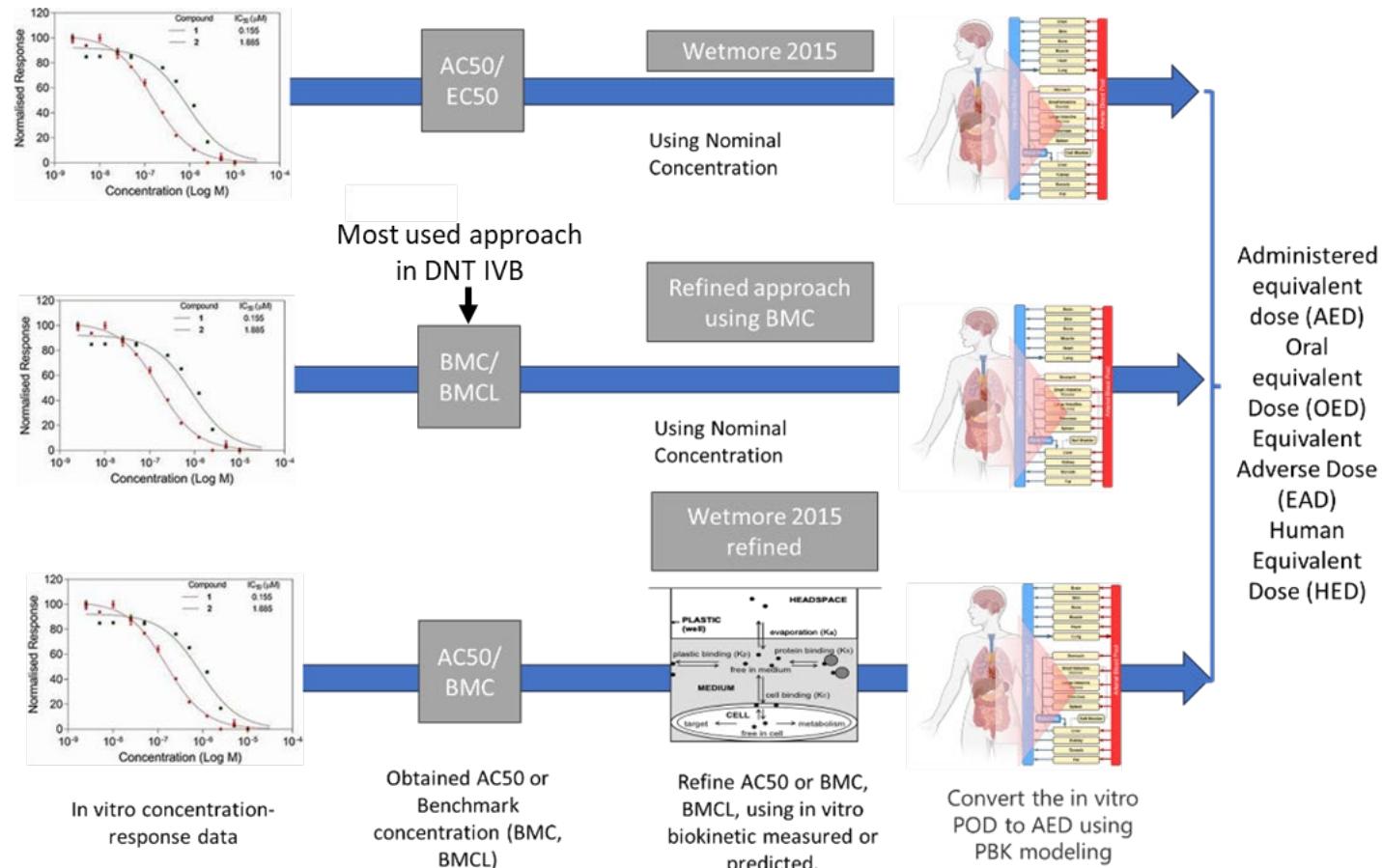


## QIVIVE Definition

The term 'QIVIVE' describes the process of converting an *in vitro* concentration associated with a specific bioactivity to an external dose (Chang et al., 2022a).



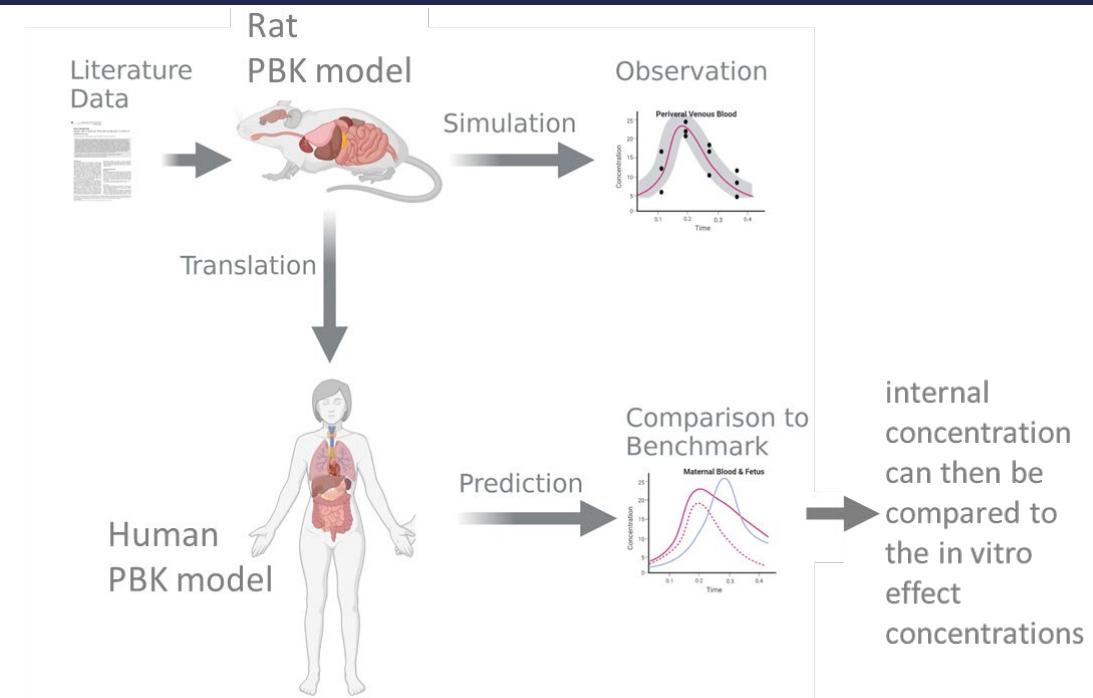
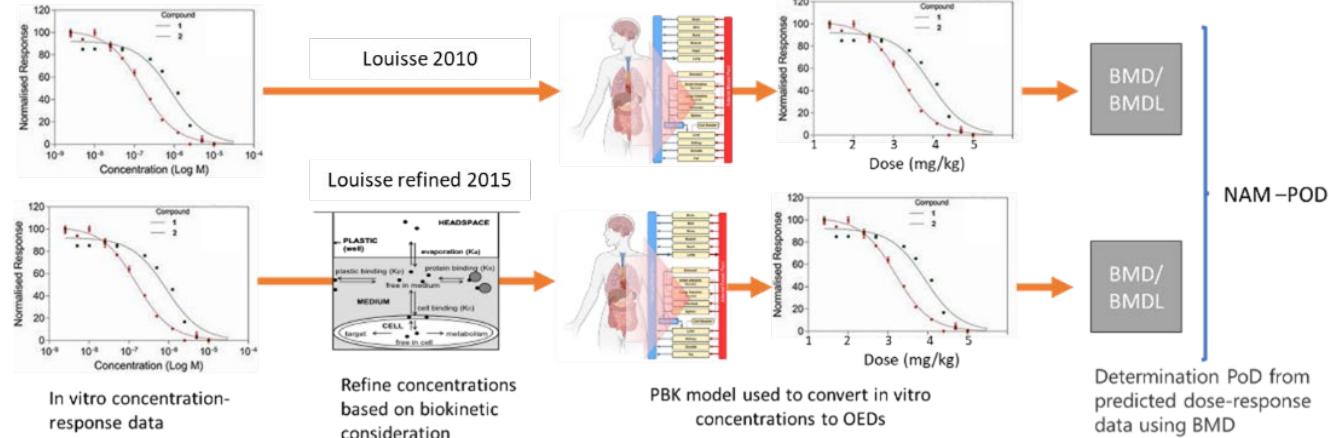
# QUANTITATIVE IN VITRO TO IN VIVO EXTRAPOLATION (QIVIVE)



Schematic presentation of a QIVIVE approach in which an in vitro POD is translated to an external dose using PBK modelling. As highlighted in the published Initial Recommendations on Evaluation of Data from the DNT IVB, to date, results from the DNT IVB are expressed as in vitro benchmark concentrations (BMC).



# QUANTITATIVE IN VITRO TO IN VIVO EXTRAPOLATION (QIVIVE)



Schematic presentation of a QIVIVE approach in which in vitro concentration-response data are translated into in vivo dose-response data using PBK modelling, from which an external POD can be derived.

In forward dosimetry, a PBK model is used to predict the internal concentrations resulting from a specific or a range of external doses. The predicted internal concentration can then be compared to the in vitro effect concentrations (Maass et al., 2023).



# WHAT IS NEEDED – PART I

- BMC or in vitro concentration response curve
- In vitro nominal to free concentration

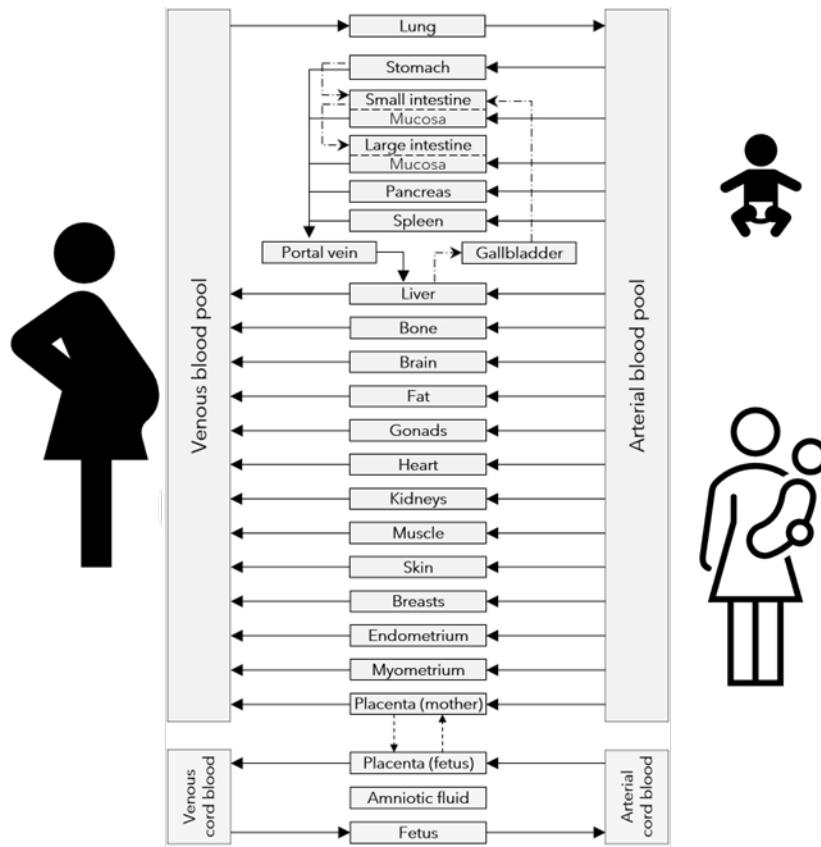
## Assays Currently in the DNT IVB mapped with information relevant for the QIVIVE approach.

Test Method (Assay)	Test System (Cell culture)	Assay Duration/ Chemical exposure	DNT Endpoint	Viability/Cytotoxicity Endpoint	Single/repeated exposure	Experimental set up (Medium composition, e.g., % FBS, lipid %, protein concentration) for in vitro distribution models	Experimental set up (Medium volume/plate format (12, 24, 48, 96)/plate material For in vitro distribution models	Is the plate coated?	Life stage - developmental period (prenatal, postnatal, 1 <sup>st</sup> , 2 <sup>nd</sup> , or 3 <sup>rd</sup> trimester)
<b>Proliferation</b>									
NPC1	human NPC grown as proliferating 3D neurospheres	72 h / 72 h	neurosphere area, BrdU incorporation in dividing cells	Resazurin reduction /LDH release	single exposure over 3 days	DMEM (#31966-021, Thermo Fisher, United States) and Hams F12 (#31765-027, Thermo Fisher, United States) in a) .....	100 µl/ 96-well plate / PS (Polystyrene), #351177 (Falcon)	Yes – poly-(2-hydroxyethyl methacrylate)	Prenatal, gestational week (GW) 16-18

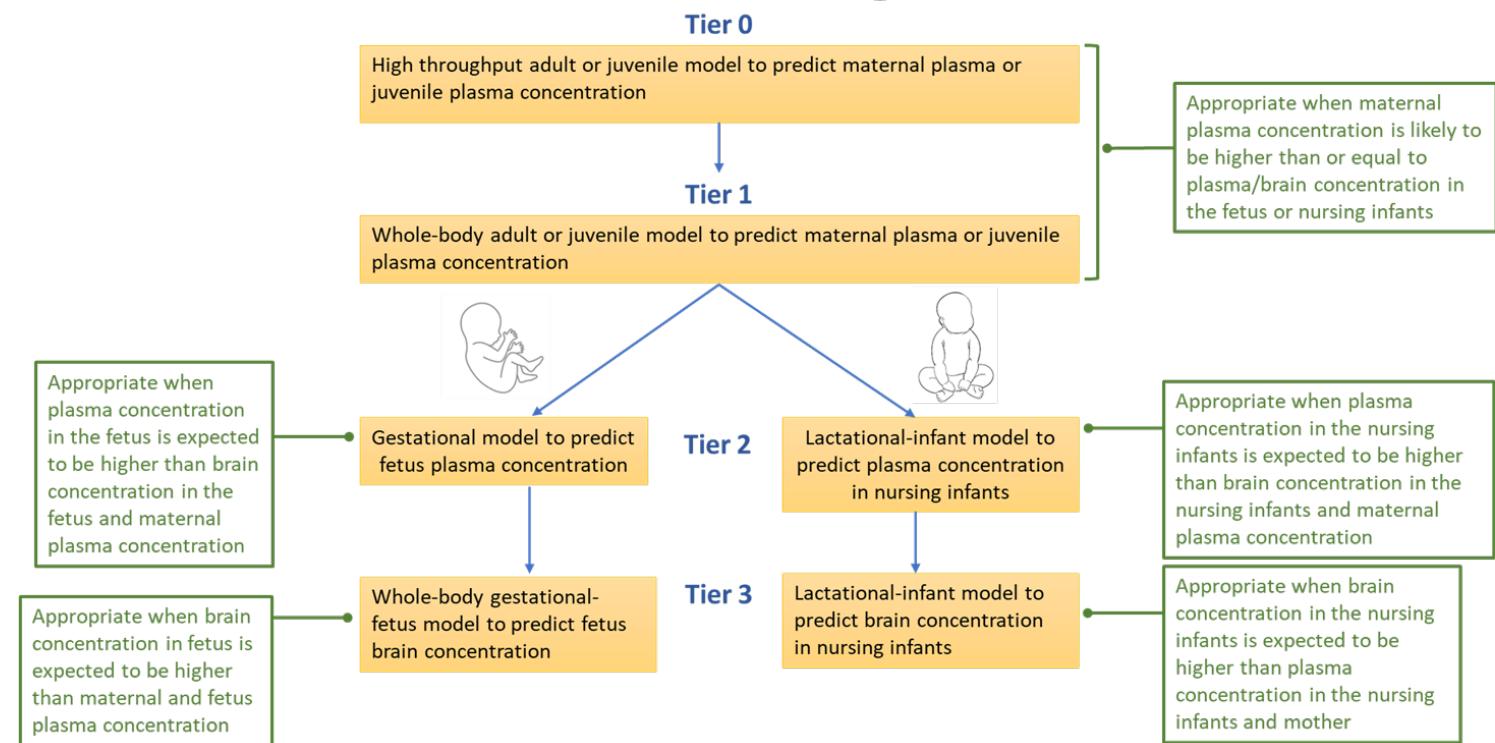


# WHAT IS NEEDED – PART II

- PBK model structure



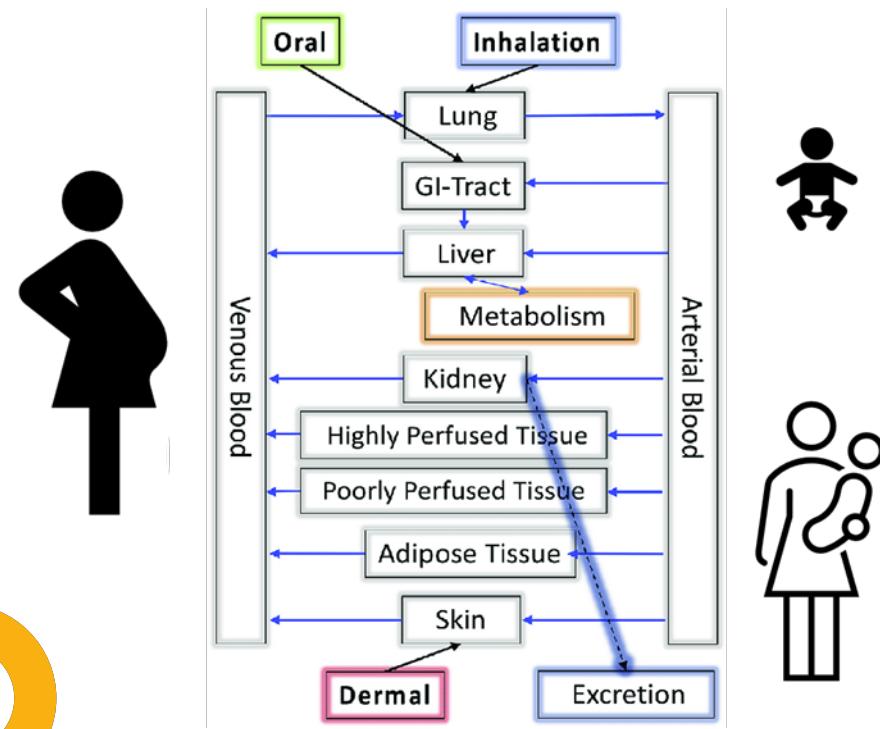
## Tiered PBK modelling framework



# WHAT IS NEEDED – PART II

- PBK model structure
- Input data for the model

- Physiology – Gestational, I, II, III or Infant + lactation.
- Biochemical – Fub, permeability, BBB, placenta, clearance., metabolism
- Physico chemical – Log Kow, Pka and MW ...

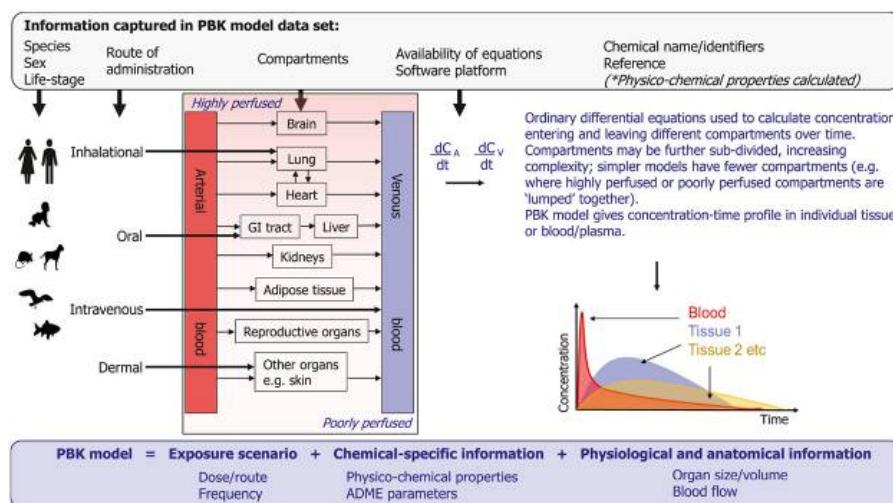


The essential set of parameters required for a model should adequately depict the physiology of the target species and the ADME of a chemical for the intended purpose, such as absorption rates, **plasma protein binding, and clearance**. Detailed recommendations for parameterizing a PBK model with in vitro and in silico approaches (OECD 331, 2021), or with in vivo data, such as those outlined in OECD TG417 (2010), can be found in published guidance documents (EPA 2006, WHO 2010).



# DATABASE OF AVAILABLE P & L PBK MODELS

Building on the  
Thompson PBK Database



Extracted in excel:

- Information on the paper
- Information on the chemical
- Chemicals identifiers
- Information on the PBK model (Pregnancy/Lactation)
- Information on PBK model parametrisation
- Information on how the model was validated



Dr. Pavani Gonnabathula

Thompson CV, Firman JW, [...] Madden JC. A Systematic Review of Published Physiologically-based Kinetic Models and an Assessment of their Chemical Space Coverage. Altern Lab Anim. 2021 Sep;49(5):197-208. doi: 10.1177/02611929211060264.

Job advertisement for a Post Doctoral Research Assistant 'In Silico Methods in PBK' - Fixed term at Liverpool John Moores University. The job is located in the School/Department of Pharmacy and Biomolecular Sciences, with a fixed-term contract. The job type is Academic, Research, and the salary range is £31,387 - £36,924 per annum. The position is full-time, with a closing date of 17/11/2024. Ref No: 4890. The document is a PDF of 268.07kb. The advertisement includes links for 'View this job description' and 'Send to a friend'.

17/11/2024



<https://jobs.ljmu.ac.uk/vacancies/9797-491997-Research%20jobs.html>

## TAKE HOME MSG PART II

- Provide principles and information to perform QIVIVE for OECD DNT in vitro test battery!
- QIVIVE approaches are presented!
- PBK model framework is proposed!
- PBK model availability is provided!

The document should  
be ready by end of the year

Overall, there is a need for a general QIVIVE guidance!!!



# WHAT NEXT?

# STILL TO BE ACCEPTED BY OECD WP!!!

## Working Party on Hazard Assessment (WPHA)

Project Title

Update of the OECD 331 Guidance document on the characterisation, validation and reporting of Physiologically Based Kinetic (PBK) models for regulatory purposes

Submitted by:

USEPA/EFSA

Date of Submission to the Secretariat:

December 1, 2024

Details of Lead Country(ies)/Organisation

<u>Country /Organisation:</u>	European Commission United States
<u>Agency/ministry/Other:</u>	European Food Safety Authority US Environmental Protection Agency
<u>Phone:</u>	Alicia Paini: +39 0521 036172 Cecilia Tan:
<u>Email:</u>	alicia.paini@efsa.europa.eu Tan.Cecilia@epa.gov

## OECD TEST GUIDELINES PROGRAMME

### Standard Project Submission Form

If you require further information please contact the OECD Secretariat  
Return completed forms to:

Anne Gourmelon (anne.gourmelon@oecd.org)  
and Lesley Smith (Lesley.smith@oecd.org)

PROJECT TITLE

Development of Test Guidelines for Measuring Human Hepatic Clearance and Plasma Protein Binding Using In Vitro Methods and Integration in a Defined Approach for Toxicokinetic Characteristics

SUBMITTED BY (Country / European Commission / Secretariat)

European Commission and United States

DATE OF SUBMISSION TO THE SECRETARIAT

24 October 2024

Working Group of National Co-ordinators of the TGs programme (WNT)



# WHAT NEXT? @EFSA PPR WG QIVIVE 4 DNT IVB

PESTICIDES PEER REVIEW - OTHER AREAS

**Art 29 - Scientific opinion**

EFSA-Q-2024-00299 | Status: Ongoing Risk Assessment

Last updated: 14/05/2024

**Subject**  
Self-task mandate of the Plant Protection Products and their Residues (PPR) Panel for a Scientific Opinion on the application of physiologically based kinetic (PBK) modelling for the quantitative in vitro to in vivo extrapolation (QIVIVE) of developmental neurotoxicity in vitro battery (DNT IVB) data for pesticide active substances

**Output**  
No Output has been formed yet for this question.

**Supporting documents**

Document Type	Action
Acceptance of the Mandate	<a href="#">Download file (974.8KB)</a>
Mandate	<a href="#">Download file (280.6KB)</a>

**Timeline**

**General Info**

Dossier number: Not applicable

Applicants:

Mandate number: M-2023-00164

Question number: EFSA-Q-2024-00299

Question type: Art 29 - Scientific opinion

Regulation: Regulation (EC) No 178/2002

**Self-task mandate of the Plant Protection Products and their Residues (PPR) Panel for a Scientific Opinion on the application of physiologically based kinetic (PBK) modelling for the quantitative in vitro to in vivo extrapolation (QIVIVE) of developmental neurotoxicity in vitro battery (DNT IVB) data for pesticide active substances.**

<https://open.efsa.europa.eu/questions/EFSA-Q-2024-00299>

**Kick – off July 2024**



## ACKNOWLEDGEMENTS

Cecilia Tan (US EPA, US)

Magda Sachana (OECD, F)

Jochem Louisse (EFSA, IT)

OECD GD 331 drafting Team !!!

OECD QIVIVE4DNTIVB drafting Team !!!





THANK YOU!

